

**WestminsterResearch**

<http://www.westminster.ac.uk/westminsterresearch>

**Potential use of electronic noses, electronic tongues and biosensors, as multisensor systems for spoilage examination in foods**

**Ghasemi-Varnamkhasti, M., Apetrei, C., Lozano, J. and Anyogu, A.**

NOTICE: this is the authors' version of a work that was accepted for publication in Trends in Food Science and Technology. Changes resulting from the publishing process, such as peer review, editing, corrections, structural formatting, and other quality control mechanisms may not be reflected in this document. Changes may have been made to this work since it was submitted for publication. A definitive version was subsequently published in Trends in Food Science and Technology, DOI: 10.1016/j.tifs.2018.07.018, 2018.

The final definitive version in Trends in Food Science and Technology is available online at:

<https://dx.doi.org/10.1016/j.tifs.2018.07.018>

© 2018. This manuscript version is made available under the CC-BY-NC-ND 4.0 license <https://creativecommons.org/licenses/by-nc-nd/4.0/>

---

The WestminsterResearch online digital archive at the University of Westminster aims to make the research output of the University available to a wider audience. Copyright and Moral Rights remain with the authors and/or copyright owners.

---

Whilst further distribution of specific materials from within this archive is forbidden, you may freely distribute the URL of WestminsterResearch: (<http://westminsterresearch.wmin.ac.uk/>).

In case of abuse or copyright appearing without permission e-mail [repository@westminster.ac.uk](mailto:repository@westminster.ac.uk)

# Accepted Manuscript

Potential use of electronic noses, electronic tongues and biosensors as multisensor systems for spoilage examination in foods

Mahdi Ghasemi-Varnamkhasti, Constantin Apetrei, Jesus Lozano, Amarachukwu Anyogu



PII: S0924-2244(17)30557-5

DOI: [10.1016/j.tifs.2018.07.018](https://doi.org/10.1016/j.tifs.2018.07.018)

Reference: TIFS 2279

To appear in: *Trends in Food Science & Technology*

Received Date: 29 August 2017

Revised Date: 2 March 2018

Accepted Date: 11 July 2018

Please cite this article as: Ghasemi-Varnamkhasti, M., Apetrei, C., Lozano, J., Anyogu, A., Potential use of electronic noses, electronic tongues and biosensors as multisensor systems for spoilage examination in foods, *Trends in Food Science & Technology* (2018), doi: 10.1016/j.tifs.2018.07.018.

This is a PDF file of an unedited manuscript that has been accepted for publication. As a service to our customers we are providing this early version of the manuscript. The manuscript will undergo copyediting, typesetting, and review of the resulting proof before it is published in its final form. Please note that during the production process errors may be discovered which could affect the content, and all legal disclaimers that apply to the journal pertain.

1 **Potential use of electronic noses, electronic tongues and biosensors as**  
2 **multisensor systems for spoilage examination in foods**

3 Mahdi Ghasemi-Varnamkhasti <sup>\*1</sup>, Constantin Apetrei <sup>2</sup>, Jesus Lozano <sup>3</sup>,  
4 Amarachukwu Anyogu <sup>4</sup>

5  
6 *1, Department of Mechanical Engineering of Biosystems, Shahrekord University, Shahrekord,*  
7 *Iran*

8 *2, Department of Chemistry, Physics and Environment, Faculty of Sciences and Environment,*  
9 *“Dunarea de Jos” University of Galati, Romania.*

10 *3, Department of Electrical Engineering, Electronics and Automation, Industrial Engineering*  
11 *School, University of Extremadura, Badajoz, Spain*

12 *4, Department of Life Sciences, Faculty of Science and Technology, University of*  
13 *Westminster, 115 New Cavendish Street, London W1W 6UW, United Kingdom*

14  
15 \*Corresponding author: [ghasemymahdi@gmail.com](mailto:ghasemymahdi@gmail.com) , Tel/fax: +98-3832324428

16

17

18

19

20

21

22 **Abstract**

23 Development and use of reliable and precise detecting systems in the food supply  
24 chain must be taken into account to ensure the maximum level of food safety and  
25 quality for consumers. Spoilage is a challenging concern in food safety considerations  
26 as it is a threat to public health and is seriously considered in food hygiene issues  
27 accordingly. Although some procedures and detection methods are already available  
28 for the determination of spoilage in food products, these traditional methods have  
29 some limitations and drawbacks as they are time-consuming, labour intensive and  
30 relatively expensive. Therefore, there is an urgent need for the development of rapid,  
31 reliable, precise and non-expensive systems to be used in the food supply and  
32 production chain as monitoring devices to detect metabolic alterations in foodstuff.  
33 Attention to instrumental detection systems such as electronic noses, electronic  
34 tongues and biosensors coupled with chemometric approaches has greatly increased  
35 because they have been demonstrated as a promising alternative for the purpose of  
36 detecting and monitoring food spoilage. This paper mainly focuses on the recent  
37 developments and the application of such multisensor systems in the food industry.  
38 Furthermore, the most traditionally methods for food spoilage detection are  
39 introduced in this context as well. The challenges and future trends of the potential  
40 use of the systems are also discussed. Based on the published literature, encouraging  
41 reports demonstrate that such systems are indeed the most promising candidates for  
42 the detection and monitoring of spoilage microorganisms in different foodstuff.

43 **Keywords:** Spoilage; Multisensors; Electronic noses; Biosensors; Electronic tongues

44

45

## 46 1. Introduction

47 Nowadays food safety is a worldwide public health issue that considers different  
48 aspects which could promote hygiene and society health. The presence of foodborne  
49 pathogens is a major global threat to public health and is one of the substantial  
50 concerns from the production to consumption chain. Many death or illness cases  
51 associated with unsafe food as a plethora of diseases including diarrhoea, dysentery  
52 due to some food pathogens (e.g. *Salmonella* spp., *Shigella* spp., *Listeria*  
53 *monocytogenes*) being reported around the world. Furthermore, some spoilage  
54 microorganisms (e.g. *Botrytis* spp., *Pseudomonas* spp., *Acinetobacter* spp.) can  
55 significantly cause economic losses to the food manufactures by providing suitable  
56 conditions for spoiling remaining food materials (Pinu, 2016).

57 Microbiological quality and safety of foodstuff should be monitored and checked to  
58 ensure the consumption security of foods to human beings. Therefore, the originating  
59 factors and detection of spoilage in any microbiological stage across the entire food  
60 supply chain is of particular importance. The identification of microbial species in  
61 foodstuff are still routinely carried out by conventional methods such as biochemical  
62 and culturing approaches which have the disadvantages of being labour-intensive and  
63 time-consuming. Additionally, some analytical techniques enabling identification of  
64 spoilage indicators have been reported in the literature. They include purge and trap  
65 (PT), Proton transfer reaction mass Spectrometry (PTR-MS), Secondary Electrospray  
66 Ionization Mass Spectrometry (SESI-MS), Solid Phase Microextraction (SPME),  
67 Selected Ion Flow Tube Mass Spectrometry (SIFT-MS), Gas Chromatography Mass  
68 Spectrometry (GC-MS), Gas Chromatography Time of Flight Mass Spectrometry  
69 (GC-TOFMS). Apart from the fact that most of these methods require specific  
70 analytical skills and the cost of the sample preparation is relatively expensive, they are

71 also not appropriate for continuous monitoring in food industry (Ghasemi-  
72 Varnamkhasti et al., 2012). Moreover some techniques mentioned above, for instance  
73 PTR-MS, are not readily available to be used in the food industry. Hence, there is a  
74 necessity for the development and use of innovative instrumental techniques as fast,  
75 reliable, non-expensive devices for the purpose of food spoilage characterization.

76 Spoilage can occurs in either stages of slaughtering or harvesting, cleaning, blanching,  
77 processing, packaging and storage, handling and distribution (Wang, Li, Yang, Ruan,  
78 & Sun, 2016). It is worth mentioning the nature of spoilage and the constituents  
79 produced during this phenomenon are enormously complicated because the food  
80 matrix including fat, carbohydrate, and protein can support microbial growth and the  
81 exponential acceleration of spoilage. Awareness of such issues is necessary while  
82 developing and using instrumental systems. Since the changes are created either in  
83 aroma profile or food body, therefore more efficient monitoring of both mediums  
84 could result in better judgment of spoilage (Kiani, Minaei, & Ghasemi-Varnamkhasti,  
85 2016).

86 In recent decades, some diagnostic tools such as electronic noses, electronic tongues  
87 and biosensors have attracted much interest in food spoilage detection and could be  
88 considered as potential alternatives for detection of food spoilage. The development  
89 of such multisensor systems is currently an on-going activity. In recent years  
90 computerized techniques called chemometric tools have been coupled with such  
91 instruments and the capability promotion has been reported in the literature  
92 accordingly (Ghasemi-Varnamkhasti & Aghbashlo, 2014). However, the industrial  
93 use of such instruments in detecting food spoilage is still in its early stages. In  
94 particular for the case of biosensors and electronic tongue, some technical problems  
95 still need to be solved before they can be used in the food industry.

96 In this paper, different aspects of food spoilage along with conventional detection  
97 methods are reviewed. In addition, the basic principles of multisensor tools which are  
98 the candidates to be used in food detection are discussed and their applications for  
99 spoilage identification are also reviewed. New ideas for detecting instruments to  
100 monitor the food production lines are substantial needs in the food industry (Peris &  
101 Escuder-Gilabert, 2013) and as the paper presents, the use of such detection systems  
102 is the future of food spoilage evaluation domain and consequently promising future  
103 could be imagined for industrial and commercial usage of such systems in food  
104 supply chain, from production to consumption.

## 105 **2. The nature of food spoilage and factors involved in the process**

106 Food spoilage remains a global economic problem that is not yet under control. It is  
107 estimated that annually about 1.3 billion tonnes of food, amounting to 30% of global  
108 food production intended for human consumption is lost or wasted. This loss occurs at  
109 all levels of the food supply chain ‘from farm to fork’ with spoilage an important  
110 contributing factor (FAO, 2011).

111 Food spoilage describes a variety of cumulative undesirable changes in a food product  
112 that renders it unacceptable to consumers (Huis in’t Veld., 1996). Food spoilage is a  
113 complex process and loss of quality is associated with two main events; changes in  
114 the physical and chemical characteristics of the food product and the microbial  
115 activity of a wide range of microorganisms (Dalgaard et al., 2006; Ercolini et al.,  
116 2006). It should be noted that the distinction between both processes is not always  
117 clear. For instance, undesirable enzymes in milk are responsible for producing the  
118 rancidity and bitterness associated with spoilage. These enzymes can either be

119 indigenous or of microbial origin (The et al., 2004) but together catalyse the  
120 proteolytic and lipolytic reactions that lead to undesirable changes in the product.

121 Physicochemical spoilage processes are usually observed as changes in the flavour  
122 and colour of a food product and are also often interlinked. Physical treatments such  
123 as excessive heat, high hydrostatic pressure and ultrasound technologies can initiate  
124 chemical changes in food. Likewise, chemical reactions such as lipolysis and  
125 lipid/enzyme oxidation can cause colour change and increased viscosity, gelation or  
126 sedimentation (Ghanbari et al., 2013; Zhou et al., 2010).

127 Biochemical and microbial changes after harvest have a major impact on the final  
128 quality and shelf life of food products. Apart from physical and chemical damage,  
129 other changes to the sensory quality of a food product such as slime production, off-  
130 flavours, off-odours and blown pack spoilage of vacuum-packaged foods can be  
131 attributed to the metabolic activities of microorganisms (Brightwell et al., 2007;  
132 Parlapani et al., 2015; Wang et al., 2017; Yang and Bedoni, 2013).

133 A vast range of bacterial and fungal species play an important role in food spoilage  
134 therefore the microbial aspects of spoilage have been the subject of intensive research  
135 for decades. Initial studies used conventional microbiology methods for identifying  
136 microbial populations involved in food spoilage (Dainty and Mackay, 1992; Dalgaard,  
137 1995). However, the evolution of more powerful molecular tools, particularly those  
138 based on 16S rRNA bacterial species classification and culture independent  
139 techniques allow for a more accurate assessment of the overall microbial food  
140 ecosystem and in some cases a reconsideration of the diversity of food spoilage flora  
141 (Ercolini et al., 2006; Jaaskelainen et al., 2016; Jaffres et al., 2009; Sade et al., 2017).

142 An important point to note is that not all microorganisms present or growing in food  
143 product cause spoilage. Microbial species that directly contribute to food spoilage

144 have been described using terms such as ‘specific spoilage organisms (SSO) or  
145 ‘metabiotic spoilage associations’, the latter term was introduced to recognize the  
146 importance of microbial interactions in food spoilage (Jorgensen et al., 2000; Gram et  
147 al., 2002).

148 Many studies have reported on the major microbial species associated with spoilage  
149 for a wide range of food types (for reviews see Andre et al., 2017; Casaburi et al.,  
150 2015; Hungaro et al., 2016; Quigely et al., 2013) . It is generally acknowledged that  
151 every food product has a distinct microbial flora associated with it during each stage  
152 of processing and storage. The composition of this microbial community depends on  
153 the microorganisms present on the raw product as well as the conditions under which  
154 the food is processed, preserved or stored (Gram et al., 2002; Parpalani et al., 2014).

155 Many interrelated factors influence the shelf life and quality indicators of a food  
156 product. Intrinsic, processing and extrinsic factors individually or in combination  
157 determine the selection of SSOs that will dominate and cause deterioration of a  
158 specific food product (Mossel et al., 1995; Nychas et al., 2008). Intrinsic factors  
159 describe the inherent physical, chemical and structural properties of the food product  
160 such as water activity ( $a_w$ ), pH, nutrient availability and the presence of antimicrobial  
161 compounds for e.g. bacteriocins. Common characteristics of highly perishable foods  
162 such as milk, poultry, fish and meat is their high protein and moisture content,  $a_w >$   
163 0.998 and neutral to acidic pH. These conditions provide a suitable growth  
164 environment for a diverse range of bacterial and fungal species.

165 Physical or chemical preservation methods are applied during processing to inhibit the  
166 survival and growth of microorganisms. Baked products are usually poorly  
167 susceptible to microbial spoilage as the heat treatment during the baking process

168 eliminates most of the raw microbial flora. Post-processing contamination thus  
169 becomes an important contributory factor to spoilage.

170 The conditions under which food is stored markedly influences the composition of the  
171 microbial flora that will contribute to the spoilage of the food product (Doulgeracki et  
172 al., 2010). Extrinsic factors relate to the environment the food is exposed to during  
173 processing and storage. Temperature and the gaseous phase surrounding a food are  
174 the most important factors that affect microbial growth (Ercolini et al., 2008; Casaburi  
175 et al., 2015). Modifications to these conditions e.g. refrigeration, modified atmosphere  
176 or vacuum packaging can be used to delay spoilage by slowing down microbial  
177 metabolic activity.

178 As previously mentioned, SSOs typically represent a small percentage of microbial  
179 species associated with a food product. This is because antagonistic and synergistic  
180 interactions between the factors described above, referred to as implicit parameters,  
181 will select for specific specie(s) adapted to occupy these ecological niches depending  
182 on their physiology and nutrient assimilation ability (Mossel et al., 1995). Table 1  
183 summarises the influence of these factors on the microbial species associated with  
184 major food products.

185 For example, lactic acid bacteria (LAB) such as *Carnobacterium* spp. have been  
186 shown to dominate the spoilage microbiota of different meat and fish products stored  
187 at low temperature under modified atmospheres (Paludan-Muller et al., 1998; Barakat  
188 et al., 2000; Laursen et al., 2005). However, in similar products stored aerobically  
189 within the same temperature range, psychrotolerant aerobes like *Pseudomonas* spp.  
190 often dominate (Del Rio et al., 2007; Nychas et al., 2008; Paparlani and Boziaris,  
191 2016).

192 Table 1. Reports on spoilage microorganisms in selected food products as influenced by  
193 intrinsic and extrinsic factors

### 194 **3. Traditional methods and recent developments**

195 Food spoilage is of great economic significance. The ability to predict shelf-life  
196 during the development of new products and to determine remaining shelf life during  
197 storage of food products is important for all stakeholders in the food value chain. This  
198 has necessitated the development of fast, accurate and reproducible methods for  
199 monitoring food spoilage (Blixt and Borsh, 1999). Traditional methods used for  
200 quality control typically rely on microbiological, chemical and sensory analysis  
201 (Haugen et al., 2006; Gobbi et al., 2010; Spadafora et al., 2016).

202 Early studies focused on determining the microbiological status of food products  
203 relied mainly on total viable counts (TVC) and phenotyping microbial isolates using  
204 biochemical tests (Dainty and Mackey, 1992; Haugen et al., 2006). These methods are  
205 time consuming and sometimes provide limited information as the extent of spoilage  
206 does not always correspond to the number of microorganisms present in the food  
207 (Blixt and Borsh, 1999; Ramirez-Guizar et al., 2017). Furthermore, they often  
208 underestimate the true microbial community. More recently, molecular approaches  
209 based on rRNA gene sequences or metagenomics are increasingly used to identify  
210 microbial communities involved in spoilage (Jaaskelainen et al., 2016; Jaffres et al.,  
211 2009; Sade et al., 2017).

212 Chemical methods can be used as an indirect means to detect and quantify microbial  
213 contamination of food based on the analysis of certain chemical markers. The quantity  
214 of cell wall components such as chitin and ergosterol are used to assess spoilage of oil  
215 seeds during storage (Gancarz et al., 2017). The colour change associated with  
216 spoilage of chicken meat can be measured using colorimetry and spectrophotometry

217 (Mancini et al., 2005). The amounts of total volatile basic nitrogen (TVBN) and  
218 trimethylamine can be indicative of fish spoilage (Jaffres et al., 2011) but as these  
219 markers only increase in fish during the late stages of storage, they cannot be used as  
220 an indication of freshness (Oehlenschangler, 2014). Organic acid profile and pH are  
221 also routinely measured. A drawback of some of these methods is the requirement for  
222 laborious sampling and extraction procedures. Despite technological advances,  
223 sensory analysis using trained panellists remains an important aspect of investigating  
224 the direct quantification of spoilage (Parpalani et al., 2014; Lytjou et al., 2017);  
225 however this is not always practical for routine analysis as it is time consuming and  
226 requires skilled personnel.

227 Nowadays, the detection of characteristic volatile compounds (VOC) of microbial  
228 origin has become a viable option to investigate the presence and growth of spoilage  
229 organisms in food and has been used in clinical settings (Tait et al., 2014). Wang et  
230 al., (2016) recently reviewed the range of methods used for the sampling, detection  
231 and analysis of these microbial volatile organic compounds in foods.

232 Solid phase microextraction (SPME) coupled with gas chromatography/mass  
233 spectroscopy (GC/MS) is one of the most common methods for studying volatile  
234 organic compounds. The use SPME-GCMS to evaluate the degree of spoilage in  
235 several food products including yoghurt (Ndagijimana et al., 2008), shrimp (Jaffres et  
236 al., 2011), ham (Martin et al., 2010) has been reported. However, VOC profiles are  
237 influenced by sample preparation, extraction and chromatographic procedures which  
238 may create inconsistencies (Ramirez-Guizar, 2017).

239 The development of more rapid and efficient identification methods continues to be  
240 the focus of intensive research. While traditional methods are for the most part cost  
241 effective, they do not always provide accurate, sensitive and reliable information.

242 Instrumentation overcomes this hurdle but widespread routine use for quality control  
243 during processing and storage is limited by cost of equipment and technical skills  
244 required by personnel (Concina et al., 2009; Wang et al., 2016). Furthermore, they  
245 mainly focus on compounds produced when food is spoiled, limiting their use for at-  
246 site quality monitoring.

247 In recent decades, there have been developments towards the use of gas sensors in  
248 devices such as the electronic nose for odour detection and electronic tongue (Gil-  
249 Sanchez et al., 2011) and biosensors. Despite all advancements in this research area,  
250 the complexity of the microbiological and biochemical processes involved in spoilage  
251 remains a challenge to developing a single quality monitoring technique for individual  
252 food products (Remenant et al., 2015).

253

#### 254 **4. Production of chemical compounds (gas and substrate) in spoiled foods**

255 As described previously, various sensory defects such as off-odours, off-flavours and  
256 discolouration in spoiled food can be attributed to the presence and metabolic activity  
257 of spoilage microorganisms. During exponential growth, spoilage microorganisms  
258 preferentially utilize the carbohydrates, sugars, proteins and fats in food to provide  
259 their metabolic needs. For example, during storage at low temperatures, bacteria  
260 present in meat use glucose as a carbon and energy source. When glucose is depleted,  
261 other substrates such as lactate, pyruvate, amino acids and nucleic acids may be  
262 metabolized (Casaburi et al., 2015). Primary metabolites such as polysaccharides,  
263 amino acids, lipids and vitamins act as precursors for the production of a range of  
264 compounds. These chemical compounds serve as indicators of spoilage and comprise

265 of organic acids, biogenic amines and a range of VOCs (alcohols, aldehydes, ketones,  
266 esters, volatile fatty acids and sulphur compounds) (Doyle, 2007; Wang et al., 2016).

267 The composition and concentration of VOCs produced in food is for the most part  
268 determined by the combined effect of both intrinsic and extrinsic factors. For  
269 example, some amino acids can be decarboxylated by microbial enzymes to produce  
270 biogenic amines such as histamine, tyramine, putrescine and cadaverine (Naila et al.,  
271 2010). Biogenic amine accumulation in fermented meat products has been reported to  
272 be influenced by fermenting strains, pH, sausage diameter (intrinsic) as well as  
273 storage temperature and relative humidity (extrinsic). These conditions favour  
274 proteolytic and decarboxylase reactions required for biogenic amine formation  
275 (Suzzia and Gardini, 2003; Lattore-Moratalla et al., 2012).

276 A list of some compounds associated with the spoilage of selected food products is  
277 reported in Table 2. Several authors have reported the detection and measurement of  
278 these molecules in spoiled food and there have been attempts to identify VOCs that  
279 are likely specific to both SSO and substrate (Concina et al., 2009; Spadafora et al.,  
280 2016). This has paved the way for more focused studies to determine the so called  
281 chemical spoilage index (CSI), a profile of microbial VOCs (MVOCs) for a particular  
282 food product (Parpalani et al., 2014). The concentration of these CSI metabolites  
283 should increase in tandem with the growth of the SSOs as well as loss of sensory  
284 quality and therefore can be used to estimate shelf life (Jay, 1986; Miks-Krajnik et al.,  
285 2016).

286 Table 2. Some spoilage substrates and metabolites typically found in spoiled food

287 Correlating sensory impressions of spoilage to the metabolic activity of SSOs is not  
288 always clear. This reflects both the complex nature of food spoilage and the limited  
289 information available regarding the metabolism of the microbial species involved.

290 Some VOCs can be produced from reactions catalysed by both SSOs and food  
291 matrix enzymes, others from complex metabolic reactions involving different  
292 microbial species (Remenant et al., 2015). Species of LAB, *Enterobacteriaceae* and  
293 *Clostridia* have been implicated in ‘blown pack’ spoilage (BPS) of refrigerated,  
294 vacuum packed meat products (Brightwell et al., 2007; Hernandez-Macedo et al.,  
295 2012). The ‘blown pack’ effect has been attributed to gas production but it remains  
296 unclear which species is directly implicated although some authors have attributed  
297 BPS to be largely due to the metabolic activities of *Clostridium estertheticum* (Cavill  
298 et al., 2016; Rajagopal et al., 2016). In addition, MVOCs identified from culture  
299 media experiments as potential CSI candidates may not be detected in food (Yu et al.,  
300 2000).

## 301 **5. Multisensor systems**

### 302 *5. 1. Electronic nose and its performance*

303 The human nose is much more complicated than other human senses like the ear and  
304 the eye. It is still the primary ‘instrument’ to assess the smell of various products and  
305 it is currently used to identify a diverse range of food spoilage. Sensory evaluation  
306 using the human sense of smell is subjective; careful design and rigorous training of  
307 assessors allows it to become a more objective, but still expensive option.  
308 Instrumental methods, such as gas chromatography/ mass spectrometry (GC/MS), are  
309 also expensive and require trained personnel. The concept of the electronic nose has  
310 attracted attention in many branches of industry for its potential in routine odour  
311 analysis.

312 The electronic nose is an electronic system that tries to mimick the structure of the  
313 human nose, but trying to reduce its limitations. An accepted definition was given by

314 Gardner in 1994: “an electronic nose is an instrument which comprises an array of  
315 electronic chemical sensors with partial specificity and an appropriate pattern  
316 recognition system, capable of recognising simple or complex odours” (Gardner &  
317 Bartlett, 1994). The similarity of electronic nose with the biological sense of smell  
318 can be observed in the smelling process: the first step in both is the interaction  
319 between volatile compounds (usually a complex mixture) with the appropriate  
320 receptors: olfactory receptors in the biological nose and a sensor array in the case of  
321 the electronic nose. The next step is the storage of the signal generated by the  
322 receptors in the brain or in a pattern recognition database (learning stage) and later the  
323 identification of one of the odour stored (classification stage). An electronic nose uses  
324 currently a number of individual sensors (typically 5-100) whose selectivities towards  
325 different molecules overlap. The response from a chemical sensor is usually measured  
326 as the change of some physical parameter, e.g. conductivity or current. There are  
327 some significant drawbacks for these devices, like the lack of selectivity and the  
328 sensors drift, that are one of the main research topics in this field. On the other hand,  
329 they have the advantage of high portability for making in situ and on-line  
330 measurements with lower costs and good reliability.

331 An electronic nose generally consists of an aroma extraction system, a sensor array, a  
332 control and measurement system, and a pattern recognition method. A simple flow  
333 chart of the typical structure of an electronic nose is shown in Fig. 1 (Lozano, 2006).

334 Fig. 1. Block diagram of an electronic nose system.

335 The aroma extraction system or sampling method carries the volatile compounds from  
336 the samples to the sensor chamber and it significantly contributes to the capability and  
337 reliability in an odour sensing system. Various techniques of the sample flow, static

338 and preconcentrator systems are available for using with an electronic nose and the  
339 most appropriate aroma extraction system should be selected for the project taking  
340 into account the type of samples, the application and the portability of the system.

341 There is a basic classification of sampling methods if concentrator is used or not. A  
342 concentrator is often used to enhance the sensitivity and can be used to autonomously  
343 enhance the selectivity of a sensor array. On the other hand, there are two main types  
344 of aroma extracting systems, the sample flow system and the static system. In the first  
345 one, the sensors are placed in the vapour flow, which allows the rapid exchange of  
346 vapour and hence many samples can be measured within a short time. In the static  
347 system, there is no vapour flow around the sensor, and measurements are usually  
348 made on the steady-state responses of the sensors exposed to vapour at a constant  
349 concentration. The most common techniques used for solid or liquid samples in food  
350 applications are static headspace (HS), purge and trap (P&T) and solid phase micro  
351 extraction (SPME) (Lozano, Santos, Gutiérrez, & Horrillo, 2007).

352 The most important part of an electronic nose is the detection system or chemical  
353 sensors, that are capable of converting a chemical change in the environment into an  
354 electric signal in the gas sensors and respond to the concentration of specific  
355 compounds from gases or liquids (Nagle, 2006). Chemical sensors can be based on  
356 electrical, thermal, mass or optical principles. Several examples of chemical sensors  
357 used in electronic noses are: conducting polymers (Guadarrama, Fernández, Íñiguez,  
358 Souto, & De Saja, 2000), semiconductor devices (Jose Pedro Santos & Lozano, 2015)  
359 quartz resonators (Sharma et al., 2015), and surface acoustic sensor (SAW) (Jose  
360 Pedro Santos et al., 2005).

361 Conducting polymers (based on polypyrrole, polyaniline, thiophenes, indoles, or  
362 furans) have been used as the active layers of gas sensors since early 1980s. The  
363 sensors made of conducting polymers have many improved characteristics: high  
364 sensitivities and short response time at room temperature. The electronic interface is  
365 straightforward, and they are suitable for portable instruments. Conducting polymers  
366 are easy to be synthesized through chemical or electrochemical processes, and their  
367 molecular chain structure can be modified conveniently by copolymerization or  
368 structural derivations. Most of the conducting polymers are doped/undoped by redox  
369 reactions; therefore, their doping level can be altered by transferring electrons from or  
370 to the analytes. Electron transferring can cause the changes in resistance and work  
371 function of the sensing material. The work function of a conducting polymer is  
372 defined as the minimal energy needed to remove an electron from bulk to vacuum  
373 energy level. This process occurred when the sensing films are exposed to redox-  
374 active gases. They can remove electrons from the aromatic rings of conducting  
375 polymers. When this occurs at a p-type conducting polymer, the doping level as well  
376 as the electric conductance of the conducting polymer is enhanced. An opposite  
377 process will occur when detecting an electro-donating gas.

378 Semiconductor chemical sensors detect gases and aromas in samples by a chemical  
379 reaction that takes place when the gas comes in direct contact with the sensor surface.  
380 This chemical reaction and the presence of the gases can be detected since the  
381 electrical resistance in the sensor is modified when it is exposed to the monitored gas.  
382 This change in resistance is measured and can be used to identify the presence of a  
383 gas, to predict the the gas concentration or other tasks. Tin dioxide in different  
384 structures (thin or thick film, nanostructures, nanowires, etc.) is the most common  
385 material used in semiconductor sensors, that are commonly used to detect hydrogen,

386 oxygen, alcohol vapor, and harmful gases such as carbon monoxide in different  
387 applications related with environment, health, food quality, etc. Operating the device  
388 at different temperatures and varying the type and thickness of the material, the  
389 sensitivity and selectivity can be optimized.

390 The piezoelectric family of sensors has two main members: quartz crystal  
391 microbalance (QCM) and surface acoustic-wave (SAW) devices. They can measure  
392 temperature, mass changes, pressure, force, and acceleration, but in the electronic  
393 nose, they are configured as mass-change-sensing devices.

394 The QCM type consists of a resonating disk a few millimeters in diameter, with metal  
395 electrodes on each side connected to a lead wire. The device resonates at a  
396 characteristic (10 MHz to 30 MHz) frequency when excited with an oscillating signal.  
397 During manufacture, a polymer coating is applied to the disk to serve as the active  
398 sensing material. In operation, a gas sample is adsorbed at the surface of the polymer,  
399 increasing the mass of the disk-polymer device and thereby reducing the resonance  
400 frequency. The reduction is inversely proportional to odorant mass adsorbed by the  
401 polymer.

402 The SAW sensor differs from QCM in several important ways. First, the wave travels  
403 over the surface of the device, not throughout its volume. SAW sensors operate at  
404 much higher frequencies, and so can generate a larger change in frequency. A typical  
405 SAW device operates in the hundreds of megahertz, while 10 MHz is more typical for  
406 a QCM, but SAW devices can measure changes in mass to the same order of  
407 magnitude as QCMs. Even though the frequency change is larger, increased surface-  
408 to-volume ratios mean the signal-to-noise ratio is usually poorer. Hence, SAW  
409 devices can be less sensitive than QCMs in some instances.

410 With QCMs, many polymer coatings are available, and as with the other sensor types,  
411 differential measurements can eliminate common-mode effects. For example, two  
412 adjacent SAW devices on the same substrate (one with an active membrane and  
413 another without) can be operated as a differential pair to remove temperature  
414 variations and power line noise. A disadvantage of both QCM and SAW devices is  
415 more complex electronics than are needed by the conductivity sensors. Another is  
416 their need for frequency detectors, whose resonant frequencies can drift as the active  
417 membrane ages.

418 The control and measurement system includes all electronic circuits needed for the  
419 measurements of signals generated by the sensors such as interface circuits, signal  
420 conditioning and A/D converters. This sensor electronics usually amplify and  
421 condition the sensor signal. The signal must be converted into a digital format to be  
422 processed by a computer, and this is carried out by an analogue to digital converter  
423 (e.g. a 12 bit converter) followed by a multiplexer to produce a digital signal which  
424 either interfaces to a serial port on the microprocessor (e.g. RS-232, USB) or a digital  
425 bus (e.g. GPIB). The microprocessor is programmed to carry out a number of tasks,  
426 including the pre-processing of the time-dependent sensor signals to compute the  
427 input vectors  $x_j$  and classify them against known vectors stored in memory. Finally,  
428 the output of the sensor array and the odour classification can be displayed on a LCD  
429 or on a PC monitor.

430 The main goal of an electronic nose is to identify an odorant sample and perhaps to  
431 estimate its concentration. The multivariate information obtained by the sensor array  
432 can be sent to a display so a human can read that information and do an action or an  
433 analysis. Also, that information, that is an electronic fingerprint of the volatile  
434 compound measured, can be sent to a computer to perform an automated analysis and

435 emulate the human sense of smell. These automated analysis that comes from  
436 methods of statistical pattern recognition, neural arrays and chemometrics (Aguilera,  
437 Lozano, Paredes, Alvarez, & Suárez, 2012), is a key part in the development of a gas  
438 sensor array capable to detect, identify or quantify different volatile compounds  
439 responsible for food spoilage. This process may be subdivided into the following  
440 steps: preprocessing and feature extraction, dimensionality reduction, classification or  
441 prediction, and decision-making.

442 Preprocessing compensates for sensor drift, compresses the transient response of the  
443 sensor array, and reduces sample-to-sample variations. Typical techniques include:  
444 manipulation of sensor baselines; normalization of sensor response ranges for all the  
445 sensors in an array (the normalization constant may sometimes be used to estimate the  
446 odorant concentration); and compression of sensor transients. Feature extraction has  
447 two purposes: to reduce the dimensionality of the measurement space, and to extract  
448 information relevant for pattern recognition. For example, in an electronic nose with  
449 32 sensors, typically one feature is extracted from each raw response of the sensor and  
450 the measurement space has 32 dimensions.

451 A dimensionality reduction stage projects this initial feature vector onto a lower  
452 dimensional space in order to avoid problems associated with high-dimensional,  
453 sparse datasets. Maybe, some of them probably respond in a similar (but not identical)  
454 way. This means that the number of dimensions in the data set can be reduced without  
455 any loss of information. It is generally performed with linear transformations such as  
456 the classical principal component analysis (PCA) and linear discriminant analysis  
457 (LDA). The resulting low-dimensional feature vector is further used to solve a given  
458 prediction problem, generally classification, regression or clustering.

459 Classification is a general process related to categorization, the process in which ideas  
460 and objects are recognized, differentiated, and understood. In this case, the  
461 identification of an unknown sample into previously learned classes is usually  
462 performed by artificial neural networks (ANNs). An artificial neural network is an  
463 information processing system that has certain performance characteristics in  
464 common with biological neural networks. It allows the electronic nose to function in  
465 the way a brain function when it interprets responses from olfactory sensors in the  
466 human nose. During training, the ANN adapts the synaptic weights to learn the  
467 patterns of the different odorants. After training, when presented with an unidentified  
468 odorant, the ANN feeds its pattern through the different layers of neurons and assigns  
469 the class label that provides the largest response.

470 Finally, the classifier produces an estimate of the class for an unknown sample along  
471 with an estimate of the confidence placed on the class assignment. A final decision-  
472 making stage may be used if any application-specific knowledge is available, such as  
473 confidence thresholds or risk associated with different classification errors. Cross  
474 validation is usually employed and training is stopped at the point of the smallest error  
475 in the validation set to detect and avoid overtraining.

476

## 477 *5.2. Electronic tongue*

478 The analysis of the substances dissolved in liquid samples with multisensor systems  
479 was firstly developed in mid-1980s (Otto & Thomas, 1985). In the beginning of the  
480 1990s, the first taste sensor was built, based on ion-selective electrodes (Hayashi,  
481 Yamanaka, Toko, & Yamafuji, 1990; Iiyama, Miyazaki, Hayashi, Toko, Yamafuji,  
482 Ikezaki, & Sato, 1992). The sensitive membrane was made of various lipid

483 membranes immobilized onto polyvinyl chloride (Toko, 2000). Later, in 1995, the  
484 concept of electronic tongue was introduced. It was based on inorganic chalcogenide  
485 glass sensors, being used for both qualitative and quantitative determinations (Legin,  
486 Rudnitskaya, Di Natale, Mazzone, & D'Amico, 2000; Vlasov, Legin, Rudnitskaya, Di  
487 Natale, & D'Amico, 2005).

488 This concept has been developed, and in the last years the bioelectronic tongue system  
489 was introduced (del Valle, Cetó, & Gutierrez-Capitán, 2014; Ghasemi-Varnamkhasti,  
490 Rodríguez-Méndez, Mohtasebi, Apetrei, Lozano, Ahmadi, Razavi, de Saja, 2012). It  
491 contains an array of biosensors and is able to qualitatively and quantitatively  
492 characterize multicomponent liquid samples (Cetó, Voelcker, & Prieto-Simón, 2016;  
493 Song, Jin, Ahn, Kim, Lee, Kim, Simons, Hong, & Park, 2014; Rodriguez-Méndez,  
494 Medina-Plaza, García-Hernández, de Saja, Fernández-Escudero, Barajas-Tola, &  
495 Medrano, 2014).

496 Conceptually speaking, electronic tongues are analytical tools which artificially  
497 determine the gustatory perceptions (del Valle, 2012; Smyth & Cozzolino, 2013).  
498 These systems consist of an array of sensors coupled with chemometric means of data  
499 processing for the characterization of complex liquid samples (Winqvist, Olsson, &  
500 Eriksson, 2011; Martínez-Bisbal, Loeff, Olivas, Carbó, García-Castillo, López-  
501 Carrero, Tormos, Tejadillos, Berlanga, Martínez-Máñez, Alcañiz, & Soto, 2017;  
502 Kumar, Ghosh, Tudu & Bandyopadhyay, 2017; Rudnitskaya, Schmidtke, Reis,  
503 Domingues, Delgadillo, Debus, Kirsanov, Legin, 2017). Following adequate  
504 calibration and training, the electronic tongue is able to determine the qualitative and  
505 quantitative chemical composition of more chemical species in complex samples  
506 (Lvova, Di Natale & Paolesse, 2017; Gutiérrez, Haddi, Amari, Bouchikhi, Mimendia,  
507 Cetó, & del Valle, 2013; Immohr, Hedfeld, Lang, & Pein, 2017).

508 The general scheme which describes the concept of electronic tongue is outlined in  
509 Fig. 2.

510 Fig. 2. General scheme of an electronic tongue system

511 Electronic tongue comprises three components: (1) automatic sampler, which may be  
512 necessary, but it is featured in the majority of commercial systems; (2) array of  
513 sensors with different selectivity and sensitivity and (3) chemometric software with  
514 proper algorithms for processing the signals from sensors and delivering the results  
515 (del Valle, 2012; Ciosek & Wróblewski, 2007; Kalit, Marković, Kalit, Vahčić, &  
516 Havranek, 2014; Tahara & Toko, 2013).

517 Usually, the initial studies dedicated to the development of electronic tongues with  
518 sensors based on various detection systems focused on the qualitative and quantitative  
519 analysis of the solutions which represent basic tastes (sweet, sour, salty, bitter and  
520 umami), as well as of other gustatory sensations or perceptions (astringency,  
521 pungency) (Riul Jr., dos Santos Jr., Wohnrath, Di Tommazo, Carvalho, Fonseca,  
522 Oliveira Jr., Taylor, & Mattoso, 2002; Eckert, Pein, Reimann, & Breitzkreutz, 2013;  
523 Tian, Feng, Xiao, Song, Li, Liu, Mao, & Li, 2015; Pioggia, Di Francesco, Marchetti,  
524 Ferro, Leardi, Ahluwalia, 2007; Jain, Panchal, Pradhan, Patel, & Pasha, 2010;  
525 Rudnitskaya, Polshin, Kirsanov, Lammertyn, Nicolai, Saison, Delvaux, Delvaux, &  
526 Legin, 2009; Toko, 1998; Legin, Rudnitskaya, Clapham, Seleznev, Lord, & Vlasov,  
527 2004; Khan, Khalilian, & Kang, 2016; Arrieta, Rodriguez-Mendez, & de Saja, 2003;  
528 Apetrei, Rodríguez-Méndez, Parra, Gutierrez, & de Saja, 2004; Arrieta, Apetrei,  
529 Rodríguez-Méndez, & de Saja, 2004). This is absolutely necessary in order to prove  
530 that the sensor responds to compounds with various organoleptic properties. The main  
531 compounds analyzed, as well as their sensorial properties, are presented in Table 3.

532 Table 3. The main sensorial properties and their relative compounds.

533 For developing the arrays of sensors, more types of sensors have been used:  
534 electrochemical (potentiometric, voltammetric, amperometric, impedimetric,  
535 conductimetric), optic or enzymatic (biosensors).

536 Most electronic tongue systems reported in the specialized literature are based on  
537 potentiometric sensors (Mimendia, Gutiérrez, Leija, Hernández, Favari, Muñoz, & del  
538 Valle, 2010; Ciosek & Wróblewski, 2011; Cuartero, Carretero, Garcia, & Ortuño,  
539 2015). By using the potentiometric methods, one measures the potential between two  
540 electrodes in the absence of an external flow of current. The value of potential  
541 measured under these circumstances is used for the quantitative determination of the  
542 analytical species of interest in the multicomponent liquid solution (Bard & Faulkner,  
543 2001; Zoski, 2007; Wang, 2000).

544 Potentiometric sensors present a number of advantages, such as: their functioning  
545 principle is well-known, there is a possibility to obtain selective sensors, low cost,  
546 high possibility of industrial production, and the detection is very similar to the  
547 principle of molecular recognition, i.e., with the principle of biologic detection of the  
548 substances responsible of taste. Their disadvantages are their being temperature  
549 dependant and the fact that the adsorption of the solution compounds in the sensitive  
550 element modifies the value of the measured potential (Bratov, Abramova, & Ipatov,  
551 2010; Bobacka, Ivaska, & Lewenstam, 2008).

552 Potentiometric sensors are most often used in the development of electronic tongues  
553 with various applications: fermentation processes monitoring, identification of the  
554 botanic origin of honey, evaluation of the impact of micro-oxygenation in the process  
555 of wine aging in the presence of oak chips, etc. (Gerstl, Joksch, & Fafilek, 2013; Peris

556 & Escuder-Gilabert, 2013; Dias, Veloso, Sousa, Estevinho, Machado, & Peres, 2015;  
557 Schmidtke, Rudnitskaya, Saliba, Blackman, Scollary, Clark, Rutledge, Delgadillo, &  
558 Legin, 2010; Mednova, Kirsanov, Rudnitskaya, Kilmartin, & Legin, 2009; Gutiérrez-  
559 Capitán, Vila-Planas, Llobera, Jiménez-Jorquera, Capdevila, Domingo, & Puig-Pujol,  
560 2014).

561 Another category of sensors which has been widely used for the development of  
562 electronic tongues are the voltammetric sensors (Bard & Faulkner, 2001; Zoski, 2007;  
563 Wang, 2000). In this case, a potential, either fix or, most often, variable, is introduced  
564 into the system, and the electroactive compounds present in the sample are oxidized  
565 or reduced, which leads to the generation of a flow of anodic or cathodic current.  
566 When the sample to be analyzed is a complex one, containing more chemical species  
567 with redox properties, the selectivity of this type of sensors is limited for a specific  
568 analyte present in the sample. The greatest disadvantage of this type of sensors is their  
569 reduced selectivity, but this aspect can be improved by using nanomaterials or by  
570 employing pulse techniques (differential pulse voltammetry and square-wave  
571 voltammetry) or by optimization of the experimental conditions (Brett & Fungaro,  
572 2000; Gupta, Jain, Radhapyari, Jadon, Agarwal, 2011; Reza Ganjali, Garkani Nejad,  
573 Beitollahi, Jahani, Rezapour, & Larijani, 2017; Rodríguez-Méndez, Apetrei, & de  
574 Saja, 2008).

575 The complexity of the voltammetric signals is even more complicated in the case of  
576 sensors which contain electroactive substances immobilized onto the sensitive  
577 element. The interpretation of results is often difficult, as the interactions are  
578 extremely complex, electrocatalytic, synergetic or inhibition effects may occur. This  
579 is why, in most cases, it is necessary to use analytical methods for multivariate data

580 (Cetó, Apetrei, del Valle, & Rodríguez-Méndez, 2014; Winqvist, 2008; Bueno, de  
581 Araujo, Salles, Kussuda, & Paixão, 2014; del Valle, 2010).

582 Numerous research groups have developed various multisensory systems based on  
583 voltammetric sensors (metallic electrodes, electrodes based on nanocomposite  
584 materials, chemically-modified electrodes, etc.) for the studies of different industrial  
585 products (Campos, Alcañiz, Aguado, Barat, Ferrer, Gil, Marrakchi, Martínez-Mañez,  
586 Soto, & Vivancos, 2012; Domínguez, Moreno-Barón, Muñoz, & Gutiérrez, 2014;  
587 Campos Sánchez, Bataller Prats, Gandía Romero, Soto Camino, Martínez Mañez, &  
588 Gil Sánchez, 2013; Winqvist, 2008; Cetó, Capdevila, Puig, & del Valle, 2014;  
589 Apetrei & Apetrei, 2014).

590 The detection principle of the conductimetric sensors is based on the change in the  
591 conductivity of the sensible material as a result of the interaction with various  
592 chemical species present in the solution to be analysed. There are only a few studies  
593 in the literature which tackle the use of conductimetric sensors in the development of  
594 electronic tongues (Winqvist, Holmin, Krantz-Rückler, Wide, Lündström, 2000; Sha,  
595 2013).

596 The measurement principle of impedance sensors is based on measuring the  
597 impedance at a certain frequency value or for a range of frequencies with the help of  
598 impedance spectroscopy. This type of sensors, based on various materials, has been  
599 largely used in the development of electronic tongues with various applications  
600 (Cabral, Bergamo, Dantas, Riul Jr, & Giacometti, 2009; Guo, Chen, Yang, & Wang,  
601 2005).

602 The detection principle of piezoelectric sensors is based on the piezoelectric  
603 phenomenon. The result of the exposure of these sensors to various substances is the

604 modification of their mass due to adsorption or absorption processes, which modify  
605 the resonance frequency of the sensor. Therefore, the electric current is modified, i.e.,  
606 the exit signal provided by the sensor. The advantages of these types of sensors are:  
607 high sensibility, durability, low costs, and reduced size. The detection principle is  
608 based on mass modification (Pearce, Schiffman, Troy Nagle, Gardner, 2006). The  
609 advantages of these types of sensors are: high sensibility, durability, low costs, and  
610 reduced size. The electronic tongues with piezoelectric sensors arrays have been used  
611 for various applications in food analysis (Sehra, Cole, & Gardner, 2004; Kalit,  
612 Marković, Kalit, Vahčić, Havranek, 2014).

613 Colorimetric sensors are based on the interaction between electromagnetic radiation  
614 and matter, from which various phenomena, such as reflection, fluorescence or  
615 absorption, result. This type of sensors contains a source of light or a series of filters  
616 for a specific wave length for increasing selectivity, an indicator, and a detector. The  
617 properties of the indicator are modified as a result of the interaction with the  
618 substance to be analysed, and consequently, a change in absorbance or fluorescence  
619 occurs. The changes are quantified by the detector, which converts the optical signal  
620 in electrical signal. Colorimetric sensors present the following advantages: simplicity,  
621 low cost, and high selectivity. In addition, it is possible for these sensors to detect  
622 non-electroactive substances which cannot be detected by electrochemical sensors.  
623 The disadvantages of the colorimetric sensors are: low durability and distortion of the  
624 exit signal, which greatly limits their applications (Piriya, Joseph, Daniel,  
625 Lakshmanan, Kinoshita, Muthusamy, 2017; Kangas, Burks, Atwater, Lukowicz,  
626 Williams, & Holmes, 2017). In the literature, there are several papers which report on  
627 the use of electronic tongues based on colorimetric sensors in food analysis

628 (Gutiérrez, Llobera, Vila-Planas, Capdevila, Demming, Büttgenbach, Mínguez, &  
629 Jiménez-Jorquera, 2010; Chung, Park, Park, Kim, Park, Son, Bae, & Cho, 2015) .

630 Bioelectronic tongue systems are endowed with biosensors arrays which can  
631 specifically determine a number of analytes of interest for a certain sample. However,  
632 when using certain detection methods, interferences are significant, and there can be  
633 obtained signals which may be assimilated to a chemical impression, which can be  
634 used for the discrimination and classification of the analyzed samples (Ahn, An,  
635 Song, Park, Lee, Kim, Jang, & Park, 2016; Song, Jin, Ahn, Kim, Lee, Kim, Simons,  
636 Hong, & Park, 2014). Bioelectronic tongue systems have been successfully used in  
637 the qualitative and quantitative analysis of various foods (Zeravik, Hlavacek, Lacina,  
638 & Skládal, 2009).

639 The comparison between electronic tongues based on different type of sensors were  
640 reported in literature. For instance, a hybrid electronic tongue based on six chemically  
641 modified graphite-epoxy voltammetric sensors and 15 potentiometric sensors was  
642 applied in the recognition of beer types (Gutiérrez, Haddi, Amari, Bouchikhi,  
643 Mimendia, Cetó, & del Valle, 2013). In other study the data obtained with two sets of  
644 voltammetric sensors, prepared using different strategies, have been combined in an  
645 electronic tongue to evaluate the antioxidant properties of red wines (Cetó, Apetrei,  
646 del Valle, & Rodríguez-Méndez, 2014). Furthermore, the purpose of a complex study  
647 was to compare the performance characteristics of six different e-tongues applied to  
648 the same set of pharmaceutical samples. Two commercially available electronic  
649 tongues (from AlphaMOS and Insent) and four laboratory prototypes (one  
650 potentiometric system from St. Petersburg University, two potentiometric systems  
651 from Warsaw University operating in flow and static modes, one voltammetric system

652 from Barcelona University) were employed (Pein, Kirsanov, Ciosek, del Valle,  
653 Yaroshenko, Wesoly, Zabadaj, Gonzalez-Calabuig, Wróblewski, & Legin, 2015).

654 The advantages of electronic tongues compared to the classical analytical methods  
655 include: high sensitivity, easy building and use, low costs of equipment and price per  
656 analysis, as well as short time necessary for analysis. Through miniaturizing and  
657 automating, electronic tongues can be used for on-line, in-line or real-time analyses,  
658 another advantage being that it is a non-destructive analytical method (Khan,  
659 Khalilian, & Kang, 2016; Cetó, González-Calabuig, del Valle, 2015; Medina-Plaza,  
660 García-Hernandez, de Saja, Fernandez-Escudero, Barajas, Medrano, García-Cabezon,  
661 Martin-Pedrosa, & Rodriguez-Mendez, 2015 ).

662 Nevertheless, research in this field is necessary in what concerns aspects such as:  
663 sensor-obtaining technologies, data processing, system calibration and validation of  
664 results. Researchers in this field grant special attention to these themes, and most of  
665 the recent studies are more and more thorough and present clear applications in  
666 various fields.

667

### 668 5.3. Biosensors

669 Biosensors are analytical devices which integrate a bioreceptor (enzymes, organelles,  
670 living cells, tissues, nucleic acids, aptamers, etc.) in a compatible transducing system,  
671 and which are capable to specifically determine certain chemical compounds (Rotariu,  
672 Lagarde, Jaffrezic-Renault, & Bala, 2016; Scognamiglio, Arduini, Palleschi, & Rea,  
673 2014; Di Rosa, Leone, Cheli, & Chiofalo, 2017). The most frequently used  
674 transducers are: electrochemical, optical, mass, thermal, but there are other types as

675 well (Compagnone, Di Francia, Di Natale, Neri, Seeber, & Tajani, 2017; Ali, Najeeb,  
676 Ali, Aslam, & Raza, 2017; Almeida Silva, Cruz Moraes, Campos Janegitz, Fatibello-  
677 Filho, 2017; Chauhan, Maekawa, & Kumar, 2017). An electric signal which can be  
678 measured and recorded is produced as a result of the specific interaction between the  
679 analyte and the biocomponent. The analytes or target compounds comprise a large  
680 and various number of chemical species, from inorganic compounds to organic  
681 compounds with small molecules and even with large molecules such as proteins  
682 (Abdulbari & Basheer, 2017; El-Nour, Salam, Soliman, & Orabi, 2017; Matysik,  
683 2017; Leca-Bouvier & Blum, 2005). The scheme of analytes detection with  
684 biosensors is presented in Fig. 3.

685 Fig. 3. Biosensor detection scheme

686 When compared to classical methods of analysis, biosensors present a number of  
687 advantages, such as: extremely high selectivity, which allows the detection of the  
688 target molecule in real complex samples, without requiring the pre-treatment of the  
689 sample, short time of analysis (from a few seconds to a few minutes), relatively low  
690 costs, possibility of miniaturizing and turning them into portable devices, which  
691 allows fast and precise on-site, in-line, on-line or real time analytical determinations  
692 (Scognamiglio, Rea, Arduini, & Palleschi, 2017; Shao, Wang, Wu, Liu, Aksay, &  
693 Lina, 2010; Mehrotra, 2016).

694 Food quality control, as well as the detection or monitoring of the food spoilage  
695 processes, requires methods and tools for the precise analysis of various parameters.  
696 Biosensors can accomplish these functions, which is why the special interest in  
697 developing new biosensors which can be used in food analysis for example, for  
698 determining freshness or spoilage, is fully justified (Dornelles Mello & Tatsuo

699 Kubota, 2002; Poltronieri, Mezzolla, Primiceri, & Maruccio, 2014; Pividori &  
700 Alegret, 2010).

701 The main research directions include the analysis of compounds of interest for food  
702 quality and that of contaminants, compounds which accidentally appear in food and  
703 which should not be there under normal conditions (McGrath, Elliott, & Fodey, 2012;  
704 Dragone, Grasso, Muccini, & Toffanin, 2017). Moreover, focus is laid on monitoring  
705 various chemical or biochemical processes related to fermentation, degradation,  
706 spoilage, maturation or freshness of foods with the help of the biosensors (Mutlu,  
707 2016; Vasilescu, Nunes, Hayat, Latif, & Marty, 2016; Adley, 2014; Ispas, Crivat, &  
708 Andreescu, 2012; Park, Kim, Lee, & Jang, 2015). Other studies lay importance on the  
709 characterization of foods in terms of biologic or geographic origins, as well as  
710 authenticity, fraud or adulteration of foods (Apetrei & Ghasemi-Varnamkhasti, 2013;  
711 Bassi, Lee, & Zhu, 1998; Narsaiah, Jha, Bhardwaj, Sharma, & Kumar, 2012;  
712 Campuzano, Ruiz-Valdepeñas Montiel, Torrente-Rodríguez, Reviejo, & Pingarrón,  
713 2016).

714 The classification of the biosensors can be made according to several criteria, the  
715 most often being the biochemical recognition mechanism (Thévenot, Toth, Durst,  
716 Wilson, 2001; Monošík, Stred'anský, Šturdík, 2012; Apetrei & Ghasemi-  
717 Varnamkhasti, 2013; Gorton, 2005).

718 Enzyme-based biosensors are the most frequently used in foods analysis (Kumar &  
719 Neelam, 2016; Prodromidis & Karayannis, 2002). Two basic principles are used in  
720 practice, one being the direct detection of the analyte (substrate) resulted from an  
721 enzymatic process, the other being the inhibition of the enzymatic activity (Upadhyay  
722 & Nishant, 2013; Murugaboopathi, Parthasarathy, Chellaram, Prem Anand, &

723 Vinurajkumar, 2013). Enzymes in the class of oxidoreductases (laccase, tyrosinase,  
724 peroxidase, dehydrogenases) are used for substrate detection, and the main  
725 electroactive compounds detected by these biosensors are o-quinone derivatives,  
726 hydrogen peroxide or reduced forms of nicotinamide adenine dinucleotide (Amine,  
727 Mohammadi, Bourais, & Palleschi, 2006; Mello & Kubota, 2002; Tembe & D'Souza,  
728 2015). The enzyme sources can be purified enzymes commercially available, but also  
729 organelles, cells, tissues, microorganisms, etc. (Apetrei & Apetrei, 2016; Rodríguez-  
730 Delgado, Alemán-Nava, Rodríguez-Delgado, Dieck-Assad, Martínez-Chapa, Barceló,  
731 Parra, 2015; Gul, Sheeraz Ahmad, Saqlan Naqvi, Hussain, Wali, Farooqi, & Ahmed,  
732 2017; Liu, Wu, Cai, Hu, Zhou, & Wang, 2014; Hasan, Nurunnabi, Morshed, Paul,  
733 Polini, Kuila, Al Hariri, Lee, & Jaffa, 2014; Lim, Ha, Lee, Lee, & Kim, 2015). For the  
734 detection of inhibitors of enzymatic activity, the activity of the enzyme is determined  
735 in the absence and in the presence of the inhibitor, determining the inhibition degree  
736 based on inhibitor concentration. The detection of target compounds does not involve  
737 its transformation (Upadhyay & Nishant, 2013; Murugaboopathi, Parthasarathy,  
738 Chellaram, Prem Anand, & Vinurajkumar, 2013).

739 The detection principle of affinity biosensors is based on molecular recognition  
740 systems, such as the interaction between DNA (Deoxyribonucleic acid) strands,  
741 antigen – antibody or hormone – receptor interactions (Patel, 2006; Turner, 2013;  
742 Rogers, 2000). Another class of compounds used in the production of this types of  
743 biosensors is molecularly imprinted polymers (Song, Xu, Chen, Wei, & Xiong, 2014;  
744 Frasco, Truta, Sales, & Moreira, 2017; Wackerlig & Schirhagl, 2016).

745 Nano biosensors are emerging as a promising tools for the applications in the food  
746 analysis. They are integrating knowledge of physical sciences, biology, chemistry,  
747 biotechnology, molecular engineering, and nanotechnology offering important

748 improvements in selectivity and sensitivity compared to classical chemical and  
749 biological methods. Nano biosensors can be used for detection and quantification of  
750 microorganisms, contaminants, and food freshness (Pérez-López, & Merkoçi, 2011;  
751 Grumezescu, 2016).

## 752 **6. Literature evidence multisensor systems to food spoilage detection**

### 753 *6.1. Electronic nose*

754 There are several electronic nose systems, including different types of and gas sensors  
755 and systems combined with other techniques and using different data processing  
756 methods for the detection and characterization of food spoilage. Some successful  
757 experiments performed by different authors have been described in the bibliography.  
758 As a general rule, there are some chemical compounds that are responsible for defects  
759 and off-flavors in food and beverages. These compounds are known by consumers as  
760 the first alarm signal linked to spoilage. It is very important to optimize the  
761 measurement system to detect these compounds. Table 4 summarizes the sensors and  
762 sensory systems applications for detection and characterization of spoilage in the food  
763 industry.

764 **Table 4.** A summarized overview on the application of electronic nose to food  
765 spoilage detection

766 There are different prototypes designed by some research groups with different  
767 features that are appropriate for different applications. In the bibliography, Laboratory  
768 equipment as well as portable instruments are designed for food spoilage detection.  
769 The following reference (Jose Pedro Santos & Lozano, (2015) shows a hand-held  
770 wireless portable electronic nose applied to the real-time detection of two common

771 aromatic defects in beer: acetaldehyde and ethyl acetate. An image of the electronic  
772 nose is illustrated in Fig. 4. These aromatic defects in beer have been measured at  
773 level between the organoleptic threshold and five times this quantity (25 ppm for  
774 acetaldehyde and 21 ppm for ethyl acetate). PCA were applied to these responses to  
775 see the data distribution among classes. Although there is some confusion between  
776 some classes corresponding to different concentrations, non-defect beer samples were  
777 separated from the other samples. In a qualitative classification among beer without  
778 defects (blank) and beer with one of the defects (ethyl acetate or acetaldehyde)  
779 regardless the concentration, the measurements were grouped into three classes:  
780 blank, ethyl acetate and acetaldehyde. The PCA score plot for the whole measurement  
781 set is shown in Fig. 4. Some partial overlapping is observed among the classes,  
782 although the ANN analysis gave a 94 % success rate in validation. Few samples are  
783 wrongly classified among the three classes. Authors explain that these results could be  
784 improved using other types of classifiers and improving the measurement system in  
785 order to a better control of the operation temperatures and flows and reducing the  
786 measurement noise.

787

788 Fig. 4. Portable e-nose system for the defect discrimination in beer and PCA score  
789 plot of measurements of beer defects.

790 It is usually recognized that electronic noses have not achieved the market penetration  
791 that was expected in the mid-90s. The prototype presented in Lozano et al., (2015)  
792 could be a first step for implementation in the wine industry. It is installed in a wine  
793 cellar for on-line monitoring of wine evolution during 9 months. The system has a  
794 novel sampling method that extracts the aroma directly from the tanks where wine is

795 stored; and it automatically carries the volatile compounds to the sensor cell with tin  
796 oxide multisensor. Linear techniques as principal component analysis (PCA) and  
797 nonlinear ones as Artificial Neural Arrays (ANN) are used for pattern recognition,  
798 and Partial Least Squares (PLS) is used for predicting GC-MS analysis. Results  
799 showed that system can detect the evolution of two different wines along 9 months  
800 stored in the monitored tanks. The evolution of the wine is confirmed with chemical  
801 and sensory analysis. Moreover, GC-MS analysis was performed to the wine of the  
802 tanks. In the whole, 19 odorants were analysed. The chemical compounds analysed  
803 were acids (butyric acid, decanoic acid, hexanoic acid, isobutyric acid, isovaleric acid,  
804 and octanoic acid), alcohols (1-hexanol and 2-phenylethanol), esters (hexyl acetate,  
805 ethyl butyrate, ethyl decanoate, ethyl hexanoate, ethyl isovalerate, ethyl lactate, ethyl  
806 octanoate, isoamyl acetate, isobutyl acetate, diethyl succinate and phenyl ethyl  
807 acetate) and phenols (4-vinyl-guaiacol). The aforementioned 19 compounds analysed  
808 in GC-MS profiles were used as predictor variables. Then, a model was created in  
809 order to predict these responses from sensor measurements. In this way, the  
810 concentration of chemical compounds in wine determined by GC-MS were correlated  
811 with electronic nose response PLS regression analysis. Correlation coefficients near to  
812 1 are obtained in the prediction of several volatile organic compounds (VOCs), i.e.  
813 ethyl butyrate, isobutyric acid, isobutyl acetate, hexyl acetate and ethyl octanoate.  
814 This system could be trained for monitoring wine preservation and evolution in tanks  
815 and therefore detecting off-odours of wine and warning the wine expert to correct it as  
816 soon as possible, preventing the wine spoilage and improving its final quality.

817 Based on the body of scientific literature, numerous considerable works on spoilage  
818 detection using electronic nose has been conducted on meat and fish products.  
819 Chemical reaction between volatile compounds involved in spoiled meat with gas

820 sensors has imperative results and this measuring principle is the basis of the spoilage  
821 detection in meat products (Wojnowski, Majchrzak, Dymerski, Gębicki, Namieśnik,  
822 2017).

823 Meat spoilage as a tremendously complex phenomenon is affected by many  
824 parameters such as storage conditions, packaging type and materials used,  
825 temperature and so on. Innovative instrumental approaches such as electronic nose  
826 have shown promising results to be used as a potential candidate for inspection of  
827 meat and its spoilage. A list of the most applications on such products is summarized  
828 in Table 4. For instance, two cases of the more recent applications are discussed here.

829 Estelles-Lopez et al., (2017) conducted a research to develop the appropriate models  
830 for predicting minced beef spoilage. For this aim, a commercial electronic nose  
831 ((LibraNose, Technobiochip, Napoli, Italy) comprising eight quartz crystal  
832 microbalance (QMB) sensors coated with different poly-pyrrole derivatives was used.  
833 Based on the planned experimental protocol, few grams of the meat was inserted in a  
834 container and left for a moment to collect the adequate headspace as called static  
835 sampling. Then the volatile compounds present in the headspace were passed over the  
836 sensors and the responses registered and saved. The authors have also used four  
837 analytical instruments to fuse the data with electronic nose. They were Gas  
838 Chromatography-Mass Spectrometry (GC-MS), High Performance Liquid  
839 Chromatography (HPLC), multispectral imaging (MSI), and Fourier Transformed  
840 Infrared Spectroscopy (FT-IR). For data fusion and analyses, numerous techniques as  
841 given in Table 4, were used and modeled. In final, they developed an on line platform  
842 to identify different types on microorganisms present in spoiled meat. Electronic nose  
843 showed satisfactory contribution for this aim.

844 Lipid oxidation as a spoilage indicator was studied by Gu, Sun, Tu, & Pan (2017) who  
845 aimed their research at evaluating the odor of Chinese-style sausage as a high-fat meat  
846 product during processing and storage using electronic nose. During lipid oxidation,  
847 some chemical changes occur in the sausage where some volatile compounds  
848 involved in the sample headspace are found such as certain aldehydes, ketones and  
849 alcohols. Monitoring these compounds could help in lipid oxidation prediction and  
850 spoilage detection consequently. They used a portable electronic nose (PEN 3, Win  
851 Muster Air-sense Analytics Inc., Germany) consisting of ten metal oxide sensors  
852 which were extremely sensitive to a lot of volatile compounds as nitrogen oxides,  
853 ammonia and aromatic compounds, Benzene, hydrogen, alkenes and aromatic  
854 compounds, Propane, methane, sulphur compounds, alcohols, sulphur organic  
855 compounds, alkane). The sensors were non selective and partial sensitive to aromatic  
856 compounds. The time of the measurement was 60 s and 110 s for odor injection and  
857 purging periods, respectively. Win Muster software was exploited to transform the  
858 information to digital signals. As mentioned in Table 4, many data processing  
859 algorithms were used to classify the samples. The authors concluded that the results  
860 show great potential use of electronic nose in judging the lipid oxidation of the high-  
861 fat meat products.

862

## 863 *6.2. Electronic tongue*

864 Electronic tongues have been successfully used for qualitative and quantitative  
865 determinations of the spoilage of many foods of interest (Haddi, El Barbri, Tahri,  
866 Bougrini, El Bari, Llobet, & B. Bouchikhi, 2015; Śliwinska, Wisniewska, Dymerski,  
867 Namiesnik, & Wardencki, 2014). As it is well-known, the foods spoilage is a complex  
868 biochemical and microbiologic process which involves atmospheric oxygen, the

869 activity of some specific enzymes and microorganisms, etc. (Sahu & Bala, 2017; de  
870 Blackburn, 2006).

871 Thus, for the quantitative case, a number of toxic compounds formed during the  
872 spoilage process has been determined, especially biogenic amines, which result from  
873 amino acids decarboxylation. The amino acids involved in these processes are free  
874 amino acids present in foods, but also the ones which originate in proteins hydrolysis  
875 (Naila, Steve Flint, Fletcher, Bremer, & Meerdink, 2010; Karovičová & Kohajdová,  
876 2005). Other quantitatively determined compounds are inosine 5'-monophosphate,  
877 inosine and xanthine and hypoxanthine, which originate from adenosine triphosphate  
878 (ATP) degradation (Vilas, Alonso, Herrera, García-Blanco, & García, 2017) (Fig. 5).

879 Fig. 5. Decomposition of ATP in the muscles (Nelson & Cox, 2017)

880 Where, ATP: Adenosine triphosphate; ADP: Adenosine diphosphate; AMP:  
881 Adenosine monophosphate, IMP: Inosine monophosphate; Ino: Inosine; Hx:  
882 Hypoxanthine; Xa: Xanthine; PI: phosphate ion.

883 Quantitative determination is generally acquired from statistic models obtained  
884 according to the data recorded with the sensor system of the electronic tongue, which  
885 allow quantitative estimations of certain physical-chemical or sensorial parameters  
886 (e.g. partial least squares–discriminant analysis (PLS-DA) or PLS2 regression  
887 models) (Haddi, El Barbri, Tahri, Bougrini, El Bari, Llobet, & B. Bouchikhi, 2015;  
888 Rodríguez-Méndez, Gay, Apetrei, & de Saja, 2009).

889 More types of foods have been analyzed and the systems used and the main results  
890 obtained are presented in the following paragraphs.

891 The concept of meat freshness is quite complex, including various physicochemical,  
892 biochemical and microbiologic characteristics related to two different processes – the  
893 former, aging, determined by the storage period required by meat in order to acquire  
894 the proper taste for consumption, and the latter, also in relation to the period of  
895 storage, which leads to meat spoilage due to bacterial growth and autolysis (Iulietto,  
896 Sechi, Borgogni & Cenci-Goga, 2015; Dave & Ghaly, 2011).

897 Gil et al. (2011) presented a case study of the use of potentiometric electronic tongue  
898 in the study of the spoilage process of a whole piece of pork loin stored under  
899 refrigeration (Gil, Barat, Baigts, Martínez-Máñez, Soto, Garcia-Breijo, Aristoy,  
900 Toldrá, Llobet, 2011). The sensors array used in the developing of the electronic  
901 tongue consisted of six electrodes made of Au, Ag, Cu, Pb, Zn and C, and a reference  
902 electrode. By using more methods in the multivariate data analysis (PCA and artificial  
903 neural arrays - multilayer perceptron and fuzzy ARTMAP), the authors proved that  
904 the potentiometric electronic tongue is capable to determine the storage time, which is  
905 in relation to the degradation of the pork loin.

906 For data validation and for establishing the correlation with the results of classical  
907 analytical methods, a number of physical-chemical, microbial and biochemical  
908 parameters were analysed. These analyses consisted in pH determination, microbial  
909 count, concentrations of inosine 5'-monophosphate, inosine and hypoxanthine. Using  
910 the PLS regression method, a very good correlation was found between pH and the  
911 data obtained from potentiometric sensors, as well as between K-index  
912 (simultaneously measures the variation in the adenosine triphosphate) and the data  
913 obtained with the electronic tongue. The conclusion of the study was that the  
914 potentiometric electronic tongues are very useful in the qualitative or semi-

915 quantitative evaluation of freshness in meat samples and they can have numerous  
916 applications in food industry in quality control of pork meat.

917 Another study, presented by Kaneki et al., (2004) described the use of a  
918 potentiometric electronic tongue based on simple solid electrodes (i.e. Pt, CuS and  
919 Ag<sub>2</sub>S) which are able to detect certain compounds responsible for the initial stage of  
920 meat putrefaction. This system was successfully used in the study of pork meat  
921 freshness (Kaneki, Miura, Shimada, Tanaka, Ito, Hotori, Akasaka, Ohkubo, & Asano,  
922 2004).

923 Microbiological contamination in dry-cured ham can occur at various stages of the  
924 maturation process, and the development of a large number of microorganisms  
925 involved in spoilage may lead to the alteration of the end product (Dikeman &  
926 Devine, 2014). These processes lead to some unpleasant and non-common odours,  
927 which are detected by an expert taster, who follows a procedure called “cala”, by  
928 which he classifies hams as good and altered hams (Paarup, Nieto, Peláez, & Reguera,  
929 1999). Girón et al. (2015) produced a potentiometric electronic tongue based on an  
930 array of sensors which contains three types of sensors, silver, nickel and copper  
931 electrodes. This electronic tongue was used for the classification of altered and  
932 unaltered hams before the classification of hams by an expert tester. The results of the  
933 analyses showed that, in the case of altered hams, the Ag potentials have the lowest  
934 values and the Cu potentials, the highest values. Starting from these experimentally  
935 observed differences, a model of classification of hams was built, but further studies  
936 are required for the system validation for industrial practice (Girón, Gil-Sánchez,  
937 García-Breijo, Pagána, Barat, & Grau, 2015).

938 Gil-Sánchez et al. (2011) presented the use of a combined multisensor system for the  
939 analysis of the spoilage of wine when it is in contact with air (Gil-Sánchez, Soto,  
940 Martínez-Máñez, Garcia-Breijo, Ibáñez, & Llobet, 2011). The system consists of a  
941 potentiometric electronic tongue and a humid electronic nose. The potentiometric  
942 electronic tongue was used for the evolution in time of the wine samples in the  
943 presence of air. The classical method of analysis used for monitoring the wine  
944 spoilage was the determination of the titratable (total) acidity. The electronic tongue  
945 used in this study is based on potentiometry. Potentiometric sensors were built using  
946 thick-film serigraphic techniques. The paste used for making the sensors was  
947 commercial, generally used for the production of thick-film resistances and  
948 conductors for hybrid electronic circuits. Each paste contains an active element,  
949 which are, in this case, Ag, Au, Cu, Ru, AgCl, and C. These sensitive materials are  
950 often used in the production of non-specific electrodes. Some materials were used in  
951 duplicate for the production of sensors, by modifying, for instance, the thickness of  
952 the sensitive layer, 9 potentiometric sensors being included in the multisensor system.  
953 Fig. 6 presents the distribution of the sensors on the multisensor pad and the tracks  
954 and pads for connecting to measuring equipment.

955 Fig. 6. The sensor array used for the potentiometric electronic tongue (Gil-Sánchez,  
956 Soto, Martínez-Máñez, Garcia-Breijo, Ibáñez, & Llobet, 2011).

957 Ruiz-Rico et al. (2013) studied the shelf-life assessment of fresh cod in cold storage  
958 using a voltammetric electronic tongue (Ruiz-Rico, Fuentes, Masot, Alcañiz,  
959 Fernández-Segovia, & Barat, 2013). The electronic tongue system is based on an  
960 array of sensors, specialised software installed on a PC and electronic equipment.  
961 Measurements relied on pulse voltammetry, the voltage pulses being applied to  
962 sensors by the electronic equipment, and the generated currents being measured

963 afterwards. For each sensor, 1,000 values were recorded, which correspond to the  
964 time evolution of the current generated in the system after applying the voltage pulse.  
965 The sensor system is made up of 8 metallic electrodes, separated into two subsystems,  
966 one made up of 4 electrodes based on noble metals (iridium, rhodium, platinum and  
967 gold) and the other, of 4 metallic electrodes based on non-noble metals (silver, cobalt,  
968 copper and nickel). Therefore, a total of 8,000 values are registered by the electronic  
969 tongue for each sample under study. For the validation of the analytical system, data  
970 resulted from physical-chemical and microbial analyses were used. For all samples  
971 analysed, the limits of the main parameters related to fish freshness, such as total  
972 volatile basic nitrogen, mesophilic and Enterobacteriaceae, were exceeded on the  
973 fourth day of storage, which means that fish has a shelf-life less than four days. The  
974 results of physical-chemical and microbial analyses showed an obvious loss of  
975 freshness from day 0 to day 4. Also, the voltammetric tongue results showed a clear  
976 difference between the freshness of fish on days 0 and 1 of storage and that in the  
977 following days. The regression patterns based on partial least squares for Total  
978 Volatile Basic Nitrogen (TVB-N) and mesophilic counts proved that the predicted  
979 values concord with the experimental results, which confirms the usefulness of  
980 voltammetric electronic tongue for assessing cod spoilage.

981 Haddi et al. (2015) implemented a voltammetric electronic tongue based on an array  
982 of seven working electrodes, a platinum counter electrode and an Ag/AgCl reference  
983 electrode (Haddi, El Barbri, Tahri, Bougrini, El Bari., Llobet, & Bouchikhi, 2015).  
984 The working electrodes were made of platinum, gold, silver, glassy carbon,  
985 palladium, copper and nickel. They were assembled in the form of an array of sensors  
986 in a stainless steel tube. The wires of each electrode were connected to a portable

987 potentiostat through a relay box. The responses of the array of sensors in the presence  
988 of the samples to be analyzed were recorded by cyclic voltammetry.

989 With the help of this system, it was objectively and rapidly assessed whether there  
990 were any significant differences between meat types (beef, goat and mutton), and  
991 between the same piece of meat in various spoilage states. The electronic tongue  
992 system, made up of 7 voltammetric sensors, was used for the detection of the specific  
993 electroactive compounds for each of the three types of meat. Data analysis was  
994 pursued using discrimination and classification methods, Principal Component  
995 Analysis (PCA) and Support Vector Machines (SVMs). The results obtained proved  
996 that the system is capable of distinguishing meats based on their biologic origin. Also,  
997 for each type of meat, the number of days passed in cold storage can be determined.

998 A number of studies reported in the literature relied on the use of voltammetric  
999 electronic tongues based on sensors modified with electroactive substances  
1000 (phthalocyanines or conducting polymers), both regular and screen-printed electrodes.

1001 A study reported the use of a novel array of voltammetric sensors used for the  
1002 detection of the principal biogenic amines resulted from the spoilage process of Tench  
1003 fish (Rodríguez-Méndez, Gay, Apetrei, & de Saja, 2009). The array of sensors  
1004 consisted of screen-printed electrodes modified with phthalocyanines. The method  
1005 conveyed in this study entailed the global detection of the chemical products resulted  
1006 from the process of spoilage of fish, including the biogenic amines.

1007 The sensors proved very good sensitivity to biogenic amines present in the solution to  
1008 be analysed (ammonia, dimethylamine, trimethylamine, cadaverine and histamine). It  
1009 was observed that biogenic amines have great influence on the chemical behaviour of  
1010 the sensors, due to the fact that some biogenic amines are electroactive and that all

1011 biogenic amines have basic and nucleophilic properties. The developed sensors are  
1012 very sensitive, reproducible, and present good stability on long term.

1013 The array of sensors was used for the determination of the freshness degree of fish  
1014 kept at 4°C in the refrigerator for 12 days. The responses recorded by cyclic  
1015 voltammetry were successfully used for assessing freshness and for determining the  
1016 post-mortem period. The voltammetric signals displayed increasing intensity with the  
1017 increasing of storage time.

1018 The ability of discriminating fish samples based on their freshness was demonstrated  
1019 by principal component analysis. The ability of classifying the fish samples according  
1020 to their freshness, as well as the prediction of freshness of some samples was  
1021 calculated by partial least squares-discriminant Analysis (PLS-DA). The results  
1022 proved that voltammetric electronic tongue is able for determining the degree of fish  
1023 freshness by monitoring the production of spoilage products. In addition, this method  
1024 is able to determine the stage of the spoilage process, which comprises 4 states.

1025 Another paper reported the use of a voltammetric electronic tongue for monitoring the  
1026 freshness of Pontic shad fish samples (Apetrei, Rodriguez-Mendez, Apetrei, de Saja,  
1027 2013). The samples were Pontic shad (*Alosa Pontica*), a species living in the north-  
1028 western part of the Black Sea. Pontic shad migrates in the Danube River for  
1029 spawning. The array of sensors was made up of a series of sensors based on carbon  
1030 screen printed electrodes modified with polypyrrole doped with different doping  
1031 agents. The electrochemical signals are complex and present redox processes related  
1032 to the electrochemical activity of the amines, and redox peaks associated to the  
1033 electrochemical activity of the electroactive material. The viability of the  
1034 voltammetric electronic tongue was tested for fish freshness monitoring. From the

1035 analysis of the signals registered by sensors, a growth of the signal currents associated  
1036 to biogenic amines was observed in the analysed samples with the increase of the  
1037 storage time.

1038 The voltammetric signals obtained with the help of the array of sensors were used to  
1039 discriminate and evaluate the state of fish freshness. Principal component analysis  
1040 confirmed the ability of the voltammetric electronic tongue to monitor the fish  
1041 freshness. The partial least squares–discriminant analysis (PLS-DA) model showed  
1042 that this electronic tongue is able to determine the post-mortem time elapsed, being  
1043 highly useful in practice.

1044 Another study was dedicated to the detection and quantification of putrescine and  
1045 ammonia resulted from the spoilage of dehydrated beef, as well as to monitoring beef  
1046 freshness under refrigeration conditions (Apetrei & Apetrei, 2016).

1047 The array of sensors used in this study was a hybrid one, made up of screen-printed  
1048 electrodes modified with bisphthalocyanines and polypyrrole doped with different  
1049 doping agents. The electrochemical responses of the sensors were analysed for two  
1050 compounds of interest in beef spoilage, namely ammonia and putrescine.

1051 The electrochemical signals are related to the redox properties of the substances used  
1052 for modifying the electrodes, which are greatly influenced by the compounds present  
1053 in the solution to be analysed. At first, it was determined that the sensors were capable  
1054 to detect amine compounds in beef extract powder with good sensitivity to the levels  
1055 of concentration at which the respective compounds are found in the initial spoilage  
1056 stages. The sensor array made up of sensors with the best performance was used for  
1057 beef freshness monitoring. The methods conveyed for the analysis of experimental

1058 data, PCA and PLS-DA, demonstrated that the electronic tongue system is able to  
1059 discriminate and classify samples according to their refrigeration time.

1060

### 1061 *6.3. Biosensors*

1062 Various types of biosensors have been used for the specific determination of some  
1063 analytes directly related to the spoilage process (Rotariu, Lagarde, Jaffrezic-Renault,  
1064 Bala, 2016). The most important are biogenic amines and the compounds resulted  
1065 from the decomposition of nucleic acids, as is the case of xanthine, hypoxanthine and  
1066 other metabolites (Ghaly, Dave, Budge, & Brooks, 2010). The following section  
1067 reviews the most relevant results reported in the specialized literature, according to  
1068 the type of food under analysis.

1069 Meat and meat products are the foods which have been most often studied using  
1070 biosensors for spoilage detection. The reason is that the products which result from  
1071 the spoilage process are toxic and may lead to intoxication, allergies, and even death  
1072 when ingested in large quantities (Stadler & Lineback, 2008). In order to be fitted  
1073 with consumption, beef must be subject to a refrigeration process for a few days, a  
1074 process that is named “aging” (Perry, 2012). During its refrigeration, besides aging,  
1075 the unwanted process of bacterial spoilage may also occur. Therefore, in order to  
1076 obtain aged meat with optimal organoleptic properties, the simultaneous monitoring  
1077 of aging and bacterial spoilage is necessary. For highlighting the bacterial spoilage  
1078 process, it is necessary to monitor the concentration of putrescine and cadaverine, two  
1079 biogenic amines, which can be considered markers of the spoilage process (Perry,  
1080 2012; Dashdorj, Tripathi, Cho, Kim, & Hwang, 2016; Apetrei & Apetrei, 2016).

1081 Yano et al. (1996) developed a direct sensing method in order to determine the quality  
1082 of beef (Yano, Yokoyama, Tamiya, & Karube, 1996). The biosensor was made of an  
1083 Ag/AgCl electrode and a platinum electrode onto which two enzymes were  
1084 immobilized, namely putrescine oxidase or xanthine oxidase. The detection method  
1085 used was potential-step chronoamperometry, the potential was stepped in the range  
1086 from 0.3 V to 0.6 V. The experimental conditions, such as pH and selectivity, were  
1087 adequate and the target compounds could be analysed on the beef surface. Sensitivity,  
1088 selectivity and stability of the biosensor were very good in detecting putrescine,  
1089 cadaverine and hypoxanthine. The experimental results demonstrated that the method  
1090 of direct determination with this biosensor could be successfully used in the non-  
1091 destructive assessment of beef quality.

1092 Kress-Rogers et al. (1993) developed a prototype biosensor (in the form of an array of  
1093 biosensors) in view of ultra-fast assessment of pork meat freshness (Kress-Rogers,  
1094 D'Costa, Sollars, Gibbs, & Turner, 1993). The biosensors array allows the  
1095 measurement of glucose concentration at 2 and 4 mm depth under the meat surface.  
1096 The array of biosensors was used to monitor the spoilage process of refrigerated pork  
1097 carrying a slaughterhouse flora. The assessment of meat freshness was pursued based  
1098 on the three-dimensional profile of glucose near the meat surface. This method can be  
1099 applied as a marker for the fast evaluation of complex foods, in what concerns the  
1100 microbial and oxidative spoilage, maturation and the fermentation process.

1101 Fish and fish products spoilage is also of great interest in food industry, as fish is  
1102 susceptible to spoilage due to storage conditions. Fish spoilage under refrigeration  
1103 conditions is attributed to the metabolic degradation of trimethylamine N-oxide  
1104 (TMAO) to trimethylamine (TMA) by psychrophilic bacteria. TMA accumulation in  
1105 tissues is responsible for the specific smell of degrading fish, while the TMA

1106 concentration depends on the stage of the spoilage process (Barrett & Kwan, 1985;  
1107 Muzaddadi, Devatkal, & Oberoi, 2016).

1108 Gamati et al. (1991) developed a biosensor for monitoring the trimethylamine  
1109 concentration, based on the difference in the oxygen uptake response of two microbial  
1110 electrodes (Gamati, Luong & Mulchandani, 1991). One of the electrodes was  
1111 produced using *Pseudomonas aminovorans* grown on TMA. It was particularly  
1112 sensitive to TMA, trimethylamine N-oxide, dimethylamine and monomethylamine.  
1113 The other electrode was produced using *Pseudomonas aminovorans* grown on  
1114 TMAO, and it was sensitive to TMA, trimethylamine N-oxide, dimethylamine and  
1115 monomethylamine. The response of biosensor is linear with TMA concentration and  
1116 the limit of detection is in pM domain. Besides, the relative standard deviation of the  
1117 biosensor response is low, the response is stable and reproducible. The results  
1118 obtained with the help of this sensor were validated by HPLC. The biosensor is useful  
1119 for TMA determination in fish tissue extracts.

1120 Another biosensor for the TMA detection was developed by Bourigua et al. (2011). It  
1121 was based on polypyrrole–flavin-containing monooxygenase (FMO3) and ferrocene.  
1122 The detection techniques employed were amperometry and impedance spectroscopy.  
1123 The biosensor presents high selectivity and sensitivity to TMA in real samples. The  
1124 validation of the biosensor was carried out using GC/SM and the real sample was fish  
1125 extract after deterioration during storage (Bourigua, El Ichi, Korri-Youssoufi, Maaref,  
1126 Dzyadevych, & Jaffrezic Renault, 2011).

1127 In food industry, fish processing is difficult because of its low commercial life and  
1128 high variability of the raw material, starting from the biologic species and ending with  
1129 fishing and storage. An important biomarker of fish spoilage is the level of xanthine:

1130 above certain values, it is certain that the spoilage process has begun (Costa &  
1131 Miertus, 1993).

1132 Fish freshness is the most important feature of this raw material for its processing in  
1133 food industry under safe, qualitative conditions. After the fish's death, breathing and  
1134 biosynthesis of adenosine triphosphate (ATP) nucleotide cease. Consequently, the  
1135 ATP in the muscles is degraded, according to the scheme presented in Fig. 5.

1136 Among the spoilage products, IMP is the main factor which contributes to fish  
1137 freshness flavour, and the spoilage product hypoxanthine is what gives the fish meat  
1138 its specific bitter taste. Dervisevic et al. (2015) produced a biosensor based on a host  
1139 matrix nanocomposite for immobilization of xanthine oxidase made up of MWCNT  
1140 incorporate in poly (GMA-co-VFc) copolymer film (Dervisevic, Custiuc, Çevik, &  
1141 Senel, 2015). The inclusion of MWCNT in the polymer matrix resulted in a  
1142 substantial growth of the sensitivity of the biosensor. The fabrication process of the  
1143 sensitive layer of the biosensor was characterized by scanning electron microscopy.  
1144 The electrochemical behaviour of the biosensor was studied by cyclic voltammetry  
1145 and electrochemical impedance spectroscopy. The biosensor presents maximum  
1146 response to xanthine at pH 7.0 and 45°C, when +0.35 V is applied. The biosensor  
1147 reaches 95% of steady-state current in approximately 4 seconds. The limit of  
1148 detection of the biosensor to xanthine detection is of 0.12  $\mu\text{M}$ , positive results being  
1149 obtained for the measurement of xanthine concentration in fish meat. The response of  
1150 the biosensor is stable and the interferences are very low.

1151 Dervisevic et al. (2015) studied the detection of xanthine molecules, which is an  
1152 indicator of meat spoilage (Dervisevic, Custiuc, Çevik, Durmus, Senel, Durmus,  
1153 2015). Xanthine is formed as a result of the decomposition of guanine. To this end,

1154 they developed a novel biosensor by embedding reduced expanded graphene oxide  
1155 sheets decorated with iron oxide ( $\text{Fe}_3\text{O}_4$ ) nanoparticles into poly (glycidyl  
1156 methacrylate-covinylferrocene) phase, and by covalent immobilization of xanthine  
1157 oxidase onto the surface of P(GMA-co-VFc)/REGO- $\text{Fe}_3\text{O}_4$  nanocomposite film. The  
1158 experimental conditions were studied and optimized for the high sensitivity detection  
1159 of xanthine (response time, linear range, operation and storage stability, pH and  
1160 temperature) a limit of detection of  $0.17 \mu\text{M}$  being obtained. The xanthine biosensor  
1161 was used for the analysis of xanthine content in fish real samples after 5, 8, 10, 13, 15,  
1162 and 20 days of storage. The novel biosensor proved that it could be successfully  
1163 employed in the analysis of real samples and also that it could be successfully used as  
1164 a reliable fish freshness controlling technique.

1165 Apetrei et al. (2015) developed a biocomposite screen-printed biosensor based on  
1166 immobilization of tyrosinase onto the carboxyl functionalised carbon nanotube for  
1167 assaying tyramine in fish products (Apetrei & Apetrei, 2015). Tyramine is a biogenic  
1168 amine which is especially found in fermented food products, but also in smoked,  
1169 salted or soured fish (Luten, 2006). This compound can be used as a biomarker for  
1170 spoilage monitoring. The detection principle employed was the amperometric one, by  
1171 applying the optimum potential for the electrochemical reduction of the o-quinone  
1172 formed in the enzymatic process at the surface of the sensitive layer of the biosensor.  
1173 The biosensor presented very good analytical performance in what tyramine detection  
1174 is concerned. These results are related to the presence of carboxyl functionalized  
1175 carbon nanotube in the sensitive layer which facilitates the transfer of the electrodes  
1176 involved in the electrochemical process.

1177 Histamine is a biogenic amine of low molecular weight, with biologic activity.  
1178 Histamine intoxication is also known as “scombroid fish poisoning”. Histamine  
1179 concentration is used as an indicator of fish spoilage (Luten, 2006; Feng, Teuber, &  
1180 Gershwin, 2016).

1181 Histamine is accumulated in seafood after the beginning of bacterial spoilage and  
1182 causes histamine poisoning even though the fish may not be altered in what the visual  
1183 aspect and smell is concerned (Luten, 2006; Feng, Teuber, & Gershwin, 2016).

1184 Keow et al. (2007) developed a biosensor based on diaminoxidase for the detection of  
1185 histamine in tiger prawn (*Penaeus monodon*) (Keow, Bakar, Salleh, Heng, Wagiran,  
1186 & Bean, 2007). The response time of the biosensor is below 1 minute under optimal  
1187 pH conditions of 7.4. The limit of detection is in the sub-ppm domain (under 50 ppm,  
1188 the level established by FDA USA), which recommends it for practical usage.

1189 For the validation of the biosensor on real samples, the variation of histamine  
1190 concentration was studied on tiger prawn samples after a 5-hour exposure at  $30 \pm 2^\circ\text{C}$   
1191 temperature. The results obtained were comparable to the results determined by  
1192 HPLC. There is good linear correlation between the two methods, with the  
1193 determination coefficient higher than 0.95. The biosensor is reusable and may be used  
1194 for the determination and quantification of histamine without further sample  
1195 processing, being appropriate for the analysis of histamine in tiger prawn and also for  
1196 spoilage monitoring.

1197 Bóka et al. (2012) developed a novel amperometric biosensor based on putrescine  
1198 oxidase for the selective detection and quantification of putrescine, a characteristic  
1199 which may function as an indicator of microbial spoilage (Bóka, Adányi, Szamos,  
1200 Virág, & Kiss, 2012). Putrescine oxidase was isolated from *Kocuria rosea*

1201 (*Micrococcus rubens*). The purified enzyme was immobilized onto the surface of a  
1202 graphite electrode in a hydrogel containing horseradish peroxidase, as a mediator of  
1203 electron transfer and poly (ethylene glycol) (400) diglycidyl ether as a reticular agent.

1204 This biosensor was used in an amperometric electrochemical cell in flow together  
1205 with the reference electrode Ag/AgCl (0.1 M KCl) and a platinum wire as an auxiliary  
1206 electrode. Under optimal conditions of pH, flow rate and applied potential, a vast  
1207 linearity domain was obtained between the response of the biosensor and the  
1208 putrescine concentration, with a detection limit appropriate for applications in foods  
1209 analysis. The validation of the biosensor was pursued by analysing beer samples and  
1210 comparing the results obtained with the results of the reference method HPLC.

1211 The formation of volatile compounds, such as acetaldehyde and ethylene in plants and  
1212 fruits is related to the state of their metabolism. For example, the synthesis speed of  
1213 ethylene in apples increases with the time spent after harvest, while the acetaldehyde  
1214 production is related to the anaerobic metabolism which grows in fruits after  
1215 harvesting. The quantity of ethylene and acetaldehyde is related to the metabolic state  
1216 and to the quality of fruit (Chen, Zhang, Hao, Chen, & Cheng, 2015; Maffei, 2010).

1217 Weber et al. (2009) developed and implemented a hybrid dual-channel catalytic-  
1218 biological sensor system, able to quantify the two volatile substances in situ (Weber,  
1219 Luzi, Karlsson, & Fussenegger, 2009). This biosensor is based on a mammalian cell  
1220 line engineered for constitutive expression of an *Aspergillus nidulans*, which triggers  
1221 quantitative reporter gene expression in the presence of acetaldehyde. Ethylene  
1222 oxidized to acetaldehyde through Wacker process can be quantified with the same  
1223 biosensor. The quantification of metabolites allowed the accurate assessment of the

1224 quality of fruits, the fresh apples being clearly differentiated from the old and rotten  
1225 apples.

1226 By placing in relation the catalytic processes and the detection technology of the  
1227 biosensors, it was possible to determine the metabolic state of food. Consequently,  
1228 this could be used in the assessment of foods which suffer biochemical  
1229 transformations, as well as in control processes for detecting and preventing food  
1230 spoilage (Zhang & Keasling, 2011).

1231 Fumarate is a very important intermediary in Krebs cycle (the tricarboxylic acid  
1232 cycle) and has a key role in the fundamental processes which produce energy, as well  
1233 as in the biosynthesis of amino acids and lipids (Nelson & Cox, 2017).

1234 The accumulation of fumarate in organism above a certain limit, due to fumarate  
1235 hydratase mutation, is one of the main causes of hereditary leiomyomatosis and renal  
1236 cell cancer, being considered an oncometabolite (Yang, Soga, Pollard, & Adam,  
1237 2012)

1238 On the other hand, fumarate is present in beverages, baking powders and candy, as a  
1239 result of the microbial activity which leads to spoilage. Another source of  
1240 contamination is represented by the impurities present in certain synthetic additives.  
1241 Accordingly, fumarate is an important and relevant indicator of food quality, which  
1242 can be used as a biomarker of food freshness (Hurrell, 2010; Kvasničk & Voldřich,  
1243 2000). Nevertheless, a cost-effective and fast analytical method for the detection and  
1244 quantification of fumarate is desired. Si et al. (2015) produced an electrochemical  
1245 whole-cell biosensing system for the quantification of fumarate in foods (apple juice)  
1246 (Si, Zhai, Liao, Gao, & Yong, 2015). A sensitive inwards electric output (electron  
1247 flow from electrode into bacteria) is sensitive to fumarate in *Shewanella oneidensis*

1248 MR-1. Therefore, the electrochemical fumarate biosensing system delivered  
1249 symmetric current peak immediately upon fumarate addition in the sample. The peak  
1250 area increases in direct ratio with fumarate concentration in vast concentration domain  
1251 with a limit of detection of 0.83  $\mu\text{M}$ . This biosensing system showed to be specific to  
1252 fumarate, as the interferences are very low. The validation of this biosensing system  
1253 was pursued by the successful quantification of fumarate in samples of apple juice.  
1254 The advantages of this biosensing system are: simplicity, low cost, limited time  
1255 required for analysis and its robustness in fumarate quantification.

1256

## 1257 **7. Challenges and future trends**

1258 Commercial electronic noses are designed for general-purpose use and besides  
1259 selectivity and sensitivity of the sensors in the array; they do not match the needs for a  
1260 particular application. It is necessary to design an array of sensors with optimized  
1261 conditions for each application in order to increase the performance for food spoilage  
1262 detection.

1263 So far, electronic noses as sensory detectors of food spoilage have been widely used  
1264 in the laboratory of different research groups. It is also clear that the utility of using  
1265 electronic noses in an industrial or consumer context is high; the chemical compounds  
1266 responsible of food spoilage are usually detected by electronic noses at lower  
1267 concentrations than human nose, so efforts must be made by researchers to transfer  
1268 this technology to them. For the food industry, faster and more efficient sampling  
1269 techniques suitable for successive batches need to be developed in the future. On the  
1270 sensors side, major focus must be given to the design and development of high  
1271 sensitivity and selectivity drift free sensors that can be used reliably over long

1272 temporal horizons. Novel and promising materials like grapheme or silicene should be  
1273 used for developing ambient temperature sensors and novel nanostructures like  
1274 nanowires and nanofibers and other nanostructures could enhance the response and  
1275 reduce the time of response and consumption. Data processing methods not only must  
1276 be made for classification and prediction problems, but also for sensor replacements,  
1277 compensating drift, stability and reliability of the sensors. It will allow a long-term  
1278 use that will be a convincing factor for industry when considering the uptake of such a  
1279 device. On the consumers' side, there are now available in the market miniature gas  
1280 sensors with low size (less than 2x3mm) and consumption (less than 7mw) that will  
1281 allow to develop very small electronic noses systems for consumers in order to advise  
1282 them if food they are going to consume is of adequate quality. Moreover, mobile  
1283 phones have been increasing the number of sensors they contain; from one or two  
1284 sensors in 2003 to more than 16 sensors in 2016. Predictions of the sensor market say  
1285 that in the near future, smart phones will include gas sensors, and with it hundreds of  
1286 apps for detecting compounds, odours and aromas related with food spoilage.

1287 The future of the electronic tongue systems and the biosensors are closely related  
1288 because improving the sensitivity and selectivity of the sensor array remain  
1289 challenging tasks.

1290 It seems that the trends will include the development of novel sensitive nanomaterials  
1291 and the nanotechnologies for the preparation of the sensors as well as the use of  
1292 hybrid array of sensors. The inclusion of the biosensors in the sensors arrays could be  
1293 a factor that will improve the multi-analyte detection, the quantitative analyses  
1294 becoming more significant and more precise. This is necessary in the detection of  
1295 food spoilage in early stage, when it starts and not when the food product is spoiled  
1296 and not suitable for human consumption. Other important research directions will

1297 include the miniaturization of the systems able to measure in-flow in real-time  
1298 analysis, coupled with wireless signal transmitters, expert systems for data analysis  
1299 and feed-back action. These multisensory systems will assure a rapid and accurate  
1300 control of food spoilage, important for the producers and for the consumers.

1301

## 1302 **8. Conclusion**

1303 In this paper, we have outlined the major contributions of electronic nose, biosensors,  
1304 and electronic tongue technologies related with food spoilage. There is a great interest  
1305 for handheld instruments that respond to simple questions related with food spoilage  
1306 posed by producers, food inspectors and general consumers. A great number of  
1307 references can be found with different applications of food spoilage detection,  
1308 including wine spoilage monitoring and detection of off-flavors, beer defects,  
1309 microbial contamination in tomatoes, egg quality detection, grain spoilage,  
1310 enterobacteriaceae in vegetable soups, spoilage of bakery products, contamination of  
1311 soft drinks, apple defects, milk spoilage and olive oil defects, fish freshness  
1312 monitoring, meats freshness, seafood spoilage, apple juice spoilage, among others.  
1313 Electronic noses and gas sensors have shown in the last years an important  
1314 enhancement in the time response and time life as well as a decrease in the size and  
1315 consumption. The latest works about the electronic tongue systems for detection of  
1316 food spoilage demonstrates one significant progress in the terms of high sensitive  
1317 sensor arrays based on different methods of detection and the use of improved data  
1318 analyses. The biosensors were used in the detection of target analytes related to food  
1319 spoilage with high sensitivity, improved selectivity, and low detection limit. These  
1320 superior analytical characteristics are principally related to the use of nanomaterials  
1321 and nanotechnologies in the development of biosensors.

1322

1323 **Acknowledgments**

1324 The supports of Shahrekord University (Shahrekord, Iran) and University of  
1325 Extremadura (Spain) are gratefully appreciated. Also, this work was supported in a  
1326 part by a grant of the Romanian National Authority for Scientific Research and  
1327 Innovation, CNCS - UEFISCDI, project number PN-II-RU-TE-2014-4-1093.

1328

1329 **References**

1330 Abdulbari, H. A., Basheer, E. A. M. (2017) Electrochemical Biosensors: Electrode  
1331 Development, Materials, Design, and Fabrication. *ChemBioEng Reviews* **4**, 92 – 105.  
1332 doi:10.1002/cben.201600009.

1333 Adley, C. C. (2014) Past, Present and Future of Sensors in Food Production. *Foods* **3**,  
1334 491 –510.

1335 Aguilera, T., Lozano, J., Paredes, J. A., Alvarez, F. J., & Suárez, J. I. (2012).  
1336 Electronic nose based on independent component analysis combined with partial least  
1337 squares and artificial neural arrays for wine prediction. *Sensors* (Basel, Switzerland),  
1338 12(6), 8055–72. <http://doi.org/10.3390/s120608055>

1339

1340 Ahn, S.R, An, J.H., Song, H.S., Park, J.W., Lee, S.H., Kim, J.H., Jang, J., Park, T.H.  
1341 (2016) Duplex Bioelectronic Tongue for Sensing Umami and Sweet Tastes Based on  
1342 Human Taste Receptor Nanovesicles. *ACS Nano* **10**, 7287 - 7296.

1343

- 1344 Ali, J., Najeeb, J., Ali, M.A., Aslam, M.F., Raza, A. (2017) Biosensors: Their  
1345 Fundamentals, Designs, Types and Most Recent Impactful Applications: A Review.  
1346 *Journal of Biosensors & Bioelectronics* **8**, 235. doi: 10.4172/2155-6210.1000235.  
1347
- 1348 Almeida Silva, T., Cruz Moraes, F., Campos Janegitz, B., Fatibello-Filho, O. (2017)  
1349 Electrochemical Biosensors Based on Nanostructured Carbon Black: A Review.  
1350 *Journal of Nanomaterials* **2017**, Article ID 4571614, 14 pages.
- 1351 Amine, A., Mohammadi, H., Bourais, I., Palleschi, G. (2006) Enzyme inhibition-  
1352 based biosensors for food safety and environmental monitoring. *Biosensors and*  
1353 *Bioelectronics* **21**, 1405 – 1423.  
1354
- 1355 Andre, S., Vallaey, T., Planchon, S. (2017) Spore-forming bacteria responsible for  
1356 food spoilage. *Research in Microbiology* **52**, 123-138.  
1357
- 1358 Apetrei, C., Ghasemi-Varnamkhasi, M. (2013) *Biosensors in food PDO*  
1359 *authentication*. *Comprehensive Analytical Chemistry*, Volume 60, 279 - 297.
- 1360 Apetrei, C., Rodríguez-Méndez, M. L., Parra, V., Gutierrez, F., de Saja, J.A. (2004)  
1361 Array of voltammetric sensors for the discrimination of bitter solutions. *Sensors and*  
1362 *Actuators B: Chemical* **103**, 145 – 152.  
1363
- 1364 Apetrei, I.M., Apetrei, C. (2014) Detection of virgin olive oil adulteration using a  
1365 voltammetric e-tongue. *Computers and Electronics in Agriculture* **108**, 148 – 154.
- 1366 Apetrei, I.M., Apetrei, C. (2015) The biocomposite screen-printed biosensor based on  
1367 immobilization of tyrosinase onto the carboxyl functionalised carbon nanotube for  
1368 assaying tyramine in fish products. *Journal of Food Engineering* **149**, 1 – 8.

1369

1370 Apetrei, I.M., Apetrei, C. (2016) Amperometric biosensor based on diamine  
1371 oxidase/platinum nanoparticles/graphene/chitosan modified screen-printed carbon  
1372 electrode for histamine detection. *Sensors* **16**, Article number 422, doi:  
1373 10.3390/s16040422.

1374 Apetrei, I.M., Apetrei, C. (2016) Application of voltammetric e-tongue for the  
1375 detection of ammonia and putrescine in beef products. *Sensors and Actuators B:  
1376 Chemical* **234**, 371 - 379.

1377

1378 Apetrei, I.M., Rodriguez-Mendez, M.L., Apetrei, C., de Saja, J.A. (2013) Fish  
1379 freshness monitoring using an E-tongue based on polypyrrole modified screen-printed  
1380 electrodes. *IEEE Sensors Journal* **13**, 2548 – 2554.

1381

1382 Arrieta, A., Rodriguez-Mendez, M. L., de Saja, J.A. (2003) Langmuir–Blodgett film  
1383 and carbon paste electrodes based on phthalocyanines as sensing units for taste.  
1384 *Sensors and Actuators B: Chemical* **95**, 357 – 365.

1385

1386 Arrieta, A.A. Apetrei, C., Rodríguez-Méndez, M. L., de Saja, J. A. (2004)  
1387 Voltammetric sensor array based on conducting polymer-modified electrodes for the  
1388 discrimination of liquids. *Electrochimica Acta* **49**, 4543 – 4551.

1389

1390 Barakat, R.K., Griffiths, M.W., Harris, L.J. (2000) Isolation and characterization of  
1391 *Carnobacterium*, *Lactococcus* and *Enterococcus* spp. from cooked, modified  
1392 atmosphere packaged, refrigerated, poultry meat. *International Journal of Food  
1393 Microbiology* **62**, 83 – 94.

1394

1395 Bard, A. J., Faulkner, L. R. (2001) *Electrochemical Methods: Fundamentals and*  
1396 *Applications*, 2<sup>nd</sup> Edition, John Willey & Sons, Inc., New York.

1397 Barrett, E.L., Kwan, H.S. (1985) Bacterial reduction of trimethylamine oxide. *Annual*  
1398 *Review of Microbiology* **39**, 131 - 149.

1399

1400 Bassi, A.S., Lee, E., Zhu J.-X. (1998) Carbon paste mediated, amperometric, thin film  
1401 biosensors for fructose monitoring in honey. *Food Research International* **31**, 119 –  
1402 127.

1403 Benabdellah, N. Bourhaleb, M., Benazzi, N., Nasri, M., Dahbi, S. 2017. The detection  
1404 of smell in spoiled meat by TGS822 gas sensor for an electronic nose used in rotten  
1405 food. *Advances in Intelligent Systems and Computing*. 520, 279-286.

1406 Berna, A. Z., Trowell, S., Cynkar, W., & Cozzolino, D. (2008). Comparison of metal  
1407 oxide-based electronic nose and mass spectrometry-based electronic nose for the  
1408 prediction of red wine spoilage. *Journal of Agricultural and Food Chemistry*, 56(9),  
1409 3238–3244. <http://doi.org/10.1021/jf7037289>

1410

1411 Blixt, Y., Borch, E. (1999) Using an electronic nose for determining the spoilage of  
1412 vacuum packaged beef. *Int J. Food. Microbiol.* 46, 123 -134.

1413

1414 Bobacka, J., Ivaska, A., Lewenstam, A. (2008) Potentiometric Ion Sensors. *Chemical*  
1415 *Reviews* **108**, 329 - 351.

1416

- 1417 Bóka, B., Adányi, N., Szamos, J., Virág, D., Kiss A. (2012) Putrescine biosensor  
1418 based on putrescine oxidase from *Kocuria rosea*. *Enzyme and Microbial Technology*  
1419 **51**, 258 – 262.
- 1420
- 1421 Bourigua, S., El Ichi, S., Korri-Yousoufi, H., Maaref, A., Dzyadevych, S., Jaffrezic  
1422 Renault, N. (2011) Electrochemical sensing of trimethylamine based on polypyrrole–  
1423 flavin-containing monooxygenase (FMO3) and ferrocene as redox probe for  
1424 evaluation of fish freshness. *Biosensors and Bioelectronics* 28, 105-111.
- 1425
- 1426 Bratov, A., Abramova, N., Ipatov A. (2010) Recent trends in potentiometric sensor  
1427 arrays—A review. *Analytica Chimica Acta* **678**, 149 – 159.
- 1428
- 1429 Brett, C.M.A., Fungaro, D. A. (2000) Modified Electrode Voltammetric Sensors for  
1430 Trace Metals in Environmental Samples. *Journal of the Brazilian Chemical Society*  
1431 **11**, 298 – 303.
- 1432
- 1433 Brightwell, G., Clemens, R., Ulrich, S., Boerema, J. (2007) Possible involvement of  
1434 psychrotolerant *Enterobacteriaceae* in blown pack spoilage of vacuum-packaged raw  
1435 meats. *International Journal of Food Microbiology* **119**, 334 – 339.
- 1436
- 1437 Bueno, L., de Araujo, W. R., Salles, M. O., Kussuda, M. Y., Paixão, T. R. L. C.  
1438 (2014) Voltammetric Electronic Tongue for Discrimination of Milk Adulterated with  
1439 Urea, Formaldehyde and Melamine. *Chemosensors* **2**, 251-266.
- 1440

- 1441 Cabañes, F. J., Sahgal, N., Bragulat, M. R., & Magan, N. (2009). Early discrimination  
1442 of fungal species responsible of ochratoxin A contamination of wine and other grape  
1443 products using an electronic nose. *Mycotoxin Research*, 25(4), 187–192.  
1444 <http://doi.org/10.1007/s12550-009-0027-x>.
- 1445
- 1446 Cabral, F.P., Bergamo, B.B., Dantas, C.A., Riul Jr., A., Giacometti, J.A. (2009)  
1447 Impedance e-tongue instrument for rapid liquid assessment. *Review of Scientific*  
1448 *Instruments* **80**, 026107. doi: 10.1063/1.3084210.
- 1449 Campos Sánchez, I., Bataller Prats, R., Gandía Romero, J.M., Soto Camino, J.,  
1450 Martínez Mañez, R., Gil Sánchez, L. (2013) Monitoring grape ripeness using a  
1451 voltammetric electronic tongue. *Food Research International* **54**, 1369 - 1375.
- 1452
- 1453 Campos, I., Alcañiz, M., Aguado, D., Barat, R., Ferrer, J., Gil, L., Marrakchi, M.,  
1454 Martínez-Mañez, R., Soto, J., Vivancos, J.-L. (2012) A voltammetric electronic  
1455 tongue as tool for water quality monitoring in wastewater treatment plants. *Water*  
1456 *Research* **46**, 2605 – 2614.
- 1457
- 1458 Campuzano, S., Ruiz-Valdepeñas Montiel, V., Torrente-Rodríguez, R. M., Reviejo,  
1459 Á. J., Pingarrón, J. M. (2016) *Electrochemical Biosensors for Food Security:*  
1460 *Allergens and Adulterants Detection*, 287-307. In book: *Biosensors for Security and*  
1461 *Bioterrorism Applications*. Springer.
- 1462
- 1463 Casaburi, A., Nasi, A., Ferrochino, L., Di Monaco, R., Mauriello, G., Villiani, F.,  
1464 Ercolini, D. (2011) Spoilage-related activity of *Carnobacterium maltaromaticum*

1465 strains in air stored and vacuum packed meat. *Applied and Environmental*  
1466 *Microbiology* **77**, 7382 - 7393.

1467

1468 Casaburi, A., Piombino, P., Nychas, G-J., Villiani, F. (2015) Bacterial populations  
1469 and the volatilome associated to meat spoilage. *Food Microbiology* **45**, 83 – 102.

1470

1471 Cetó, X., Apetrei, C., Del Valle, M., Rodríguez-Méndez, M.L. (2014) Evaluation of  
1472 red wines antioxidant capacity by means of a voltammetric e-tongue with an  
1473 optimized sensor array. *Electrochimica Acta* **120**, 180 – 186.

1474

1475 Cetó, X., Capdevila, J., Puig, A., del Valle, M. (2014) Cava wine authentication  
1476 employing a voltammetric electronic tongue. *Electroanalysis* **26**, 1504 - 1512.

1477

1478 Cetó, X., González-Calabuig, A., del Valle, M. (2015) Use of a Bioelectronic Tongue  
1479 for the Monitoring of the Photodegradation of Phenolic Compounds. *Electroanalysis*  
1480 **27**, 225 – 233.

1481

1482 Cetó, X., Voelcker, N. H., Prieto-Simón, B. (2016) Bioelectronic tongues: New trends  
1483 and applications in water and food analysis. *Biosensors and Bioelectronics* **79**, 608 –  
1484 626.

1485

1486 Chatterjee, D., Bhattacharjee, P., & Bhattacharyya, N. (2014). Development of  
1487 methodology for assessment of shelf-life of fried potato wedges using electronic

- 1488 noses: Sensor screening by fuzzy logic analysis. *Journal of Food Engineering*, 133,  
1489 23–29. <http://doi.org/10.1016/j.jfoodeng.2014.02.009>
- 1490
- 1491 Chauhan, N., Maekawa, T., Kumar, D. (2017) Graphene based biosensors—  
1492 Accelerating medical diagnostics to new-dimensions. *Journal of Materials Research*,  
1493 1 - 23. doi:10.1557/jmr.2017.91
- 1494
- 1495 Chen, S., Zhang, R., Hao, L., Chen, W., Cheng, S. (2015) Profiling of Volatile  
1496 Compounds and Associated Gene Expression and Enzyme Activity during Fruit  
1497 Development in Two Cucumber Cultivars. *PLoS One* **10**, e0119444. doi:  
1498 10.1371/journal.pone.0119444
- 1499
- 1500 Chung, S., Park, T. S., Park, S. H., Kim, J. Y., Park, S., Son, D., Bae, Y. M., Cho S. I.  
1501 (2015) Colorimetric Sensor Array for White Wine Tasting. *Sensors* **15**, 18197 -  
1502 18208.
- 1503
- 1504 Ciosek, P., Wróblewski W. (2007) Sensor arrays for liquid sensing – electronic  
1505 tongue systems. *Analyst* **132**, 963 – 978.
- 1506
- 1507 Ciosek, P., Wróblewski, W. (2011) Potentiometric Electronic Tongues for Foodstuff  
1508 and Biosample Recognition—An Overview. *Sensors* **11**, 4688 - 4701.
- 1509
- 1510 Cleto, S., Matos, S., Kluskens, L., Vieira, M.J. (2012) Characterization of  
1511 contaminants from a sanitized milk processing plant. *PLoS One*. 7, e40189.

1512

1513 Compagnone, D., Di Francia, G., Di Natale, C., Neri, G., Seeber, R., Tajani A. (2017)  
1514 Chemical Sensors and Biosensors in Italy: A Review of the 2015 Literature. *Sensors*  
1515 **17**, 868; doi:10.3390/s17040868.

1516

1517 Concina, I., Bornšek, M., Baccelliere, S., Falasconi, M., Gobbi, E., & Sberveglieri, G.  
1518 (2010). Alicyclobacillus spp.: Detection in soft drinks by Electronic Nose. *Food*  
1519 *Research International*, **43**(8), 2108–2114.  
1520 <http://doi.org/10.1016/j.foodres.2010.07.012>

1521

1522 Concina, I., Falasconi, M., Gobbi, E., Bianchi, F., Musci, M., Mattarozzi, M., Pardo,  
1523 M., Mangia, A., Careri, M., Sberveglieri, G., (2009) Early detection of microbial  
1524 contamination in processed tomatoes by electronic nose. *Food Control*, **20**, 873-880.

1525

1526 Costa, C., Miertus, S. (1993) *Trends in Electrochemical Biosensors: Proceedings of*  
1527 *the Conference*. World Scientific, Singapore.

1528

1529 Cuartero, M., Carretero, A., Garcia, M. S., Ortuño, J. A. (2015) New Potentiometric  
1530 Electronic Tongue for Analysing Teas and Infusions. *Electroanalysis* **27**, 782 – 788.

1531

1532 Cynkar, W., Cozzolino, D., Damberg, B., Janik, L., & Gishen, M. (2007). Feasibility  
1533 study on the use of a head space mass spectrometry electronic nose (MS e\_nose) to  
1534 monitor red wine spoilage induced by Brettanomyces yeast. *Sensors and Actuators, B:*  
1535 *Chemical*, **124**(1), 167–171. <http://doi.org/10.1016/j.snb.2006.12.017>

1536

1537 Dainty, R.H., Mackey, B.M., (1992) The relationship between phenotypic properties  
1538 of bacteria from chill-stored meat and spoilage processes. *Society of Applied*  
1539 *Bacteriology Symposium Series* **73**, 103S – 114S.

1540

1541 Dalgaard, P. (1995) Qualitative and quantitative characterization of spoilage bacteria  
1542 from packed fish. *International Journal of Food Microbiology* **26**, 319 – 333.

1543

1544 Dalgaard, P., Madsen, H., Samieian, N., Emborg, J. (2006) Biogenic amine formation  
1545 and microbial spoilage in chilled garfish (*Belone belone*) effect of modified  
1546 atmosphere packaging and previous frozen storage. *Journal of Applied Microbiology*  
1547 **101**, 80 – 95.

1548

1549 Dashdorj, D., Tripathi, V. K., Cho, S., Kim, Y., Hwang, I. (2016) Dry aging of beef.  
1550 *Journal of Animal Science and Technology* **58**: 20. doi: 10.1186/s40781-016-0101-9

1551 Dave, D., Ghaly A. E. (2011). Meat Spoilage Mechanisms and Preservation  
1552 Techniques: A Critical Review. *American Journal of Agricultural and Biological*  
1553 *Sciences* **6**, 486-510.

1554

1555 de Blackburn, C. (2006) *Food Spoilage Microorganisms*, 1<sup>st</sup> Edition, Elsevier.

1556

1557 Del Rio, E., Panizo-Moran, M., Prieto, M., Alonso-Calleja C., Capita, R. (2007)  
1558 Effect of various chemical decontamination treatments on natural microflora and

- 1559 sensory characteristics of poultry. *International Journal of Food Microbiology* **76**,  
1560 201 – 207.
- 1561
- 1562 del Valle M. (2010) Electronic Tongues Employing Electrochemical Sensors.  
1563 *Electroanalysis* **22**, 1539 – 1555.
- 1564 del Valle, M. (2012) Sensor Arrays and Electronic Tongue Systems. *International*  
1565 *Journal of Electrochemistry* **2012**, Article ID 986025, 11 pages.  
1566 <http://dx.doi.org/10.1155/2012/986025>
- 1567 del Valle, M. (2012) Sensor Arrays and Electronic Tongue Systems. *International*  
1568 *Journal of Electrochemistry* **2012**, Article ID 986025, 11 pages.  
1569 doi:10.1155/2012/986025
- 1570 del Valle, M., Cetó, X., Gutierrez-Capitán, M. (2014) *BioElectronic tongues: When*  
1571 *the sensor array incorporate biosensors*, pp. 211-246, in *Multisensor Systems for*  
1572 *Chemical Analysis: Materials and Sensors*. Lvova, L., Kirsanov, D., Di Natale, C.,  
1573 Legin, A. (Eds.) CRC Press.
- 1574 Dervisevic, M., Custiuc, E., Çevik, E., Durmus, Z., Senel, M., Durmus, A. (2015)  
1575 Electrochemical biosensor based on REGO/Fe<sub>3</sub>O<sub>4</sub> bionanocomposite interface for  
1576 xanthine detection in fish sample. *Food Control* **57**, 402 – 410.
- 1577 Dervisevic, M., Custiuc, E., Çevik, E., Senel, M. (2015) Construction of novel  
1578 xanthine biosensor by using polymeric mediator/MWCNT nanocomposite layer for  
1579 fish freshness detection. *Food Chemistry* **181**, 277 – 283.
- 1580 Di Rosa, A. R., Leone, F., Cheli, F., Chiofalo V. (2017) Fusion of electronic nose,  
1581 electronic tongue and computer vision for animal source food authentication and  
1582 quality assessment - A review. *Journal of Food Engineering* **210**, 62 – 75.

- 1583 Dias, L. G., Veloso, A. C.A., Sousa, M. E. B. C., Estevinho, L., Machado, A. A. S. C.,  
1584 Peres, A. M. (2015) A novel approach for honey pollen profile assessment using an  
1585 electronic tongue and chemometric tools. *Analytica Chimica Acta* **900**, 36 – 45.
- 1586 Dikeman, M., Devine C., (Eds) (2014) *Encyclopedia of Meat Sciences*, 2<sup>nd</sup> edition,  
1587 Elsevier.
- 1588 Domínguez, R. B., Moreno-Barón, L., Muñoz, R., Gutiérrez, J. M. (2014)  
1589 Voltammetric Electronic Tongue and Support Vector Machines for Identification of  
1590 Selected Features in Mexican Coffee. *Sensors* **14**, 17770 – 17785.
- 1591 Doulgeracki, A.I., Ercolini, D., Villani, F., Nychas, G.J. (2012) Spoilage microbiota  
1592 associated to the storage of raw meat in different conditions. *International Journal of*  
1593 *Food Microbiology* **157**, 130-141.
- 1594 Doyle, M.E. (2007) Microbial Food Spoilage – Losses and Control Strategies: a brief  
1595 review of the literature. Food Research Institute. Briefings. University of Wisconsin  
1596 – Madison. [www.wisc.edu/fri](http://www.wisc.edu/fri).
- 1597 Dragone, R., Grasso, G., Muccini, M., Toffanin, S. (2017) Portable  
1598 Bio/Chemosensoristic Devices: Innovative Systems for Environmental Health and  
1599 Food Safety Diagnostics. *Frontiers in Public Health* **5**, 80. doi:  
1600 10.3389/fpubh.2017.00080
- 1601 dry-cured ham, *J. Food Sci.* 75 M360–M365.
- 1602 Eckert, C., Pein, M., Reimann, J., Breitzkreutz, J. (2013) Taste evaluation of  
1603 multicomponent mixtures using a human taste panel, electronic taste sensing systems  
1604 and HPLC. *Sensors and Actuators B: Chemical* **182**, 294-299.

- 1605 El Barbri, N., Llobet, E., El Bari, N., Correig, X., and Bouchikhi, B., 2008. Electronic  
1606 Nose Based on Metal Oxide Semiconductor Sensors as an Alternative Technique for  
1607 the Spoilage Classification of Red Meat. *Sensors*, 8, 142-156
- 1608 El-Nour, K. M. A., Salam, E. T. A., Soliman, H. M., Orabi A. S. (2017) Gold  
1609 Nanoparticles as a Direct and Rapid Sensor for Sensitive Analytical Detection of  
1610 Biogenic Amines. *Nanoscale Research Letters* **12**, 231. DOI 10.1186/s11671-017-  
1611 2014-z
- 1612 Ercolini, D., Russo, F., Nasi, A., Ferranti, P., Villiani, F. (2009) Mesophilic and  
1613 psychrotrophic bacteria from meat and their spoilage potential in vitro and in beef.  
1614 *Applied and Environmental Microbiology* **75**, 1990-2001.
- 1615 Ercolini, D., Russo, F., Torrieri, E., Masi, P., Villani, F. (2006) Changes in the  
1616 spoilage –related microbiota of beef during refrigerated storage under different  
1617 packaging conditions. *Applied and Environmental Microbiology* **72**, 4663-4671.
- 1618 Esposto, S., Servili, M., Selvaggini, R., Ricc, I., San, V., & Perugia, C. (2006).  
1619 Discrimination of virgin olive oil defects - comparison of two evaluation methods :  
1620 HS-SPME GC-MS and electronic nose, 315–318.
- 1621 Estelles-Lopez et al., 2017. An automated ranking platform for machine learning  
1622 regression models for meat spoilage prediction using multi-spectral imaging and  
1623 metabolic profiling. *Food Research International*, In press,  
1624 <http://dx.doi.org/10.1016/j.foodres.2017.05.013>
- 1625 FAO. (2011) Global food losses and food waste – Extent, causes and prevention.  
1626
- 1627 Feng, C., Teuber, S., Gershwin, M.E. (2016) Histamine (Scombroid) Fish Poisoning:  
1628 a Comprehensive Review. *Clinical Reviews in Allergy & Immunology* **50**, 64-69.

1629

1630 Fonnesbech Vogel, B., Venkateswaran, K., Satomi, M., Gram, L. (2005)

1631 Identification of *Shewanella baltica* as the most important H<sub>2</sub>S-producing species1632 during iced storage of Danish marine fish. *Appl Environ Microbiol.* 71, 6689-97.

1633

1634 Frasco, M. F., Truta, L. A. A. N. A., Sales, M. G. F., Moreira, F. T. C. (2017)

1635 Imprinting Technology in Electrochemical Biomimetic Sensors. *Sensors* **17**, 523.

1636 doi:10.3390/s17030523

1637 Gamati, S., Luong, J. H. T., Mulchandani, A. (1991) A Microbial Biosensor for

1638 Trimethylamine Using *Pseudomonas aminovorans* Cells. *Biosensors and*1639 *Bioelectronics* **6**, 125-131

1640 Gancarz, M., Wawrzyniak, J., Gawrysiak-Witulska, M., Wiącek, D., Nawrocka, A.,

1641 Tadla, M., &amp; Rusinek, R. (2017). Application of electronic nose with MOS sensors to

1642 prediction of rapeseed quality. *Measurement*, 103, 227–234.1643 <http://doi.org/10.1016/j.measurement.2017.02.042>

1644 Ganjali, M. R., Nejad, F. G, Beitollahi, H., Jahani, S., Rezapour, M., Larijani B.

1645 (2017) Highly Sensitive Voltammetric Sensor for Determination of Ascorbic Acid

1646 Using Graphite Screen Printed Electrode Modified with ZnO/Al<sub>2</sub>O<sub>3</sub> Nanocomposite.1647 *International Journal of Electrochemical Science* **12**, 3231 – 3240.1648 Gardner, J. W., & Bartlett, P. N. (1994). A brief history of electronic noses. *Sensors*1649 and Actuators B, 19, 18–19. [http://doi.org/10.1016/0925-4005\(94\)87085-3](http://doi.org/10.1016/0925-4005(94)87085-3)

1650 Gerstl, M., Joksch, M., Fafilek, G. (2013) Designing a Simple Electronic Tongue for

1651 Fermentation Monitoring. *Journal of Analytical & Bioanalytical Techniques* **S12**:

1652 002. doi: 10.4172/2155-9872.S12-002

- 1653 Ghaly, A.E., Dave, D., Budge, S., Brooks, M.S. (2010) Fish Spoilage Mechanisms  
1654 and Preservation Techniques: Review. *American Journal of Applied Sciences* **7**, 859 -  
1655 877.
- 1656 Ghanbari, M., Ja,I, M., Domig, K.J., Kneifel, W. (2013) Seafood biopreservation by  
1657 lactic acid bacteria – A review. *LWT Food Science and Technology* **54**, 315 – 324.
- 1658 Ghasemi-Varnamkhasti M, Aghbashlo M (2014) Electronic nose and electronic  
1659 mucosa as innovative instruments for real time monitoring of food dryers Trends in  
1660 Food Science & Technology 38:158-166.
- 1661 Ghasemi-Varnamkhasti M, Mohtasebi S, Rodriguez-Mendez M, Lozano J, Razavi S,  
1662 Ahmadi H, Apetrei C (2012) Classification of non-alcoholic beer based on aftertaste  
1663 sensory evaluation by chemometric tools. *Expert Systems with Applications* 39,4315-  
1664 4327.
- 1665 Ghasemi-Varnamkhasti, M., Rodríguez-Méndez, M. L., Mohtasebi, S. S., Apetrei, C.,  
1666 Lozano, J., Ahmadi, H., Razavi, S. H., de Saja, J. A. (2012) Monitoring the aging of  
1667 beers using a bioelectronic tongue. *Food Control* **25**, 216-224.
- 1668 Gil, L., Barat, J. M., Baigts, D., Martínez-Mañez, R., Soto, J., Garcia-Breijo, E.,  
1669 Aristoy, M. C., Toldrá, F., Llobet E. (2011) Monitoring of physical–chemical and  
1670 microbiological changes in fresh pork meat under cold storage by means of a  
1671 potentiometric electronic tongue. *Food Chemistry* **126**, 1261 – 1268.
- 1672 Gil-Sanchez, L., Soto, J., Martinez-Mañez, R., Garcia-Breijo, E., Ibañez, J., & Llobet,  
1673 E. (2011). A novel humid electronic nose combined with an electronic tongue for  
1674 assessing deterioration of wine. *Sensors and Actuators, A: Physical*, 171(2), 152–158.  
1675 <http://doi.org/10.1016/j.sna.2011.08.006>.

- 1676 Gil-Sánchez, L., Soto, J., Martínez-Máñez, R., Garcia-Breijo, E., Ibáñez, J., Llobet, E.  
1677 (2011) A novel humid electronic nose combined with an electronic tongue for  
1678 assessing deterioration of wine. *Sensors and Actuators A* **171**, 152 – 158.
- 1679 Girón, J., Gil-Sánchez, L., García-Breijo, E., Pagána, M. J., Barat, J. M., Grau, R.  
1680 (2015) Development of potentiometric equipment for the identification of altered dry-  
1681 cured hams: A preliminary study. *Meat Science* **106**, 1 – 5.
- 1682 Gobbi, E., Falasconi, M., Concina, I., Mantero, G., Bianchi, F., Mattarozzi, M., ...  
1683 Sberveglieri, G. (2010). Electronic nose and Alicyclobacillus spp. spoilage of fruit  
1684 juices: An emerging diagnostic tool. *Food Control*, 21(10), 1374–1382.  
1685 <http://doi.org/10.1016/j.foodcont.2010.04.011>
- 1686 Gobbi, E., Falasconi, M., Concina, I., Mantero, G., Bianchi, F., Mattarozzi, M.,  
1687 Musci, M., Sberveglieri, G., 2010. Electronic nose and Alicyclobacillus spp. spoilage  
1688 of fruit juices: an emerging diagnostic tool. *Food Control* 21, 1374–1382.
- 1689 Gobbi, E., Falasconi, M., Zambotti, G., Sberveglieri, V., Pulvirenti, A., &  
1690 Sberveglieri, G. (2015). Rapid diagnosis of Enterobacteriaceae in vegetable soups by  
1691 a metal oxide sensor based electronic nose. *Sensors and Actuators, B: Chemical*,  
1692 207(PB), 1104–1113. <http://doi.org/10.1016/j.snb.2014.10.051>
- 1693 Godziszewska, J., et al., 2017. A simple method of the detection of pork spoilage  
1694 caused by *Rahnella aquatilis*. *LWT, IN press*, DOI; 10.1016/j.lwt.2017.05.049.
- 1695 Gorton L. (2005) *Biosensors and Modern Biospecific Analytical Techniques*. Elsevier.
- 1696 Gram, L., Ravn, L., Rasch, M., Bruhn, J.B., Christensen, A.B., Givskov, M. (2002)  
1697 Food spoilage – interactions between food spoilage bacteria. *International Journal of*  
1698 *Food Microbiology* **78**, 79-97.

- 1699 Grumezescu A. (Ed.) (2016) *Nanobiosensors*, Volume 8, 1<sup>st</sup> Edition. Academic Press,  
1700 Amsterdam.
- 1701 Gu, X., Sun, Y., Tu, K., & Pan, L., 2017. Evaluation of lipid oxidation of Chinese-  
1702 style sausage during processing and storage based on electronic nose. *Meat Science*,  
1703 133, 1–9
- 1704 Guadarrama, A., Fernández, J. A., Íñiguez, M., Souto, J., & De Saja, J. A. (2000).  
1705 Array of conducting polymer sensors for the characterisation of wines. *Analytica*  
1706 *Chimica Acta*, 411(1–2), 193–200. [http://doi.org/10.1016/S0003-2670\(00\)00769-8](http://doi.org/10.1016/S0003-2670(00)00769-8)
- 1707 Gul, I., Sheeraz Ahmad, M., Saqlan Naqvi, S. M., Hussain, A., Wali, R., Farooqi, A.  
1708 A., Ahmed, I. (2017) Polyphenol oxidase (PPO) based biosensors for detection of  
1709 phenolic compounds: A Review. *Journal of Applied Biology & Biotechnology* **5**, 072  
1710 – 085.
- 1711 Guo, X.S., Chen, Y.Q., Yang, X.L., Wang, L.R. (2005) Development of a novel  
1712 electronic tongue system using sensor array based on polymer films for liquid phase  
1713 testing. *2005 IEEE Engineering in Medicine and Biology*, 27<sup>th</sup> Annual Conference,  
1714 Shanghai, 259 - 262.
- 1715 Gupta, V.K., Jain, R., Radhapyari, K., Jadon, N., Agarwal, S. (2011) Voltammetric  
1716 techniques for the assay of pharmaceuticals-a review. *Analytical Biochemistry* **408**,  
1717 179 - 96.
- 1718 Gutiérrez, J. M., Haddi, Z., Amari, A., Bouchikhi, B., Mimendia, A., Cetó, X., del  
1719 Valle, M. (2013) Hybrid electronic tongue based on multisensor data fusion for  
1720 discrimination of beers. *Sensors and Actuators B: Chemical* **177**, 989 – 996.
- 1721 Gutiérrez, M., Llobera, A., Vila-Planas, J., Capdevila, F., Demming, S., Büttgenbach,  
1722 S., Mínguez, S., Jiménez-Jorquera, C. (2010) Hybrid electronic tongue based on

- 1723 optical and electrochemical microsensors for quality control of wine. *Analyst* **135**:  
1724 1718 - 1725.
- 1725 Gutiérrez-Capitán, M., Vila-Planas, J., Llobera, A., Jiménez-Jorquera, C., Capdevila,  
1726 F., Domingo, C., Puig-Pujol, A. (2014) Hybrid electronic tongues based on  
1727 microsensors applied to wine quality control. *IEEE Sensors 2014 Proceedings*,  
1728 Valencia, 2130 - 2133.
- 1729 Haddi, Z., El Barbri, N., Tahri, K., Bougrini, M., El Bari, N., Llobet, E., Bouchikhi,  
1730 B. (2015) Instrumental assessment of red meat origins and their storage time using  
1731 electronic sensing systems. *Analytical Methods* **7**, 5193 – 5203.
- 1732 Hasan, A., Nurunnabi, M., Morshed, M., Paul, A., Polini, A., Kuila, T., Al Hariri, M.,  
1733 Lee, Y., Jaffa, A. A. (2014) Recent Advances in Application of Biosensors in Tissue  
1734 Engineering. *BioMed Research International* **2014**, Article ID 307519, 18 pages.
- 1735 Haugen, J.E., 2006. Rapid control of smoked Atlantic salmon (*Salmo salar*) quality by  
1736 electronic nose: Correlation with classical evaluation methods. *Sensors and Actuators*  
1737 *B* **116**, 72–77.
- 1738 Hayashi, K., Yamanaka, M., Toko, K., Yamafuji, K. (1990) Multi-channel taste  
1739 sensor using lipid membranes. *Sensors and Actuators B: Chemical* **2**, 205 – 213.
- 1740 Hernandez-Macedo, M.L., Contreras-Castillo, C.J., Tsai, S.M., Da Cruz, S.H.,  
1741 Sarantoupoulos, C.I.G.L., Padula, M., Dias, C.T.S. (2012) Gases and volatile  
1742 compounds associated with microorganisms in blown pack spoilage of Brazilian  
1743 vacuum-packed beef. *Letters in Applied Microbiology* **55**, 467-475.
- 1744 <http://www.fao.org/docrep/014/mb060e/mb060e.pdf>
- 1745 Huang, X. C., Guo, C. F., Yuan, Y. H., Luo, X. X., & Yue, T. L. (2015). Detection of  
1746 medicinal off-flavor in apple juice with artificial sensing system and comparison with

- 1747 test panel evaluation and GC-MS. Food Control.  
1748 <http://doi.org/10.1016/j.foodcont.2014.11.037>
- 1749 Huis int. Veld., J.H.J. (1996) Microbial and biochemical spoilage of food; an  
1750 overview. *International Journal of Food Microbiology* **33**, 1-18.
- 1751 Hungaro, H.M., Caturla, M.Y.R., Horita, C.N., Furtado, M.M. Sant Ana, A.S. (2016)  
1752 Blown pack spoilage in vacuum-packaged meat: a review on clostridia as causative  
1753 agents, sources, detection methods, contributing factors and mitigation strategies.  
1754 *Trends in Food Science and Technology*, **52**, 123 -138.
- 1755 Hurrell, R. (2010) Use of ferrous fumarate to fortify foods for infants and young  
1756 children. *Nutrition Reviews* **68**, 522-530.
- 1757 Iiyama, S., Miyazaki, Y., Hayashi, K., Toko, K., Yamafuji, K., Ikezaki, H., Sato, K.  
1758 (1992) Highly sensitive detection of taste substances using monolayer lipid  
1759 membranes. *Sensors Materials* **4**, 21 – 27.
- 1760 Immohr, L. I., Hedfeld, C., Lang, A., Pein, M. (2017) Suitability of e-tongue sensors  
1761 to assess taste-masking of pediatric liquids by different beverages considering their  
1762 physico-chemical properties. *AAPS PharmSciTech* **18**, 330 – 340.
- 1763 Ispas, C. R., Crivat, G., Andreescu, S. (2012) Review: Recent Developments in  
1764 Enzyme-Based Biosensors for Biomedical Analysis. *Analytical Letters* **45**, 168 – 186.
- 1765 Iulietto, M. F., Sechi, P., Borgogni, E., Cenci-Goga, B. T. (2015) Meat Spoilage: A  
1766 Critical Review of a Neglected Alteration Due to Ropy Slime Producing Bacteria.  
1767 *Italian Journal of Animal Science* **14**, Article 4011,  
1768 <http://dx.doi.org/10.4081/ijas.2015.4011>  
1769

- 1770 Jaaskelainen, E., Hultman, J., Parshintsev, J., Riekkola, M-L., Bjorkroth, J. (2016)  
1771 Development of spoilage bacterial community and volatile compounds in chilled beef  
1772 under vacuum or high oxygen atmospheres. *International Journal of Food*  
1773 *Microbiology* **223**, 25-32.
- 1774 Jaffres, E., Lalanne, V., Mace, S., Cornet, J., Cardinal, M., Serot, T., Dousset, X.  
1775 Joffraud, J.J., (2011) Sensory characteristics of spoilage and volatile compounds  
1776 associated with bacteria isolated from cooked and peeled tropical shrimps using  
1777 SPME-GC-MS analysis. *International Journal of Food Microbiology* **147**, 195 - 202.
- 1778 Jaffres, E., Sohier, D., Leroi, F., Pilet, M.F., Prevost, H., Joffraud, J.J., Dousset, X.  
1779 (2009) Study of the bacterial ecosystem in tropical cooked and peeled shrimps using a  
1780 polyphasic approach. *International Journal of Food Microbiology* **131**, 20-29.
- 1781 Jain, H., Panchal, R., Pradhan, P., Patel, H., Pasha, T. (2010) Electronic tongue: A  
1782 new taste sensor. *International Journal of Pharmaceutical Sciences Review and*  
1783 *Research* **5**, 91 – 96.
- 1784 Jay, J.M. (1986) Microbial spoilage indicators and metabolites. In: Pierson, M.D.,  
1785 Sterm, N.J. (Eds.) *Food-Borne Microorganisms and their toxins: Developing*  
1786 *Methodology*, Marcel Dekker, Inc., New York, N.Y., pp. 219-240.
- 1787 Jorgenson, L.V., Huss, H.H, Dalgaard, P. (2000) The effect of biogenic amine  
1788 production by single bacterial cultures and metabiosis on cold-smoked salmon.  
1789 *Journal of Applied Microbiology* **89**, 920 – 934.
- 1790 Kalit, M. T., Marković, K., Kalit, S., Vahčić, N., Havranek, J. (2014) Electronic nose  
1791 and electronic tongue in the dairy industry. *Mljekarstvo* **64**, 228 – 244.

- 1792 Kaneki, N., Miura, T., Shimada, K., Tanaka, H., Ito, S., Hotori, K., Akasaka, C.,  
1793 Ohkubo, S., Asano, Y. (2004) Measurement of pork freshness using potentiometric  
1794 sensor. *Talanta* **62**, 217 - 221.
- 1795 Kangas, M. J., Burks, R. M., Atwater, J., Lukowicz, R. M., Williams, P., Holmes, A.  
1796 E. (2017) Colorimetric Sensor Arrays for the Detection and Identification of Chemical  
1797 Weapons and Explosives. *Critical Reviews in Analytical Chemistry* **47**, 138 – 153.
- 1798 Karovičová, J., Kohajdová, Z. (2005) Biogenic Amines in Food. *Chemical Papers* **59**,  
1799 70 – 79.
- 1800 Keow, C. M., Bakar, F. A, Salleh, A. B., Heng, L. Y., Wagiran, R., Bean, L. S.  
1801 (2007) An amperometric biosensor for the rapid assessment of histamine level in tiger  
1802 prawn (*Penaeus monodon*) spoilage. *Food Chemistry* **105**, 1636 – 1641.
- 1803 Khan, M. R. R., Khalilian, A., Kang S. W. (2016) A High Sensitivity IDC-Electronic  
1804 Tongue Using Dielectric/Sensing Membranes with Solvatochromic Dyes. *Sensors* **16**,  
1805 668. doi:10.3390/s16050668
- 1806 Khan, M. R. R., Khalilian, A., Kang, S.-W. (2016) A High Sensitivity IDC-Electronic  
1807 Tongue Using Dielectric/Sensing Membranes with Solvatochromic Dyes. *Sensors* **16**,  
1808 668. doi: 10.3390/s16050668.
- 1809 Khulal, U., Zhao, J., Hu, W., Chen, Q., 2017. Intelligent evaluation of total volatile  
1810 basic nitrogen (TVB-N) content in chicken meat by an improved multiple level data  
1811 fusion model. *Sensors and Actuators B* **238** (2017) 337–345
- 1812 Kiani S, Minaei S, Ghasemi-Varnamkhasti M (2016) Fusion of artificial senses as a  
1813 robust approach to food quality assessment *Journal of Food Engineering* **171**:230-239.
- 1814 Kodogiannis, V.S., 2017. Application of an Electronic Nose Coupled with Fuzzy-  
1815 Wavelet Network for the Detection of Meat Spoilage. *Food Bioprocess Technology*,  
1816 10,730–749.

- 1817 Korkeala, H., Suortti, T., MäkeläRopy, P. (1998) Slime formation in vacuum-packed  
1818 cooked meat products caused by homofermentative lactobacilli and a *Leuconostoc*  
1819 species. *International Journal of Food Microbiology*, 7, 339-347.
- 1820 Koutsoumanis, K., Nychas, G.J.E. (2000) Application of a systematic experimental  
1821 procedure to develop a microbial model for rapid fish shelf-life predictions.  
1822 *International Journal of Food Microbiology*, 60, 171 -174.
- 1823 Kress-Rogers, E., D'Costa, E. J., Sollars, J. E., Gibbs, P. A., Turner, A. P. F. (1993)  
1824 Measurement of meat freshness *in situ* with a biosensor array. *Food Control* 4, 149 –  
1825 154.
- 1826 Kumar, H., Neelam, R. (2016) Enzyme-based electrochemical biosensors for food  
1827 safety: a review. *Nanobiosensors in Disease Diagnosis* 5, 29 – 39.
- 1828 Kumar, S., Ghosh, A., Tudu, B., Bandyopadhyay, R. (2017) An equivalent electrical  
1829 network of an electronic tongue: A case study with tea samples, 2017 *ISOCS/IEEE*  
1830 *International Symposium on Olfaction and Electronic Nose (ISOEN)*, Montreal, QC,  
1831 Canada, 1 - 3.
- 1832 Kvasničk, F., Voldřich, M. (2000) Determination of fumaric acid in apple juice by on-  
1833 line coupled capillary isotachopheresis–capillary zone electrophoresis with UV  
1834 detection. *Journal of Chromatography A* 891, 175 – 181.
- 1835 Latorre-Moratalla, M.L., Bover-Cidl, S., Bosch-Fuste, J., Vidal-Carou, M.C. (2012)  
1836 Influence of technological conditions of sausage fermentation on the aminogenic  
1837 activity of *L. curvatus* CTC273. *Food Microbiology* 29, 43 -48.
- 1838 Laursen, B.G., Bay, L., Cleenwerck, I., Vancanneyt, M., Swings, J., Dalgaard, P.,  
1839 Leisner, J.J. (2005) *Carnobacterium divergens* and *Carnobacterium maltaromaticum*

- 1840 as spoilers or protective cultures in meat and seafood: phenotypic and genotypic  
1841 characterization. *Systematic and Applied Microbiology* **28**, 151 – 164.
- 1842 Leca-Bouvier, B., Blum, L. J. (2005) Biosensors for Protein Detection: A Review.  
1843 *Analytical Letters* **38**, 1491 – 1517.
- 1844 Ledenbach, L.H., and Marshall, R.T. (2009) Microbiological Spoilage of Dairy  
1845 Products. In W.H. Sperber, M.P. Doyle (eds.), *Compendium of the Microbiological*  
1846 *Spoilage of Foods and Beverages, Food Microbiology and Food Safety*, Springer  
1847 Verlag New York.
- 1848 Leduc, F., Tournayre, P., Kondjoyan, N., Mercier, F., Malle, P., Kol, O., Berdagué,  
1849 J.L., Duflos, G. (2012) Evolution of volatile odorous compounds during the storage of  
1850 European seabass (*Dicentrarchus labrax*). *Food Chemistry*, 131, 1304-1311.
- 1851 Legin, A., Rudnitskaya, A., Clapham, D., Seleznev, B., Lord, K. & Vlasov, Y. (2004)  
1852 Electronic tongue for pharmaceutical analytics: quantification of tastes and masking  
1853 effects. *Analytical and Bioanalytical Chemistry* **380**, 36 – 45.
- 1854
- 1855 Legin, A., Rudnitskaya, A., Di Natale, C., Mazzone, E., D'Amico, A. (2000)  
1856 Application of electronic tongue for qualitative and quantitative analysis of complex  
1857 liquid media. *Sensors and Actuators B: Chemical* **65**, 232 – 234.
- 1858 Lerma-García, M. J., Cerretani, L., Cevoli, C., Simó-Alfonso, E. F., Bendini, A., &  
1859 Toschi, T. G. (2010). Use of electronic nose to determine defect percentage in oils.  
1860 Comparison with sensory panel results. *Sensors and Actuators, B: Chemical*, 147(1),  
1861 283–289. <http://doi.org/10.1016/j.snb.2010.03.058>
- 1862 Li, C., Heinemann, P., & Sherry, R. (2007). Neural array and Bayesian array fusion  
1863 models to fuse electronic nose and surface acoustic wave sensor data for apple defect

- 1864 detection. *Sensors and Actuators, B: Chemical*, 125(1), 301–310.  
1865 <http://doi.org/10.1016/j.snb.2007.02.027>.
- 1866 Lim, J. W., Ha, D., Lee, J., Lee, S. K., Kim T. (2015) Review of  
1867 Micro/Nanotechnologies for Microbial Biosensors. *Frontiers in Bioengineering and*  
1868 *Biotechnology* **3**, 61. doi: 10.3389/fbioe.2015.00061
- 1869 Liu, Q., Wu, C., Cai, H., Hu, N., Zhou, J., Wang, P. (2014) Cell-Based Biosensors  
1870 and Their Application in Biomedicine. *Chemical Reviews* **114**, 6423 – 6461.
- 1871 Lopez-Caballero, M.E., Sanchez-Fernandez, J.A., Moral, A., (2001) Growth and  
1872 metabolic activity of *Shewanella putrefaciens* maintained under different CO<sub>2</sub> and O<sub>2</sub>  
1873 concentrations. *International Journal of Food Microbiology* **64**, 277-287.
- 1874 Lovdal, T. (2015) The microbiology of cold salmon. *Food Control* **54**, 360 – 373.
- 1875 Lozano, J. (2006) New technology in sensing odours: from human to artificial noses.  
1876 In: Teixeira da Silva JA (Ed) (2006) Floriculture, Ornamental and Plant  
1877 Biotechnology: Advances and Topical Issues (1st Edn, Vol IV), Global Science  
1878 Books, London, pp 152-161
- 1879 Lozano, J., Álvarez, F., Santos, J.P., Horrillo, C., (2011). Detection of acetic acid in  
1880 wine by means of an electronic nose. In: AIP Conference Proceedings - ISOEN11, pp.  
1881 176–177.
- 1882 Lozano, J., Santos, J. P., Gutiérrez, J., & Horrillo, M. C. (2007). Comparative study of  
1883 sampling systems combined with gas sensors for wine discrimination. *Sensors and*  
1884 *Actuators B: Chemical*, 126(2), 616–623. <http://doi.org/10.1016/j.snb.2007.04.018>
- 1885 Lozano, J., Santos, J. P., Suarez, J. I., Cabellos, M., Arroyo, T., & Horrillo, C. (2015).  
1886 Automatic Sensor System for the Continuous Analysis of the Evolution of Wine.

- 1887 American Journal of Enology and Viticulture, 66(2), 148–155.  
1888 <http://doi.org/10.5344/ajev.2014.14103>
- 1889 Lutén J.B. (2006) *Seafood Research from Fish to Dish: Quality, Safety and*  
1890 *Processing of Wild and Farmed Fish*. Wageningen Academic Pub.
- 1891 Lvova, L., Di Natale, C., Paolesse, R. (2017) Electronic tongue based on porphyrins  
1892 for Apulian red wines defects detection, *2017 ISOCS/IEEE International Symposium*  
1893 *on Olfaction and Electronic Nose (ISOEN)*, Montreal, QC, Canada, 1 - 2.
- 1894 Lytougou, A.E., Panagou, Z.E., Nychas, G.J.E (2017) Effect of different marinating  
1895 conditions on the evolution of spoilage microbiota and metabolomic profile of  
1896 chicken breast fillets. *Food Microbiol.* 66, 141 -179.
- 1897 Mace, S., Cornet, J., Chevalier, F., Cardinal, M., Pilet, M-F., Dousset, X., Joffrau, J-J.  
1898 (2012) Characterisation of the spoilage microbiota in raw salmon (*Salmo salar*) steaks  
1899 stored under vacuum or modified atmosphere packaging combining conventional  
1900 methods and PCR-TTGE. *Food Microbiology* 30, 164-172.
- 1901 Macías, M., Manso, A., Orellana, C., Velasco, H., Caballero, R., & Chamizo, J.  
1902 (2012). Acetic Acid Detection Threshold in Synthetic Wine Samples of a Portable  
1903 Electronic Nose. *Sensors*, 13(1), 208–220. <http://doi.org/10.3390/s130100208>
- 1904 Maffei M.E. (2010) Sites of synthesis, biochemistry and functional role of plant  
1905 volatiles. *South African Journal of Botany* 76, 612 – 631.
- 1906 Magan, N., & Evans, P. (2000). Volatiles as an indicator of fungal activity and  
1907 differentiation between species, and the potential use of electronic nose technology  
1908 for early detection of grain spoilage. *Journal of Stored Products Research*.  
1909 [http://doi.org/10.1016/S0022-474X\(99\)00057-0](http://doi.org/10.1016/S0022-474X(99)00057-0)

- 1910 Magan, N., Pavlou, A., & Chrysanthakis, I. (2001). Milk-sense: A volatile sensing  
1911 system recognizes spoilage bacteria and yeasts in milk. *Sensors and Actuators, B:*  
1912 *Chemical*, 72(1), 28–34. [http://doi.org/10.1016/S0925-4005\(00\)00621-3](http://doi.org/10.1016/S0925-4005(00)00621-3)
- 1913 Mancini, R. A., & Hunt, M. C. (2005). Current research in meat colour. *Meat Science*,  
1914 71, 100-121.
- 1915 Marín, S., Vinaixa, M., Brezmes, J., Llobet, E., Vilanova, X., Correig, X., ... Sanchis,  
1916 V. (2007). Use of a MS-electronic nose for prediction of early fungal spoilage of  
1917 bakery products. *International Journal of Food Microbiology*, 114(1), 10–16.  
1918 <http://doi.org/10.1016/j.ijfoodmicro.2006.11.003>
- 1919 Martin, A., Benito, M.J., Aranda, E., Ruiz-Moyano, S., Cordoba, J.J., Cordoba, M.J.  
1920 (2010) Characterization by volatile compounds of microbial deep spoilage in Iberian
- 1921 Martínez-Bisbal, M. C., Loeff, E., Olivas, E., Carbó, N., García-Castillo, F. J., López-  
1922 Carrero, J., Tormos, I., Tejadillos, F. J., Berlanga, J. G., Martínez-Mañez, R., Alcañiz,  
1923 M., Soto, J. (2017) A Voltammetric Electronic Tongue for the Quantitative Analysis  
1924 of Quality Parameters in Wastewater. *Electroanalysis* **29**, 1147 – 1153.
- 1925 Matysik F. M. (Ed.) (2017) *Trends in Bioelectroanalysis*. Springer International  
1926 Publishing AG.
- 1927 McGrath, T.F., Elliott, C.T., Fodey, T.L. (2012) Biosensors for the analysis of  
1928 microbiological and chemical contaminants in food. *Analytical and Bioanalytical*  
1929 *Chemistry* **403**, 75 - 92.
- 1930 Medina-Plaza, C., García-Hernandez, C., de Saja, J.A., Fernandez-Escudero, J.A.,  
1931 Barajas, E., Medrano, G., García-Cabezón, C., Martín-Pedrosa, F., Rodríguez-  
1932 Mendez, M.L. (2015) The advantages of disposable screen-printed biosensors in a

- 1933 bioelectronic tongue for the analysis of grapes. *LWT - Food Science and Technology*  
1934 **62**, 940 – 947.
- 1935 Mednova, O., Kirsanov, D., Rudnitskaya, A., Kilmartin, P., Legin, A. (2009)  
1936 Application Of A Potentiometric Electronic Tongue For The Determination Of Free  
1937 SO<sub>2</sub> And Other Analytical Parameters In White Wines From New Zealand. *AIP*  
1938 *Conference Proceedings* **1137**, 263. doi: <http://dx.doi.org/10.1063/1.3156521>
- 1939 Mehrotra, P. (2016) Biosensors and their applications – A review. *Journal of Oral*  
1940 *Biology and Craniofacial Research* **6**, 153 – 159.
- 1941 Mello, L. D., Kubota, L. T. (2002) Review of the use of biosensors as analytical tools  
1942 in the food and drink industries. *Food Chemistry* **77**, 237 – 256.
- 1943 Membre, J-M., Dagnas, S. (2016) Modelling microbial responses: application to food  
1944 spoilage. *Modelling in Food Microbiology* **2016**, 33 – 60.
- 1945 Mikš-Krajnik, M., Yoon, Y.J., Ukuku, D.O., Yuk, H.G. (2016) Volatile chemical  
1946 spoilage indexes of raw Atlantic salmon (*Salmo salar*) stored under aerobic condition  
1947 in relation to microbiological and sensory shelf lives. *Food Microbiol.* 53(Pt B), 182-  
1948 91.
- 1949 Mimendia, A., Gutiérrez, J. M., Leija, L., Hernández, P. R., Favari, L., Munoz, R., del  
1950 Valle, M. (2010) A review of the use of the potentiometric electronic tongue in the  
1951 monitoring of environmental systems. *Environmental Modelling & Software* **25**, 1023  
1952 – 1030.
- 1953 Monošík, R., Stred'anský, M., Šturdík, E. (2012) Biosensors - classification,  
1954 characterization and new trends. *Acta Chimica Slovaca* **5**, 109 - 120.

- 1955 Mossel, D.A.A., Corry, J.E.L., Struijk, C.B. and Baird, R.M. (1995) Essentials of the  
1956 Microbiology of Foods: a textbook for advanced studies. Wiley, England, pp. 175 –  
1957 214.
- 1958 Murugaboopathi, G., Parthasarathy, V., Chellaram, C., Prem Anand, T.,  
1959 Vinurajkumar, S. (2013) Applications of Biosensors in Food Industry. *Biosciences*  
1960 *Biotechnology Research Asia* **10**, 711 - 714.
- 1961 Mutlu, M. (2016) *Biosensors in Food Processing, Safety, and Quality Control*. CRC  
1962 Press.
- 1963 Muzaddadi, A.U., Devatkal, S., Oberoi, H.S. (2016) *Seafood Enzymes and Their*  
1964 *Application in Food Processing* – Chapter 9. Agro-Industrial Wastes as Feedstock for  
1965 Enzyme Production. Apply and Exploit the Emerging and Valuable Use Options of  
1966 Waste Biomass, 201 – 232.
- 1967 Nagle, H. T. J. W. G. T. C. P. S. S. S. (2006). Handbook of machine olfaction:  
1968 electronic nose technology. *Aging* (Vol. 7). <http://doi.org/10.1002/3527601597>.
- 1969 Naila, A., Flint, S., Fletcher, G., Bremer, P., Meerdink, G. (2010) Control of Biogenic  
1970 Amines in Food—Existing and Emerging Approaches. *Journal of Food Science* **75**,  
1971 R139 – R150.
- 1972 Narsaiah, K., Jha, S. N., Bhardwaj, R., Sharma, R., Kumar, R. (2012) Optical  
1973 biosensors for food quality and safety assurance—a review. *Journal of Food Science*  
1974 *and Technology* **49**, 383 – 406.
- 1975 Ndagijimana, M., Chaves-Lopez, C., Corsetti, A., Tofalo, R., Sergi, M., Paparella, A.  
1976 (2008) Growth and metabolites production by *Penicillium brevicompactum* in  
1977 yoghurt, *Int. J. Food Microbiol.* **127** 276–283.

- 1978 Nelson, D. L., Cox, M. M. (2017) *Lehninger Principles of Biochemistry*, 7<sup>th</sup> Edition,  
1979 Macmillan Higher Education, New York.
- 1980 Nychas, G.J.E., Skandamis, P.N., Tassou, C.C., Koutsoumanis, K.P., (2008) Meat  
1981 spoilage during distribution. *Meat Science* **78**, 77 -89.
- 1982 Oehlenschlager, J. (2014) Seafood quality assessment. In Boziaris, I.S. (Ed.) *Seafood*  
1983 *Processing. Technology, Quality and Safety. IFST Advances in Food Science Series*,  
1984 Wiley Blackwell, 361 – 386.
- 1985 Otto, M., Thomas, J. D. R. (1985) Model studies on multiple channel analysis of free  
1986 magnesium, calcium, sodium, and potassium at physiological concentration levels  
1987 with ion-selective electrodes. *Analytical Chemistry* **57**, 2647 – 2651.
- 1988 Paarup, T., Nieto, J. C., Peláez, C., Reguera, J. I. (1999) Microbiological and physico-  
1989 chemical characterisation of deep spoilage in Spanish dry-cured hams and  
1990 characterisation of isolated Enterobacteriaceae with regard to salt and temperature  
1991 tolerance. *European Food Research and Technology* **209**, 366 – 371.
- 1992 Paludan-Muller, C., Dalgaard, P., Huss, H.H., Gram, L. (1998) Evaluation of the role  
1993 of *Carnobacterium piscicola* in spoilage of vacuum- and modified-atmosphere-  
1994 packed cold-smoked salmon stored at 5°C. *International Journal of Food*  
1995 *Microbiology*, **39**, 155 – 168.
- 1996 Panigrahi, S., Balasubramanian, S., Gu, H., Logue, C., Marchello, M., 2006a. Neural-  
1997 network-integrated electronic nose system for identification of spoiled beef. *LWT* **39**,  
1998 135–145.
- 1999 Panigrahi, S., Balasubramanian, S., Gu, H., Logue, C., Marchello, M., 2006b. Design  
2000 and development of a metal oxide based electronic nose for spoilage classification of  
2001 beef. *Sensors and Actuators B* **119**, 2–14

- 2002 Papadopoulou, O.S., Panagou, E.Z., Mohareb, F.R., Nychas, G.E., 2013. Sensory and  
2003 microbiological quality assessment of beef fillets using a portable electronic nose in  
2004 tandem with support vector machine analysis. *Food Research International* 50, 241–  
2005 249.
- 2006 Park, Y. W., Kim, S. M., Lee, J. Y., Jang, W. (2015) Application of biosensors in  
2007 smart packaging. *Molecular & Cellular Toxicology* **11**, 277 – 285.
- 2008 Parlapani, F.F., Mallouchos, A., Haroutounian, S.A., Boziaris, I.S. (2017)  
2009 Microbiological spoilage and investigation of volatile profile during storage of sea  
2010 bream fillets under various conditions. *International Journal of Food Microbiology*  
2011 **189**, 153 – 163.
- 2012 Parpalani, F.F., Boziaris, I.S. (2016) Monitoring of spoilage and determination of  
2013 microbial communities based on 16S rRNA gene sequence analysis of whole sea  
2014 bream stored at various temperatures. *LWT Food Science and Technology*, **66**, 553 –  
2015 559.
- 2016 Parpalani, F.F., Haroutounian, S.A., Nychas, G.J., Boziaris, I.S. (2015)  
2017 Microbiological spoilage and volatiles production of gutted European sea bass stored  
2018 under air and commercial modified atmosphere package at 2°C. *Food Microbiology*  
2019 **50**, 44 – 53.
- 2020 Parpalani, F.F., Meziti, A., Kormas, K.Ar., Boziaris., I.S. (2013) Indigenous and  
2021 spoilage microbiota of farmed sea bream stored on ice identified by phenotypic and  
2022 16S rRNA gene analysis. *Food Microbiology* **33**, 85 – 89.
- 2023 Patel, P. D. (2006) Overview of affinity biosensors in food analysis. *Journal of AOAC*  
2024 *International* **89**, 805-818.

- 2025 Pathange, L. P., Mallikarjunan, P., Marini, R. P., O'Keefe, S., & Vaughan, D. (2006).  
2026 Non-destructive evaluation of apple maturity using an electronic nose system. *Journal*  
2027 *of Food Engineering*, 77(4), 1018–1023.  
2028 <http://doi.org/10.1016/j.jfoodeng.2005.08.034>
- 2029 Pearce, T. C., Schiffman, S. S., Nagle, H. T., Gardner, J. W. (2006) *Handbook of*  
2030 *Machine Olfaction: Electronic Nose Technology*. John Wiley & Sons.
- 2031 Pein, M., Kirsanov, D., Ciosek, P., del Valle, M., Yaroshenko, I., Wesoly, M.,  
2032 Zabadaj, M., Gonzalez-Calabuig, A., Wróblewski, W., Legin, A. (2015) Independent  
2033 comparison study of six different electronic tongues applied for pharmaceutical  
2034 analysis. *Journal of Pharmaceutical and Biomedical Analysis* **114**, 321-329.
- 2035 Pennacchia, C., Ercolini, D., Villiani, F. (2011) Spoilage-related microbiota  
2036 associated with chilled beef stored in air or vacuum pack. *Food Microbiology* **28**, 84 –  
2037 93.
- 2038 Pérez-López, B., Merkoçi, A. (2011) Nanomaterials based biosensors for food  
2039 analysis applications. *Trends in Food Science & Technology* **22**, 625-639.
- 2040 Peris, M., Escuder-Gilabert, L. (2013) On-line monitoring of food fermentation  
2041 processes using electronic noses and electronic tongues: A review. *Analytica Chimica*  
2042 *Acta* **804**, 29 – 36.
- 2043 Peris, M., Escuder-Gilabert, L. (2013). On-line monitoring of food fermentation  
2044 processes using electronic noses and electronic tongues: a review. *Analytica Chimica*  
2045 *Acta*, 804(4), 29-36.
- 2046 Perry, N. (2012) Dry aging beef. *International Journal of Gastronomy and Food*  
2047 *Science* **1**, 78 – 80.

- 2048 Pinu, F., 2016. Early detection of food pathogens and food spoilage microorganisms:  
2049 Application of metabolomics. *Trends in Food Science & Technology* 54, 213-215.
- 2050 Pioggia, G., Di Francesco, F., Marchetti, A., Ferro, M., Leardi, R., Ahluwalia, A.  
2051 (2007) A composite sensor array impedentiometric electronic tongue: Part II.  
2052 Discrimination of basic tastes. *Biosensors and Bioelectronics* 22, 2624 – 2628.
- 2053 Piriya, V. S. A., Joseph, P., Daniel, S. C. G. K., Lakshmanan, S., Kinoshita, T.,  
2054 Muthusamy, S. (2017) Colorimetric sensors for rapid detection of various analytes.  
2055 *Materials Science & Engineering. C, Materials for Biological Applications* 78, 1231 -  
2056 1245.
- 2057 Pividori, M. I., Alegret S. (2010) Electrochemical biosensors for food safety.  
2058 *Contributions to Science* 6, 173 – 191.
- 2059 Poltronieri, P., Mezzolla, V., Primiceri, E., Maruccio G. (2014) Biosensors for the  
2060 Detection of Food Pathogens. *Foods* 3, 511 – 526.
- 2061 Prodromidis, M. I., Karayannis, M. I. (2002) Enzyme Based Amperometric  
2062 Biosensors for Food Analysis. *Electroanalysis* 14, 241 – 261.
- 2063 Quigely, L., O’Sullivan, O., Stanton, C., Beresford, T.P., Ross, R.P., Fitzgeralds, G.F.,  
2064 Cotter, P.D. (2013) The complex microbiota of raw milk. *FEMS Microbiol Rev.* 37,  
2065 664 -698.
- 2066 Ragazzo-Sanchez, J. A., Chalier, P., Chevalier-Lucia, D., Calderon-Santoyo, M., &  
2067 Ghommidh, C. (2009). Off-flavours detection in alcoholic beverages by electronic  
2068 nose coupled to GC. *Sensors and Actuators, B: Chemical*, 140(1), 29–34.  
2069 <http://doi.org/10.1016/j.snb.2009.02.061>

- 2070 Rajamaki, T., Alakomi, H., Ritvanen, T., Skytta, E., Smolander, M., Ahvenainen, R.,  
2071 2006. Application of an electronic nose for quality assessment of modified  
2072 atmosphere packaged poultry meat. *Food Control* 17, 5–13.
- 2073 Ramirez-Guizar, S., Sykes, H., Perry, J.D., Schwalbe, E.C., Stanforth, S.P., Perez-  
2074 Perez, C.I.M., Dean, J.R. (2017) A chromatographic approach to distinguish Gram-  
2075 positive from Gram-negative bacteria using exogenous volatile organic compound  
2076 metabolites. *Journal of Chromatography A* **1501**, 79-88.
- 2077 Remenant, B., Jaffres, E., Dousset, X., Pilet, M-F., Zagorec, M. (2015) Bacterial  
2078 spoilers of food: Behaviour, fitness and functional properties. *Food Microbiology* **45**,  
2079 45 -53.
- 2080 Riul Jr., A., dos Santos Jr., D. S., Wohnrath, K., Di Tommazo, R., Carvalho, A. C. P.  
2081 L. F., Fonseca, F. J., Oliveira Jr., O. N., Taylor, D. M., Mattoso, L. H. C. (2002)  
2082 Artificial Taste Sensor: Efficient Combination of Sensors Made from Langmuir-  
2083 Blodgett Films of Conducting Polymers and a Ruthenium Complex and Self-  
2084 Assembled Films of an Azobenzene-Containing Polymer. *Langmuir* **18**, 239 – 245.
- 2085 Rodríguez-Delgado, M. M., Alemán-Nava, G. S., Rodríguez-Delgado, J. M., Dieck-  
2086 Assad, G., Martínez-Chapa, S. O., Barceló, D., Parra, R. (2015) Laccase-based  
2087 biosensors for detection of phenolic compounds. *Trends in Analytical Chemistry* **74**,  
2088 21 – 45.
- 2089 Rodríguez-Méndez, M. L., Medina-Plaza, C., García-Hernández, C., de Saja, J. A.,  
2090 Fernández-Escudero J. A., Barajas-Tola, E., Medrano G. (2014) Analysis of grapes  
2091 and wines using a voltammetric bioelectronic tongue. Correlation with the phenolic  
2092 and sugar content. *IEEE Sensors 2014 Proceedings*, Valencia, 2139 - 2142.

- 2093 Rodríguez-Méndez, M.L., Apetrei, C., de Saja, J.A. (2008) Evaluation of the  
2094 polyphenolic content of extra virgin olive oils using an array of voltammetric sensors.  
2095 *Electrochimica Acta* **53**, 5867 – 5872.
- 2096 Rodríguez-Méndez, M.L., Gay, M., Apetrei, C., de Saja, J.A. (2009) Biogenic amines  
2097 and fish freshness assessment using a multisensor system based on voltammetric  
2098 electrodes. Comparison between CPE and screen-printed electrodes. *Electrochimica*  
2099 *Acta* **54**, 7033 – 7041.
- 2100 Rogers, K. R. (2000) Principles of affinity-based biosensors. *Molecular*  
2101 *Biotechnology* **14**, 109 – 129.
- 2102 Rotariu, L., Lagarde, F., Jaffrezic-Renault, N., Bala, C. (2016) Electrochemical  
2103 biosensors for fast detection of food contaminants –trends and perspective. *TrAC*  
2104 *Trends in Analytical Chemistry* **79**, 80 – 87.
- 2105 Rudnitskaya, A., Polshin, E., Kirsanov, D., Lammertyn, J., Nicolai, B., Saison, D.,  
2106 Delvaux, F. R., Delvaux, F., Legin, A. (2009) Instrumental measurement of beer taste  
2107 attributes using an electronic tongue. *Analytica Chimica Acta* **646**, 111 – 118.
- 2108 Rudnitskaya, A., Schmidtke, L. M., Reis, A., Domingues, M. R., Delgadillo, I.,  
2109 Debus, B., Kirsanov, D., Legin, A. (2017) Measurements of the effects of wine  
2110 maceration with oak chips using an electronic tongue. *Food Chemistry* **229**, 20 - 27.
- 2111 Ruiz-Rico, M., Fuentes, A., Masot, R., Alcañiz, M., Fernández-Segovia, I., Barat, J.  
2112 M. (2013) Use of the voltammetric tongue in fresh cod (*Gadus morhua*) quality  
2113 assessment. *Innovative Food Science and Emerging Technologies* **18**, 256 – 263.
- 2114 Sade, E., Penttinen, K., Bjorkroth, J., Hultman, J. (2017) Exploring lot-to-lot variation  
2115 in spoilage bacterial communities on commercial modified atmosphere packaged  
2116 beef. *Food Microbiology* **62**, 147 – 152.

- 2117 Sahu, M., Bala, S. (2017) Food Processing, Food Spoilage and their Prevention: An  
2118 Overview. *International Journal of Life-Sciences Scientific Research* **3**, 753 - 759.
- 2119 Santos, J. P., & Lozano, J. (2015). Real time detection of beer defects with a hand  
2120 held electronic nose. In Proceedings of the 2015 10th Spanish Conference on Electron  
2121 Devices, CDE 2015. Institute of Electrical and Electronics Engineers Inc.
- 2122 Santos, J. P., Fernández, M. J., Fontecha, J. L., Lozano, J., Aleixandre, M., García,  
2123 M., et al., Horrillo, M. C. (2005). SAW sensor array for wine discrimination. *Sensors*  
2124 and *Actuators* B: Chemical, 107(1), 291–295.  
2125 <http://doi.org/10.1016/j.snb.2004.10.013>.
- 2126 Santos, J. P., Lozano, J., Aleixandre, M., Arroyo, T., Cabellos, J. M., Gil, M., &  
2127 Horrillo, M. del C. (2010). Threshold detection of aromatic compounds in wine with  
2128 an electronic nose and a human sensory panel. *Talanta*, 80(5), 1899–906.  
2129 <http://doi.org/10.1016/j.talanta.2009.10.041>
- 2130 Sarnoski, P.J., Jahncke, M.L., O’Keefe, S.F., Mallikarjunan, P., & Flick, G.J.  
2131 2008. *Journal of Aquatic Food Product Technology*, 17(3), 234-252.
- 2132 Schmidt, V.S.J., Kaufmann, V., Kulozik, U., Scherer, S., Wenning, M. (2012)  
2133 Microbial biodiversity, quality and shelf life of microfiltered and pasteurized  
2134 extended shelf life (ESL) milk from Germany, Austria and Switzerland. *International*  
2135 *Journal of Food Microbiology* **154**, 1-9.
- 2136 Schmidtke, L. M., Rudnitskaya, A., Saliba, A. J., Blackman, J. W., Scollary, G. R.,  
2137 Clark, A. C., Rutledge, D. N., Delgado, I., Legin A. (2010) Sensory, Chemical, and  
2138 Electronic Tongue Assessment of Micro-oxygenated Wines and Oak Chip  
2139 Maceration: Assessing the Commonality of Analytical Techniques. *Journal of*  
2140 *Agricultural and Food Chemistry* **58**, 5026 – 5033.

- 2141 Scognamiglio, V., Arduini, F., Palleschi, G., Rea, G. (2014) Biosensing technology  
2142 for sustainable food safety. *Trends in Analytical Chemistry* **62**, 1 – 10.
- 2143 Scognamiglio, V., Rea, G., Arduini, F., Palleschi, G. (Eds.) (2016) *Biosensors for*  
2144 *Sustainable Food - New Opportunities and Technical Challenges*. Comprehensive  
2145 Analytical Chemistry, Volume 74, 3-432, Elsevier BV.
- 2146 Sehra, G., Cole, M., Gardner, J. W. (2004) Miniature taste sensing system based on  
2147 dual SH-SAW sensor device: an electronic tongue. *Sensors and Actuators B:*  
2148 *Chemical* **103**, 233 – 239.
- 2149 Shah, J. S. (2013) An Electronic tongue for core taste identification based on  
2150 conductometry. *International Journal of Engineering Research and Applications* **3**,  
2151 961 - 963
- 2152 Shao, Y., Wang, J., Wu, H., Liu, J., Aksay. I. A., Lina, Y. (2010) Graphene Based  
2153 Electrochemical Sensors and Biosensors: A Review. *Electroanalysis* **22**, 1027 – 1036.
- 2154 Sharma, P., Ghosh, A., Tudu, B., Sabhapondit, S., Baruah, B. D., Tamuly, P., ...  
2155 Bandyopadhyay, R. (2015). Monitoring the fermentation process of black tea using  
2156 QCM sensor based electronic nose. *Sensors and Actuators, B: Chemical*, 219, 146–  
2157 157. <http://doi.org/10.1016/j.snb.2015.05.013>
- 2158 Si, R. W., Zhai, D. D., Liao, Z. H., Gao, L., Yong, Y. C. (2015) A whole-cell  
2159 electrochemical biosensing system based on bacterial inward electron flow for  
2160 fumarate quantification. *Biosensors and Bioelectronics* **68**, 34 – 40.
- 2161 Śliwiska, M., Wisniewska, P., Dymerski, T., Namiesnik, J., Wardencki, W. (2014)  
2162 Food Analysis Using Artificial Senses. *Journal of Agricultural and Food Chemistry*  
2163 **62**, 1423 – 1448.

- 2164 Smyth, H., Cozzolino, D. (2013) Instrumental methods (spectroscopy, electronic nose,  
2165 and tongue) as tools to predict taste and aroma in beverages: advantages and  
2166 limitations. *Chemical Reviews* **113**, 1429 – 1440.
- 2167 Song, H. S., Jin, H. J., Ahn, S. R., Kim, D., Lee, S. H., Kim, U. K., Simons, C. T.,  
2168 Hong, S., Park, T. H. (2014) Bioelectronic tongue using heterodimeric human taste  
2169 receptor for the discrimination of sweeteners with human-like performance. *ACS*  
2170 *Nano* **8**, 9781 - 9789.
- 2171 Song, H. S., Jin, H. J., Ahn, S. R., Kim, D., Lee, S. H., Kim, U. K., Simons, C. T.,  
2172 Hong, S., Park, T. H. (2014) Bioelectronic Tongue Using Heterodimeric Human Taste  
2173 Receptor for the Discrimination of Sweeteners with Human-like Performance. *ACS*  
2174 *Nano* **8**, 9781 – 9789.
- 2175 Song, X., Xu, S., Chen, L., Wei, Y., Xiong, H. (2014) Recent advances in molecularly  
2176 imprinted polymers in food analysis. *Journal of Applied Polymer Science* **131**, 40766.  
2177 doi: 10.1002/app.40766
- 2178 Spadafora, N.D., Paramithiotis, S., Drosinos, E., Cammarisano, L., Rogers, H.J.,  
2179 Muller, C.T. (2016) Detection of *Listeria monocytogenes* in cut melon fruit using  
2180 analysis of volatile organic compounds. *Food Microbiology* **54**, 52 – 59.
- 2181 Stadler, R. H., Lineback, D. R. (2008) *Process-Induced Food Toxicants: Occurrence,*  
2182 *Formation, Mitigation, and Health Risks.* John Wiley & Sons.
- 2183 Susiluoto, T., Korkeala, H., Bjorkroth, K.J. (2003) *Leuconostoc gasicomitatum* is the  
2184 dominating lactic acid bacterium in retail modified-atmosphere-packaged marinated  
2185 broiler meat strips on sell-by-day. *International Journal of Food Microbiology* **80**, 89  
2186 – 97.

- 2187 Suzzia, G., Gardini, F. (2002) Biogenic amines in dry fermented sausages: a review.  
2188 *International Journal of Food Microbiology* 88, 41 -54.
- 2189 Tahara, Y., Toko, K. (2013) Electronic Tongues–A Review. *IEEE Sensors Journal*  
2190 **13**, 3001 - 3011.
- 2191 Tait, E., Stanforth, S.P., Reed, S., Perry, J.D., Dean, J.R. (2014) Use of volatile  
2192 compounds as a diagnostic tool for the detection of pathogenic bacteria. *Trends Anal.*  
2193 *Chem.* 53, 117 -125.
- 2194 *Technologies, Sensors (Basel)* 9, 5099–5148.
- 2195 Tembe, S., D’Souza, S. F. (2015) Immobilisation strategies for construction of  
2196 tyrosinase-based biosensors. *Materials Technology* **30**, B190 - B195.
- 2197 The, K.H., Flint, S., Palmer, J., Andrewes, P., Bremer, P., Lindsay, D. (2014) Biofilm  
2198 – An unrecognised source of spoilage enzymes in dairy products? *International Dairy*  
2199 *Journal* 34, 32-40.
- 2200 Thévenot, D. R., Toth, K., Durst, R. A., Wilson, G. S. (2001) Electrochemical  
2201 biosensors: recommended definitions and classification. *Biosensors and*  
2202 *Bioelectronics* **16**, 121 – 131.
- 2203 Tian, H., Feng, T., Xiao, Z., Song, S., Li, Z., Liu, Q., Mao, D., Li, F. (2015)  
2204 Comparison of intensities and binary interactions of four basic tastes between an  
2205 electronic tongue and a human tongue. *Food Science and Biotechnology* **24**, 1711 –  
2206 1715.
- 2207 Tian, X., Cai, Q., & Zhang, Y., 2012. Rapid Classification of Hairtail Fish and Pork  
2208 Freshness Using an Electronic Nose Based on the PCA Method. *Sensors*, 12, 260-277.

- 2209 Timsorn, K., Thoopboochagorn, T., Lertwattanasakul, N., & Wongchoosuk, C.,  
2210 (2016). Evaluation of bacterial population on chicken meats using a briefcase  
2211 electronic nose. *Biosystems Engineering*, 151, 116 -125.
- 2212 Toko K. (2000) *Biomimetic Sensor Technology*, Cambridge University Press.
- 2213 Toko, K. (1998) Electronic sensing of tastes. *Sensors Update* **3**, 131 – 160.
- 2214 Turner, A. P. F. (2013) Biosensors: sense and sensibility. *Chemical Society Reviews*  
2215 **42**, 3184 – 3196.
- 2216 ul Hasan, N., Ejaz, N., Ejaz, W., & Kim, H. S. (2012). Meat and fish freshness  
2217 inspection system based on odor sensing. *Sensors (Basel, Switzerland)*, 12(11),  
2218 15542–15557.
- 2219 Upadhyay, L. S. B., Verma, N. (2013) Enzyme Inhibition Based Biosensors: A  
2220 Review. *Analytical Letters* **46**, 225 – 241.
- 2221 Upadhyay, R., Sehwal, S., & Mishra, H. N. (2017). Electronic nose guided  
2222 determination of frying disposal time of sunflower oil using fuzzy logic analysis.  
2223 *Food Chemistry*, 221, 379–385. <http://doi.org/10.1016/j.foodchem.2016.10.089>
- 2224 Valerio, F., De Bellis, P., Di Biase, M., Lonigro, S.L., Giussani, B., Visconti, A.,  
2225 Lavermicocca, P., Sisto, A. (2012) Diversity of spore forming bacteria and  
2226 identification of *Bacillus amyloliquefaciences* as a species frequently associated with  
2227 the ropy spoilage of bread. *International Journal of Food Microbiology* **156** 278 –  
2228 285.
- 2229 Vasilescu, A., Nunes, G., Hayat, A., Latif, U., Marty, J.-L. (2016) Electrochemical  
2230 Affinity Biosensors Based on Disposable Screen-Printed Electrodes for Detection of  
2231 Food Allergens. *Sensors* **16**, 1863. doi:10.3390/s16111863
- 2232 Verma, P., & Yadava, R. D. S. (2015). Polymer selection for SAW sensor array based  
2233 electronic noses by fuzzy c-means clustering of partition coefficients: Model studies

- 2234 on detection of freshness and spoilage of milk and fish. *Sensors and Actuators, B:*  
2235 *Chemical*, 209, 751–769. <http://doi.org/10.1016/j.snb.2014.11.149>
- 2236 Vilas, C., Alonso, A.A., Herrera, J.R., García-Blanco, A., García, M.R. (2017) A  
2237 model for the biochemical degradation of inosine monophosphate in hake (*Merluccius*  
2238 *merluccius*). *Journal of Food Engineering* **200**, 95 – 101.
- 2239 Vlasov, Y., Legin, A., Rudnitskaya, A., Di Natale, C., D’Amico, A. (2005)  
2240 Nonspecific sensor arrays (“electronic tongue”) for chemical analysis of liquids. *Pure*  
2241 *and Applied Chemistry* **77**, 1965 – 1983.
- 2242 von Neubeck, M., Baur, C., Krewinkel, M., Stoeckel, M., Krantz, B., Stressler, T.,  
2243 Jorg, L.F. (2015) Biodiversity of refrigerated raw milk microbiota and their  
2244 enzymatic spoilage potential. *International Journal of Food Microbiology*, 211, 57 –  
2245 65.
- 2246 Vytrasova, J., Pribanova, P., Marvanova, L. (2002) Occurrence of xerophilic fungi in  
2247 bakery gingerbread production. *International Journal of Food Microbiology* **72**, 91-  
2248 96.
- 2249 Wackerlig, J., Schirhagl, R. (2016) Applications of Molecularly Imprinted Polymer  
2250 Nanoparticles and Their Advances toward Industrial Use: A Review. *Analytical*  
2251 *Chemistry* **88**, 250 – 261.
- 2252 Wang, C., Yang, J., Zhu, X., Lu, Y., Xue, Y., Lu, Z., 2017. Effects of Salmonella  
2253 bacteriophage, nisin and potassium sorbate and their combination on safety and shelf  
2254 life of fresh chilled pork. *Food Control*, 73, 869-877.
- 2255 Wang, G-y., Wang, H-h., Han, Y-w., Xing, T., Ye, K-p., Xu, X-l., Zhou, G-h. (2017)  
2256 Evaluation of the spoilage potential of bacteria isolated from chilled chicken in vitro  
2257 and in situ. *Food Microbiology* **63**, 139 – 146.

- 2258 Wang, H., Hu, Z., Long, F., Guo, C., Yuan, Y., Yue, T (2016) Early detection of  
2259 *Zygosaccharomyces rouxii*—spawned spoilage in apple juice by electronic nose  
2260 combined with chemometrics. *Int. J. Food Microbiol.* 217, 68 -78.
- 2261 Wang, J. C. (2000). *Analytical electrochemistry*, John Wiley & Sons, Chichester.
- 2262 Wang, Y., Li, Y., Yang, J., Ruan, J., & Sun, C., 2016. Microbial volatile organic  
2263 compounds and their application in microorganism identification in foodstuff. *Trends*  
2264 *in Analytical Chemistry*, 78, 1–16.
- 2265 Wang, Y., Li, Y., Yang, J., Ruan, J., Sun, C. (2016) Microbial volatile organic  
2266 compounds and their application in microorganism identification in foodstuff. *Trends*  
2267 *in Analytical Chemistry* **78**, 1-16.
- 2268 Weber, W., Luzi, S., Karlsson, M., Fussenegger, M. (2009) A novel hybrid dual-  
2269 channel catalytic-biological sensor system for assessment of fruit quality. *Journal of*  
2270 *Biotechnology* **139**, 314 – 317.
- 2271 Wilson, A.D., Bauietto, M. (2009) Applications and Advances in Electronic-Nose
- 2272 Winqvist F. (2008) Voltammetric electronic tongues – basic principles and  
2273 applications. *Microchimica Acta* **163**, 3 – 10.
- 2274 Winqvist, F., Holmin, S., Krantz-Rückler, C., Wide, P., Lündström, I. (2000) A  
2275 hybrid electronic tongue. *Analytica Chimica Acta* **406**, 147 – 157.
- 2276 Winqvist, F., Olsson, J., Eriksson, M. (2011) Multicomponent analysis of drinking  
2277 water by a voltammetric electronic tongue. *Analytica Chimica Acta* **683**, 192 – 197.
- 2278 Wojnowski, W., Majchrzak, T., Dymerski, T., Gębicki, J., Namieśnik, J., 2017.  
2279 Electronic noses: Powerful tools in meat quality assessment. *Meat Science* 131, 119–  
2280 131

- 2281 Yang, M., Soga, T., Pollard, P. J., Adam, J. (2012) The emerging role of fumarate as  
2282 an oncometabolite. *Frontiers in Oncology* **2**, 85. doi: 10.3389/fonc.2012.00085
- 2283 Yang, S.P., Xie, J., Qiang, Y-F (2017) Determination of Spoilage Microbiota of  
2284 Pacific White Shrimp During Ambient and Cold Storage Using Next-Generation  
2285 Sequencing and Culture-Dependent Method. *Journal of Food Science* **82**, 1178 –  
2286 1183.
- 2287 Yang, X., Badoni, M. (2013) Substrate utilization during incubation in meat juice  
2288 medium of psychrotolerant clostridia associated with blown pack spoilage. *Food*  
2289 *Microbiology* **34**, 400 -405.
- 2290 Yano, Y., Yokoyama, K., Tamiya, E., Karube, I. (1996) Direct evaluation of meat  
2291 spoilage and the progress of aging using biosensors. *Analytica Chimica Acta* **320**, 269  
2292 – 276.
- 2293 Yongwei, W., Wang, J., Zhou, B., & Lu, Q. (2009). Monitoring storage time and  
2294 quality attribute of egg based on electronic nose. *Analytica Chimica Acta*, 650(2),  
2295 183–188. <http://doi.org/10.1016/j.aca.2009.07.049>.
- 2296 Yu, K., Thomas, R., Hamilton-Kemp, T.R., Archbold, D.D., Collins, R.W., Newman,  
2297 M.C. (2000) Volatile compounds from *Escherichia coli* O157:H7 and their absorption  
2298 by strawberry fruit. *Journal of Agriculture and Food Chemistry*, **48**, 413 - 417.
- 2299 Zeravik, J., Hlavacek, A., Lacina, K., Skládal P. (2009) State of the Art in the Field of  
2300 Electronic and Bioelectronic Tongues – Towards the Analysis of Wines.  
2301 *Electroanalysis* **21**, 2509 – 2520.
- 2302 Zhang, F., Keasling, J. (2011) Biosensors and their applications in microbial  
2303 metabolic engineering. *Trends in Microbiology* **19**, 323 – 329.

2304 Zhou, G.H., Xu, X.L., Liu, Y. (2010) Preservative technologies for fresh meat – A  
2305 review. *Meat Science* **86**, 119 – 128.

2306 Zoski C. (ed) (2007). *Handbook of Electrochemistry*, 1<sup>st</sup> Edition, Elsevier Science,  
2307 New York.

2308

2309

2310

2311

2312

2313

2314

### 2315 **Captions of Figures**

2316 **Fig. 1.** Block diagram of an electronic nose system.

2317 **Fig. 2.** General scheme of an electronic tongue system

2318 **Fig. 3.** Biosensor detection scheme

2319 **Fig. 4.** Portable electronic nose system for the defect discrimination in beer

2320 **Fig. 5.** Decomposition of ATP in the muscles (Nelson & Cox, 2017)

2321 **Fig. 6.** The sensor array used for the potentiometric electronic tongue (Gil-Sánchez,

2322 Soto, Martínez-Mañez, Garcia-Breijo, Ibáñez, & Llobet, 2011).

**Table 3.** The main sensorial properties and their relative compounds.

<b>Taste</b>	<b>Compounds</b>
Sweetness	Glucose, Sucrose, Fructose, D-Amino acids, Sweeteners (natural or artificial)
Sourness	Acetic acid, Citric acid, Tartaric acid, Lactic acid, Phosphoric acid
Saltiness	NaCl, KCl
Bitterness	Quinine, Caffeine, MgCl <sub>2</sub> , Humulone, L-Amino acids
Umami	Monosodium glutamate, Glutamic acid, Disodium inosinate, Disodium guanylate
Astringency	Tannins
Pungency	Capsaicin, piperine

**Table 4.** A summarized overview on the application of electronic nose to food spoilage detection

Application	Sensor technology	Number of sensors	Additional Techniques	Data processing algorithm	References
Wine monitoring	MOX	16	GC-MS	PCA, PNN	(J. Lozano et al., 2015)
Acetic Acid in wine	MOX(PEN3)	10	-	PCA, MLP	(Macías et al., 2012)
	MOX	4	-	PCA, RBFNN	(Lozano et al., 2011)
Wine spoilage, off-flavors	Humid e-nose	5	E-tongue	PCA, K-means	(Gil-Sanchez et al., 2011)
	MOX(FOX 3000)	12	-	PCA, CLA	(Cabañes, Sahgal, Bragulat, & Magan, 2009)
	MOX (FOX4000)	18	-	PCA, DFA	(Ragazzo-Sanchez, Chalier, Chevalier-Lucia, Calderon-Santoyo, & Ghommidh, 2009)
	MOX (FOX 3000)	12	MS	PLS	(Berna, Trowell, Cynkar, & Cozzolino, 2008)
Red wine spoilage induced by <i>Brettanomyces</i> yeast	MS-enose	-	GC-MS	PCA, SLDA, PLS	(Cynkar, Cozzolino, Damberg, Janik, & Gishen, 2007)
Threshold detection wine compounds	MOX	16	Sensory panel	PCA, NN	(José Pedro Santos et al., 2010)
Beer defects	MOX	4	-	PCA, NN	(Jose Pedro Santos &

					Lozano, 2015)
Fried potato	MOX (Figaro)	8	Biochemical assays	Fuzzy logic, PCA, ANOVA	(Chatterjee, Bhattacharjee, & Bhattacharyya, 2014)
Microbial contamination in tomatoes	MOX (EOS835 – Sacmi)	6	DHS-GC-MS	PCA, Pearson correlation	(Concina et al., 2009)
Egg quality	MOX	8	-	PCA, LDA, BPNN, GANN, QPSR	(Yongwei, Wang, Zhou, & Lu, 2009)
Grain spoilage (review)	MOX	17	-	DFA, Neural Networks	(N. Magan & Evans, 2000)
Spoiled Rapeseed	MOX (Agrinose)	8	HPLC, Colony Forming Units, Fourier Transform Infrared (FT-IR) Spectra	PCA	(Gancarz et al., 2017)
Enterobacteriaceae in vegetable soups	MOX (EOS507C)	4	GC-MS	PCA,LDA, Pearson correlation	(Emanuela Gobbi et al., 2015)
Spoilage of bakery products	MS-enose	-	HPLC	PLS	(Marín et al., 2007)
Contamination of soft drinks	MOX (EOS835)	6	PCR, HPLC	PCA, LDA, kNN, SVM	(Concina et al., 2010)
Alicyclobacillus spp. spoilage of fruit juices	MOX (EOS835)	6	DHS-GC-MS	PCA, Pearson correlation	(E. Gobbi et al., 2010)
Zygosaccharomyces spoilage in apple juice	MOX (PEN3)	10	Sensory panel	LDA, PLS	(Wang et al., 2016)
Apple defects	CP (Cyranose 320)	32	-	PCA, MANOVA, DA	(Pathange, Mallikarjunan, Marini, O'Keefe, & Vaughan, 2006)
	CP	32	Z-nose	PCA, PNN, Bayesian	(Li, Heinemann, & Sherry,

	(Cyanose 320)				2007)
Medicinal off-flavor in apple juice	MOX (PEN3)	10	GC-MS, Test panel	PCA, LDA, ANOVA	(Huang, Guo, Yuan, Luo, & Yue, 2015)
Spoilage of milk and fish	SAW	6	-	Fuzzy c-means, PCA, RBNN	(Verma & Yadava, 2015)
Milk spoilage (bacteria and yeasts)	CP (BH-114)	14	-	DFA, PCA, Dendrogram, NN	(Naresh Magan, Pavlou, & Chrysanthakis, 2001)
Olive oil defects	MOX (EOS)	6	GC-MS, Test panel	PCA, SIMCA	(Esposito et al., 2006)
	MOX (EOS507)	6	Test panel	LDA, MLR, NN	(Lerma-García et al., 2010)
Rancidity of oil	MOX (EOS507)	18	Rancidity analysis	PCA, HCPC	(Upadhyay, Sehwaq, & Mishra, 2017)
Classification of Chicken meat freshness and bacterial population prediction	MOX	8	GC-MS	BPNN	Timsorn et al., 2016
Prediction of total volatile basic nitrogen (TVB-N) content in chicken meat	Colorimetric sensors array	-	Hyperspectral imaging system, Texture analysis	Data fusion techniques	Khulal et al., (2017)
Microbiological examination of beef fillets	QMB	8	Microbiological and sensory analyses	SVM, DFA	Papadopoulou et al., 2013
Identification of spoiled beef	CP	32	Microbiological analysis	ANNs	Panigrahi et al., 2006a
Determining the spoilage of vacuum packaged beef	MOSFET	10	Microbiological and sensory analyses	PLSR	Blixt & Borch, 1999
Spoilage classification of beef	MOX (M-	9	Microbiological analyses	LDA, QDA	Panigrahi et al., 2006b

	Module E-nose)				
Monitoring the spoilage of beef fillets under storage	QCM	8	Microbiological analyses	Fuzzy-Wavelet Network	Kodogiannis, 2017
Odor spoilage sensing of beef and fish	MOS	8	-	SVM, ANNs	ul Hasan et al., (2012)
Developing an automated ranking platform to predict minced beef spoilage	QMB (LibraNose)	8	HPLC, FT-IR, GC-MS and MSI	OLS-R, SL-R, PCR, PLS-R, SVM-R, RF-R and kNN-R	Estelles-Lopez et al., 2017
Spoilage detecting in hairtail fish and pork	MOX	8	Measuring total volatile basic nitrogen (TVBN)	PCA	Tian et al., 2012
Spoilage Classification of Red Meat	MOS	6	Microbiological analyses	PLS, SVM	El Barbri et al., 2008
Detection of Acetone and Ethanol in spoiled meat	MOS (TGS822)	1	Microbiological analysis	Statistical analysis	Benabdellah et al., 2017
Reduction of <i>Salmonella</i> and the spoilage bacteria on fresh chilled pork	MOS (PEN3)	10	Chemical analyses	One-way ANOVA	Wang et al., 2017
Study of lipid oxidation of Chinese-style sausage	MOS (PEN3)	10	Measuring acid value (AV) and peroxide value (POV)	PLSDA, FLDA, MLR, ANNs, SVM, HCA	Gu et al., 2017
Identification of pork meat samples spoiled by <i>R. aquatilis</i>	Heracles II	Columns: MXT-5 and MXT-17	PCR and microbiological analyses	ANOVA, Tukey's post-hoc test	Godziszewska et al., 2017
Spoilage detection of modified	MOSFET,	10	Microbiological and	PLSR, ANNs	Rajamaki et al., 2006

atmosphere packaged poultry meat	NST 3320 instrument		sensory analyses		
Evaluation of Spoilage of the blue crab (Crab ( <i>Callinectes sapidus</i> ) meat	CP (Cyranose) <sup>TM</sup>	32	Microbiological and sensory analyses	Canonical discriminant analysis (CDA), stepwise discriminant analysis (SDA)	Sarnoski et al., 2008
Quality and spoilage identification in smoked salmon	MOX - FishNose system	6	GC-MS	Partial least-squares regression (PLSR)	Haugen et al., 2006

**Table 1.** Reports on spoilage microorganisms in selected food products as influenced by intrinsic and extrinsic factors

Food product	Extrinsic		Intrinsic				Preservative	Spoilage organism(s)	Reference
	Temperature		Atmospheric conditions		pH				
	Low	High	Aerobic	Anaerobic	Low	High			
Baked products		x	x			(x)		<i>Bacillus</i> spp. Moulds	Valerio et al., (2012); Vytrasova et al., 2002
Meat	x			x		x		Lactic acid bacteria, Enterobacteriaceae, <i>Clostridium</i> , <i>Shewanella</i>	Cavill et al., 2011; Doulgeracki et al., (2010); Hernandez- Macedo et al., 2012;
Meat	x		x			x		<i>Pseudomonas</i> , <i>Brochothrix</i> <i>thermosphacta</i> , <i>Photobacterium</i> ,	Ercolini et al., 2006; Nychas et al., 2008; Pennachia et al., 2011
Meat		x	x			x		Enterobacteriaceae, <i>Pseudomonas</i> , <i>Acinetobacter</i>	Gill and Newton, 1979
Meat	x			x		x	Nisin	Enterobacteriaceae, <i>Pseudomonas</i>	Ferrocino et al., 2013
Marinated broiler	x			x		x	Spices	<i>Leuconostoc</i> <i>gasicomaticum</i>	Susuiluito et al., (2003)
Raw milk (refrigeration)	x		x			Neutral		<i>Pseudomonas</i> , <i>Lactococcus</i> , <i>Acinetobacter</i>	von Neubeck et al., (2015)
Minimally processed vegetable	x		x			(x)		<i>Pseudomonas</i> , Enterobacteriaceae, <i>Cryptococcus</i>	Ragaert et al., (2007)

**Table 1 (contd.).** Reports on spoilage microorganisms in selected food products as influenced by intrinsic and extrinsic factors

Food product	Extrinsic		Intrinsic				Preservative	Spoilage organism(s)	Reference
	Temperature		Atmospheric conditions		pH				
	Low	High	Aerobic	Anaerobic	Low	High			
Filtered milk	x		x		ND			<i>Acinetobacter</i> , <i>Chryseobacterium</i> , <i>Psychrobacter</i> , <i>Sphingomonas</i> , <i>Paenibacillus</i> , <i>Bacillus</i>	Schmidt et al., 2012
Fish	x			x			Essential oil	<i>Aeromonas</i> , <i>Lactococcus</i>	Zhang et al., 2017
Fish		x	x					<i>Pseudomonas</i> , H <sub>2</sub> S producing bacteria, Enterobacteriaceae	Parpalani et al., 2014
Fish	x			x				<i>Pseudomonas</i> , <i>Photobacterium</i> , <i>Lactococcus</i> , <i>Brocothrix</i> <i>thermosphacta</i>	Koutsoumanis et al., (2000); Mace et al., 2012
Smoked fish	x			x				Lactic acid bacteria, <i>Phospobacterium</i> , psychotrophic Enterobacteriaceae	Lovdal, 2015
Seafood		x	x					<i>Proteus</i> , <i>Vibrio</i>	Yang et al., 2017
Fruits		x	x					Yeasts	Gram et al., 2002
Fermented alcoholic beverages – sake and beer	x			x	x		Ethanol as by product of fermentation	<i>Lactobacillus</i> spp, <i>Pediococcus</i> spp., <i>Pectinatus</i> spp., <i>Megaspaera</i> spp.	Jespersen and Jackobsen, (1996); Suzuki (2011)

**Table 2.** Some spoilage substrates and metabolites typically found in spoiled food

Sensory characteristic	Spoilage compound	Spoilage substrate	Food product	Reference
Blown pack	CO <sub>2</sub>	sugars	vacuum packed meat	Hernandez-Macedo et al. (2012)
Ropiness/Slime	EPS	glucose	wine	Delarheche et al. (2004)
		starch	bread	Valerio et al. (2008)
		sugars	vacuum packed cooked meats	Korkeala et al. (1988)
<b>Off odours</b>				
Fruity	ethylhexanoate, ethyloctanoate, ethyldecnoate	glucose	air stored beef	Ercolini et al. 2010
	ethyl butanoate	ethanol	meat	La Stora et al. (2012)
	hexanal	lipids	fish	Leduc et al. (2012)
Pungent/alcoholic/fermented	3-methyl-1-butanol, 2-butanol, ethanol	sugars	fish	Miks-Krajnik et al. (2016); Parpalani et al. (2017)
	1-pentanol	sugars	RTE salads	Dias-Lula et al. (2017)
	acetic acid	glucose	fish	Mace et al. (2013)
Fishy	Trimethylamine	trimethylamine oxide	bell peppers	Pothakos et al. (2014)
Musty, mushroom	1-octen-3-ol	unsaturated fatty acids	seafood	Lopez-Caballero et al. (2001)
			baby spinach	Dias-Lula et al. (2017)
			fish	Leduc et al. (2012)
			rapeseed	Gancarz et al., 2017
Cheesy	Acetoin	glucose	fish	Miks-Krajnik et al. (2016)
	Butanoic acid	triglycerides/amino acids	meat	Ercolini et al. (2011)
	2,3-heptanedione		shrimps	Jaffres et al. (2011)
Sulphide off-odour	H <sub>2</sub> S	sulphur containing amino acids	fish	Fonnechbech Vogel et al. (2005)
	Dimethyl sulfoxide	sulphur containing amino acids	baby spinach	Dias-Lula et al. (2017)

<sup>a</sup>The combination of acrolein with polyphenols leads to the production of bitter compounds.

**Table 2 (contd.).** Some spoilage substrates and metabolites typically found in spoiled food

Sensory characteristic	Spoilage compound	Spoilage substrate	Food product	Reference
		sulphur containing amino acids	fish	Parpalani et al. (2017)
<b>Off flavours</b>				
Rancid	Volatile fatty acids	triglycerides	milk	Deeth
Bitter	acrolein <sup>a</sup>	protein glycerol	milk beer and wine	Cleto et al., (2012) Garai-Ibabe et al., (2008)

<sup>a</sup>The combination of acrolein with polyphenols leads to the production of bitter compounds.

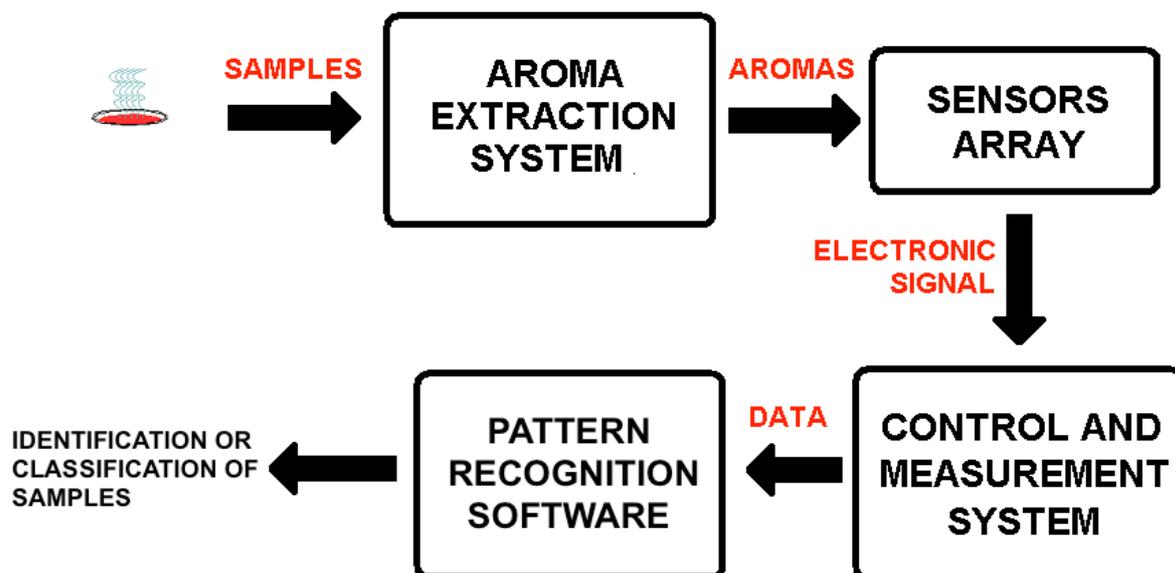
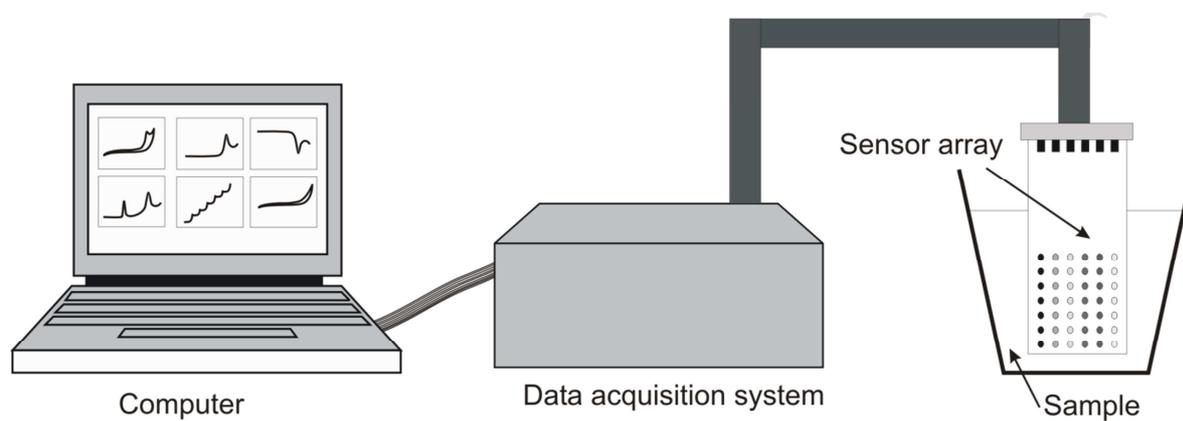
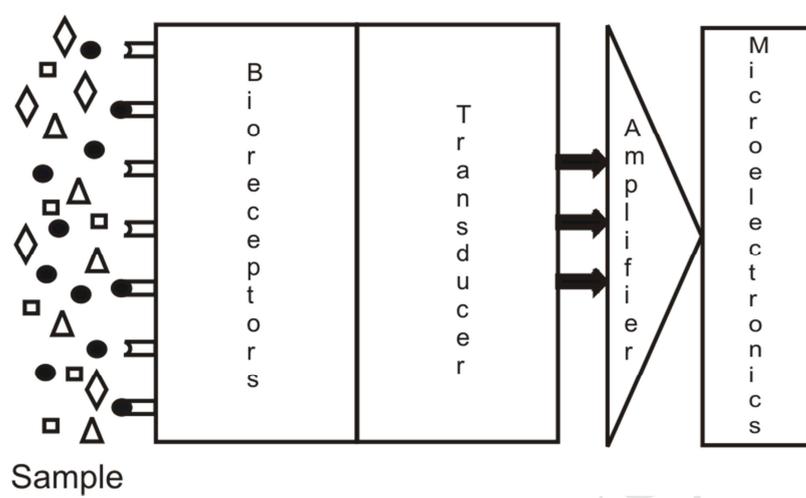


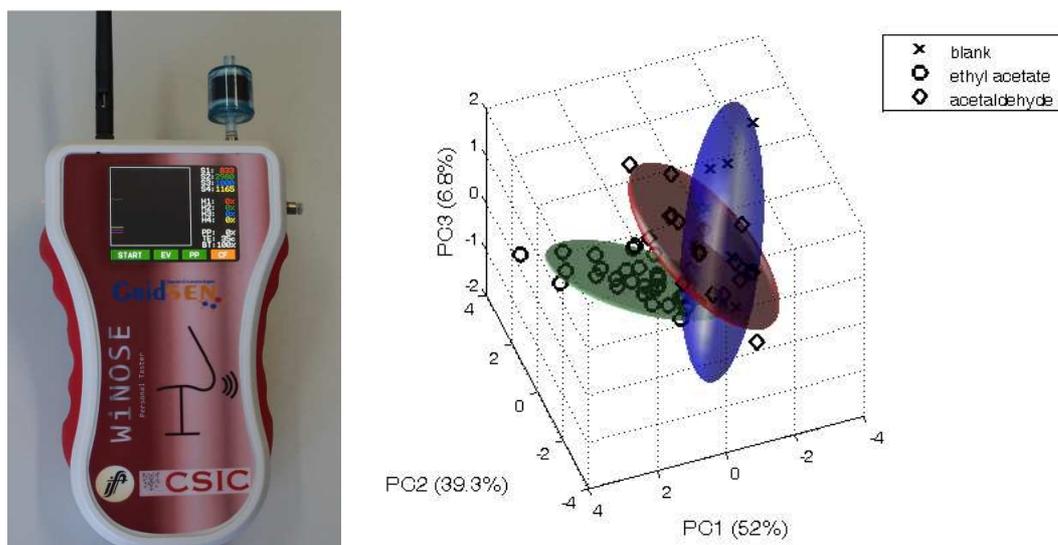
Fig. 1. Block diagram of an electronic nose system.



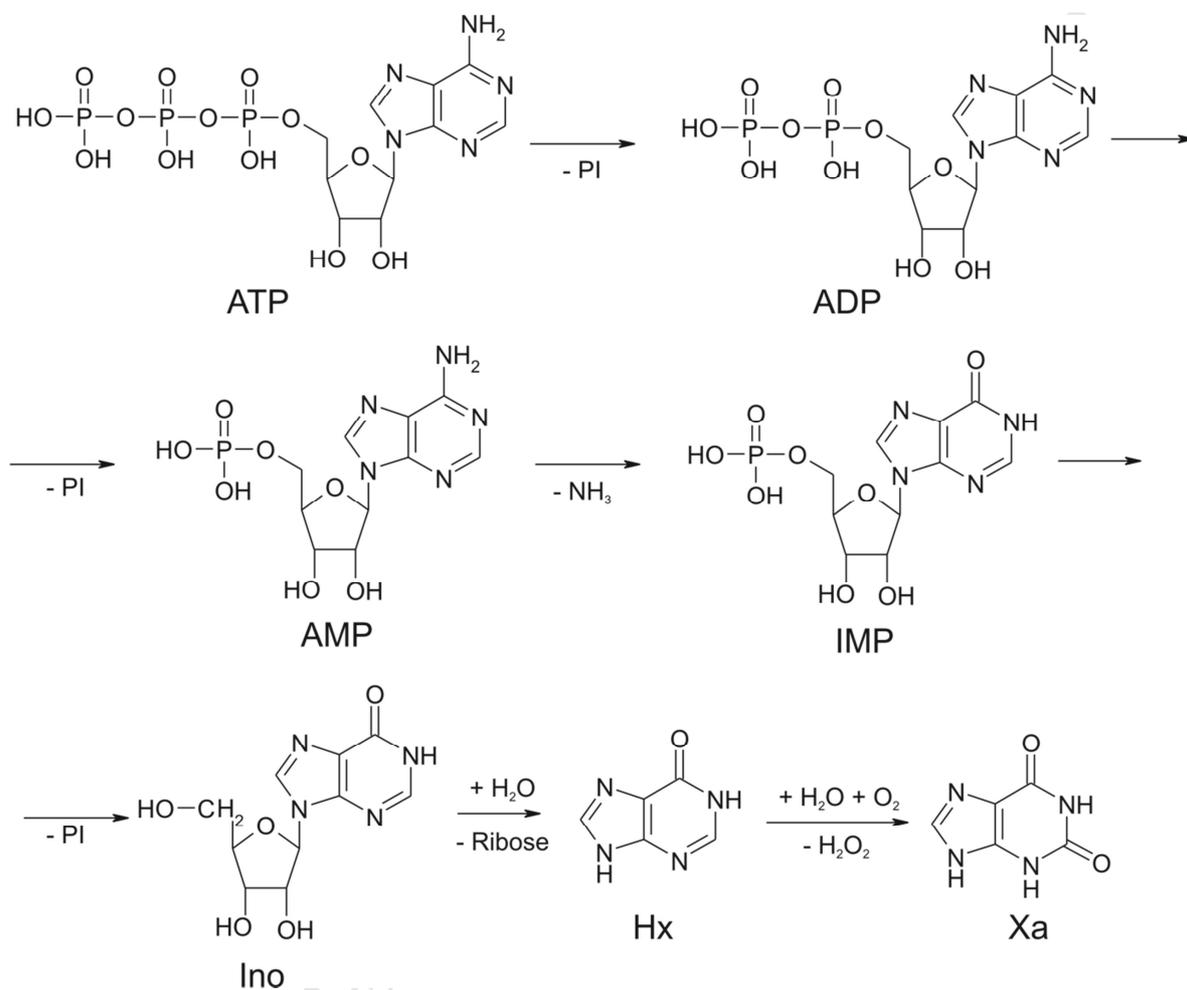
**Fig. 2.** General scheme of an electronic tongue system



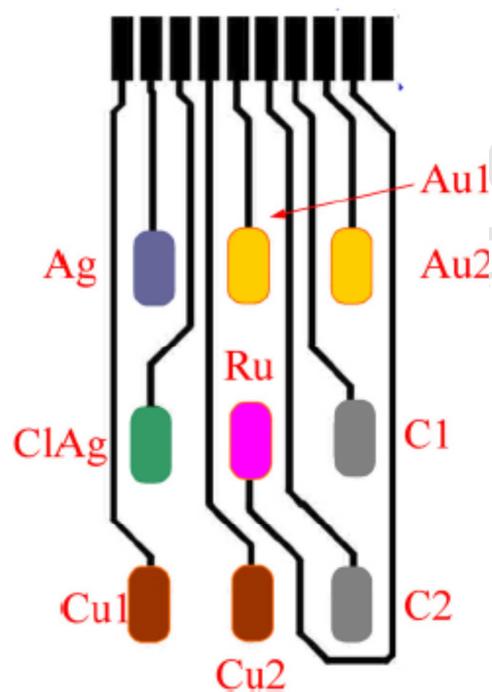
**Fig. 3.** Biosensor detection scheme



**Fig. 4.** Portable electronic nose system for the defect discrimination in beer and PCA score plot of measurements of beer defects.



**Fig. 5.** Decomposition of ATP in the muscles (Nelson & Cox, 2017)



**Fig. 6.** The sensor array used for the potentiometric electronic tongue (Gil-Sánchez, Soto, Martínez-Mañez, Garcia-Breijo, Ibáñez, & Llobet, 2011).

There is an urgent need for the development of rapid, reliable, precise and non-expensive systems to be used in the food supply and production chain.

In recent decades, some diagnostic tools such as electronic noses, electronic tongues and biosensors have attracted much interest for detection of food spoilage.

The future of the electronic tongue systems and the biosensors are closely related because improving the sensitivity and selectivity of the sensor array remain challenging tasks.

Electronic noses and gas sensors have shown in the last years an important enhancement in the time response and time life as well as a decrease in the size and consumption.